

AQT0032 - ABL1 Assay Validation

PhosphoSens®-Kinetic Assay Format

Outline for this Study



PhosphoSens-Kinetic Assay Validation

Enzyme Source, Construct, and Lot Information:

Carna ABL1 (Cat#/Lot#: 08-001/14CBS-0812R) amino acid full length; N-terminal His tag

Reference Compound Information:

Staurosporine MedChemExpress (Cat#/Lot#: HY-15141/125391) CAS No.: 62996-74-1

Experiments to be run:

Enzyme Titration

Sensor Peptide K_m Determination

ATP K_m Determination

DMSO Tolerance Test

Reference Compound IC_{50} Determination at ATP K_m

Enzyme Titration

Reaction Conditions and Set Up

AssayQuant®

Reaction Conditions:

54 mM HEPES, pH 7.5

1mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl₂

10 μM AQT0032

0.04, 0.06, 0.08, 0.1, 0.2, 0.4, 0.6, 0.8, 1, 2, 3, and 5 nM ABL1

Reaction Set Up:

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 or 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

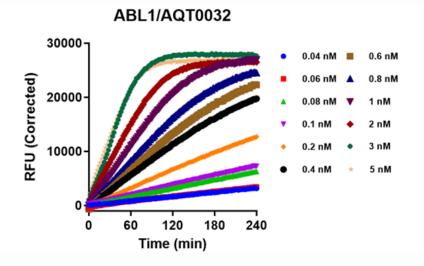
Notes:

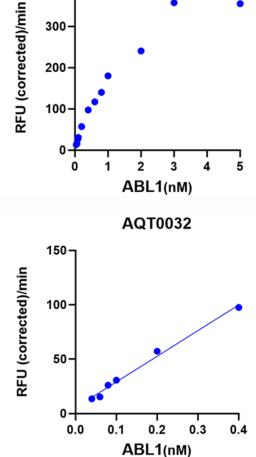
Enzyme Titration

AssayQuant®

Progress Curves

Complete **Progress** Curves



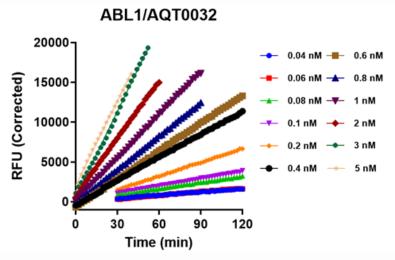


AQT0032

400-

200-

Linear Region of **Progress** Curves



Linear Range

Enzyme Titration



Reaction Rate Table

Furning Comp (nM)	Normalized	Normalized Rate	
Enzyme Conc. (nM)	Reaction Rate (RFU/pmole/min)	Stnd Error (RFU/pmole/min)	
0.04	16,875	284	
0.06	12,800	151	
0.08	16,300	136	
0.1	15,345	83	
0.2	14,325	56	
0.4	12,199	47	
0.6	9,750	16	
0.8	8,750	16	
1	9,000	22	
2	6,015	22	
3	5,958	62	
5	3,551	49	

The reaction is linear from 0.06 - 0.4 nM

Sensor Peptide K_m Determination



Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5

1mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl₂

1, 2, 3, 5, 7, 10, 15, 20, 30, 50, 70, and 100 μM AQT0032

2 nM ABL1

Reaction Set Up:

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

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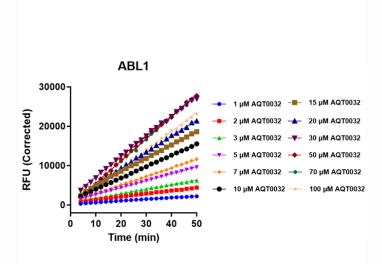
Notes:

Sensor Peptide K_m Determination

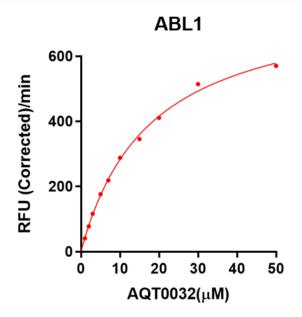


Titration Curves and K_m Plot and Table

Sensor Peptide Titration Curves



Sensor Peptide K_m Plot



Sensor Peptide K_m is 18 µM

Sensor Peptide K_m Table

Michaelis-Menten	
Best-fit values	
Vmax	784.4
Km	17.75
Std. Error	
Vmax	21.89
Km	1.075
95% CI (profile likelihood)	
Vmax	736.7 to 837.8
Km	15.46 to 20.43
Goodness of Fit	
Degrees of Freedom	8
R squared	0.9973

ATP K_m Determination



Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5

0, 2.0, 3.9, 7.8, 16, 31, 63, 125, 250, 500, 1000, and 2000 μM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl₂

10 μM AQT0032

1 nM ABL1

Reaction Set Up:

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

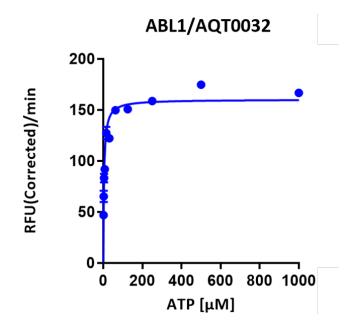
Notes:

ATP K_m Determination



Titration Curves and K_m Plot and Table





ATP K_m Table

Michaelis-Menten		
Best-fit values		
Vmax	160.0	
Km	3.876	
Std. Error		
Vmax	4.761	
Km	0.6881	
95% CI (asymptotic)		
Vmax	149.4 to 170.6	
Km	2.343 to 5.410	
Goodness of Fit		
Degrees of Freedom	10	
R squared	0.9354	

ATP K_m is 4 μM

DMSO Tolerance Test



Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5

1 mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl₂

0, .01, .02, .04, .08, .16, .31, .63, 1.3, 2.5, 5.0, and 10% DMSO

10 μM AQT0032

2 nM ABL1

Reaction Set Up:

2 or 2.5 μL 10x DMSO dilutions

14 or 17.5 μL Reaction Mix with Sensor Peptide, ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

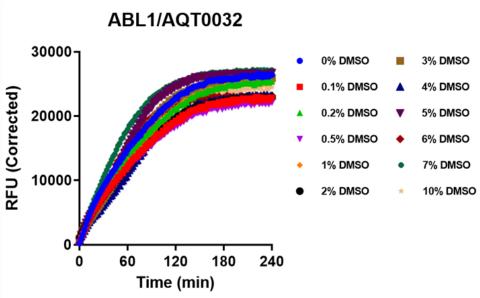
Notes:

DMSO Tolerance Test

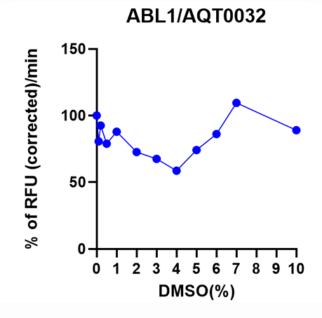


Titration Curves and Inhibition Plot





Reaction Rate vs [DMSO] Plot



No change in enzyme activity out to 1% DMSO

IC₅₀ Determination

AssayQuant®

Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5

ATP at K_m

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl₂

1% DMSO

10 μM AQT0032

2 nM Carna ABL1

0.1 mM Staurosporine was serially diluted (3-fold, 11-point) in 100% DMSO. The series was then diluted 10-fold into BSA (with a final concentration of 0.2 mg/mL BSA in 10% DMSO) to prepare the 10x compound stocks.

Reaction Set Up:

16 μL Reaction Mix with Sensor Peptide and Inhibitor

 $4 \mu L$ 1x EDB or Kinase dilutions (5x in EDB)

20 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Inhibitors are added via direct (0.4 μ L of 50X stock in 100% DMSO) or intermediate dilutions (2.0 μ L of 10X stock in 10% DMSO).

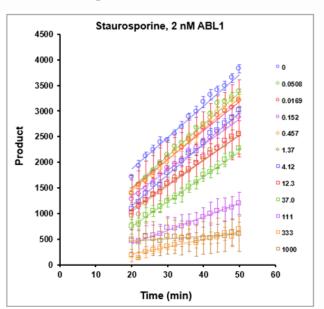
Notes:

IC₅₀ Determination

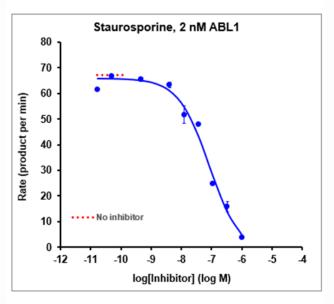


Progress Curves and IC₅₀ Curves and Table

Linear Region of Progress Curves



IC₅₀ Curve



IC₅₀ Table

Parameter	Value	
Bottom	-2.9	
Тор	65.8	
log IC50	-7.08	
IC50 (nM)	83.93	
Ki (nM)	41.97	
Slope	-0.850	
R squared	0.982	
IC50 approx SE (nM)	4.81	
50% inhibition (nM)	76.07	

The Y-axis label is RFU/min.

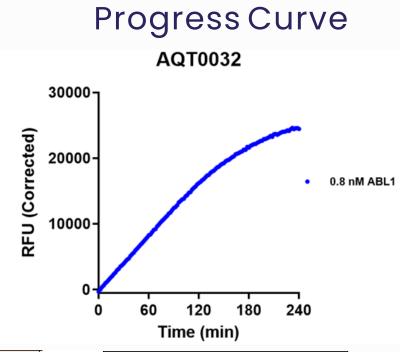
Staurosporine IC₅₀ Determination at ATP K_m is 84 nM

Summary



Assay Validation Results and Progress Curve and Assay Strength at 1 mM ATP

Experiment	Result		
Enzyme Titration Linear Range	0.06 - 0.4 nM		
Sensor Peptide K _m Value	18 μΜ		
ATP K _m Value	4 μΜ		
DMSO Tolerance (highest % recommended)	1		
Staurosporine IC ₅₀ Determination at ATP K _m	84 nM		



Kinase Name	Conc. (nM)	Sox-based Substrate Name	Normalized Reaction Rate (RFU/pmole/min)	Normalized Rate StndError (RFU/pmole/min)	Maximum Signal:Bkgd (S/B) Kinetic
ABL1	0.8	AQT0032	8,750	16	3.5

Under the conditions utilized for this experiment, the assay is Very Strong