

# AQT0150 - RIPK1 Assay Validation

PhosphoSens®-Kinetic Assay Format

1

# **Outline for this Study**



PhosphoSens-Kinetic Assay Validation

#### **Enzyme Source, Construct, and Lot Information:**

SignalChem RIPK1, (Cat & Lot #, R07-11G/O3960-16) amino acids 1-327, N-term GST-tag

#### **Reference Compound Information:**

Staurosporine

#### **Experiments to be run:**

**Enzyme Titration** 

Sensor Peptide K<sub>m</sub> Determination

ATP K<sub>m</sub> Determination

**DMSO Tolerance Test** 

Reference Compound IC<sub>50</sub> Determination at ATPK<sub>m</sub>

# **Enzyme Titration**

# AssayQuant®

### **Reaction Conditions and Set Up**

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5 1mM ATP 1.2 mM DTT 0.012% Brij-35 1% glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl<sub>2</sub> 20 μM AQT0150 0.03, 0.06, 0.12, 0.23, 0.47, 0.94, 1.88, 3.75, 7.5, 15, 30, and 60 nM RIPK1

# Reaction Set Up:<br/>2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 or 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

#### Notes:

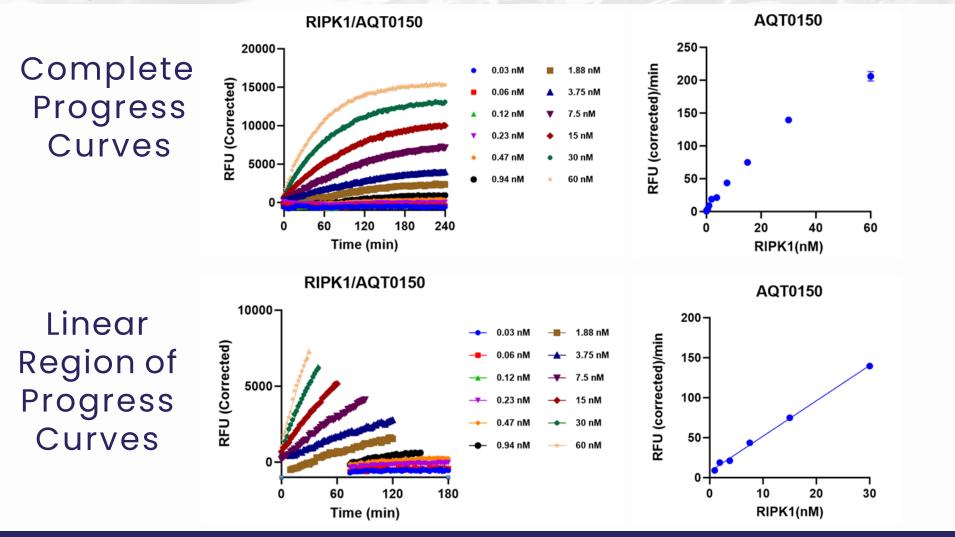
# **Enzyme Titration**



Linear

Range

#### **Progress Curves**



How Can We Help? For technical questions, please reach out at hello@assayquant.com

# **Enzyme Titration**

# AssayQuant<sup>®</sup>

## **Reaction Rate Table**

Enzyme Conc. (nM)	Normalized Reaction Rate (RFU/pmole/min)	Normalized Rate Stnd Error (RFU/pmole/min)	
0.03	1,190	325	
0.06	1,283	180	
0.12	869	66	
0.23	577	38	
0.47	414	21	
0.94	484	22	
1.88	504	10	
3.75	283	5	
7.5	291	4	
15	250	3	
30	233	6	
60	172	6	

## The reaction is linear from 3.75 - 30 nM

# Sensor Peptide K<sub>m</sub> Determination



**Reaction Conditions and Set Up** 

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5 1mM ATP 1.2 mM DTT 0.012% Brij-35 1% glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl<sub>2</sub> 1, 2, 3, 5, 7, 10, 15, 20, 30, 50, 70, and 100 μM AQT0150 20 nM RIPK1

# Reaction Set Up:<br/>2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

#### Notes:

# **Sensor Peptide K<sub>m</sub> Determination**

Titration Curves and K<sub>m</sub> Plot and Table

Sensor Peptide Titration Curves

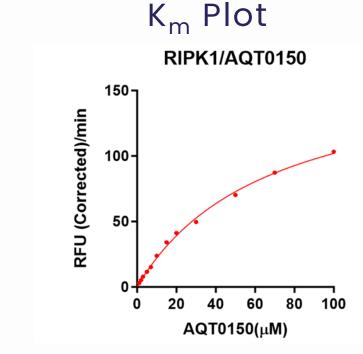
**RIPK1/AQT0150** 

60

Time (min)

8000-

RFU (Corrected) 00 00



Sensor Peptide

## Sensor Peptide K<sub>m</sub> Table

AssayQuant

Michaelis-Menten	
Best-fit values	
Vmax	170.5
Km	67.26
Std. Error	
Vmax	7.234
Km	5.196
95% CI (profile likelihood)	
Vmax	155.5 to 188.8
Km	56.69 to 80.68
Goodness of Fit	
Degrees of Freedom	10
R squared	0.9973

## Sensor Peptide K<sub>m</sub> is 67 µM

120

# **ATP K<sub>m</sub> Determination**



## **Reaction Conditions and Set Up**

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5
0, 2.0, 3.9, 7.8, 16, 31, 63, 125, 250, 500, 1000, and 2000 $\mu M$ ATP
1.2 mM DTT
0.012% Brij-35
1% glycerol
0.2 mg/ml BSA
0.55 mM EGTA
10 mM MgCl <sub>2</sub>
20 μM AQT0150
15 nM RIPK1

# Reaction Set Up:<br/>2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

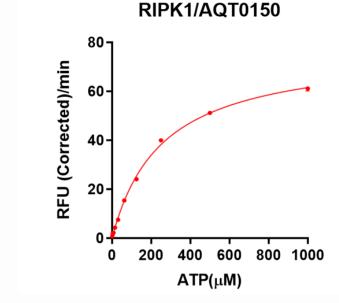
Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

#### Notes:

# **ATP K<sub>m</sub> Determination**

Titration Curves and K<sub>m</sub> Plot and Table

ATP Titration Curves



## ATP K<sub>m</sub> Table

Michaelis-Menten		
Best-fit values		
Vmax	77.22	
Km	255.5	
Std. Error		
Vmax	1.653	
Km	14.00	
95% CI (profile likelihood)		
Vmax	73.67 to 81.08	
Km	226.3 to 288.9	
Goodness of Fit		
Degrees of Freedom	9	
R squared	0.9985	

#### RIPK1/AQT0150

## ATP K<sub>m</sub> is 256 µM

# ATP K<sub>m</sub> Plot

#### How Can We Help? For technical questions, please reach out at hello@assayquant.com



# **DMSO Tolerance Test**



### **Reaction Conditions and Set Up**

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5
1 mM ATP
1.2 mM DTT
0.012% Brij-35
1% glycerol
0.2 mg/ml BSA
0.55 mM EGTA
10 mM MgCl<sub>2</sub>
0, .01, .02, .04, .08, .16, .31, .63, 1.3, 2.5, 5.0, and 10% DMSO
15 μM AQT0150
30 nM RIPK1

# Reaction Set Up:<br/>2 or 2.5 μL10x DMSO dilutions14 or 17.5 μLReaction Mix with Sensor Peptide, ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

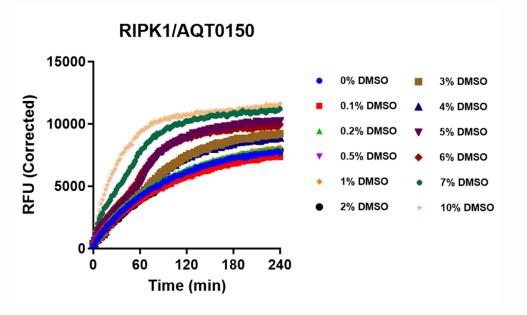
#### Notes:

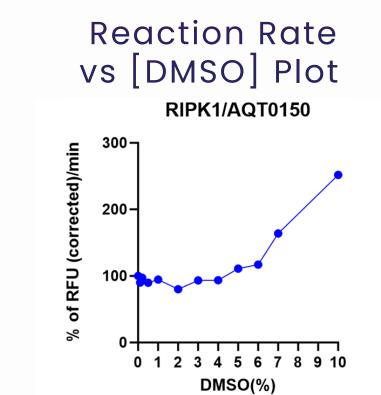
## **DMSO Tolerance Test**



**Titration Curves and Inhibition Plot** 

## Complete Progress Curves





## No change in enzyme activity out to 4% DMSO

# IC<sub>50</sub> Determination

### **Reaction Conditions and Set Up**

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5 ATP at K<sub>m</sub>

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

1% DMSO

 $20 \ \mu M \ AQT0150$ 

15 nM RIPK1

0.1 mM Staurosporine was serially diluted (3-fold, 11-point) in 100%DMSO. The series was then diluted 10-fold into BSA (with a final concentration of 0.2 mg/mL BSA in 10% DMSO) to prepare the 10x compound stocks.

#### **Reaction Set Up:**

- 16 μL Reaction Mix with Sensor Peptide and Inhibitor
- $4 \mu L$  1x EDB or Kinase dilutions (5x in EDB)
- 20 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Inhibitors are added via direct (0.4  $\mu$ L of 50X stock in 100% DMSO) or intermediate dilutions (2.0  $\mu$ L of 10X stock in 10% DMSO).

#### Notes:





# IC<sub>50</sub> Determination

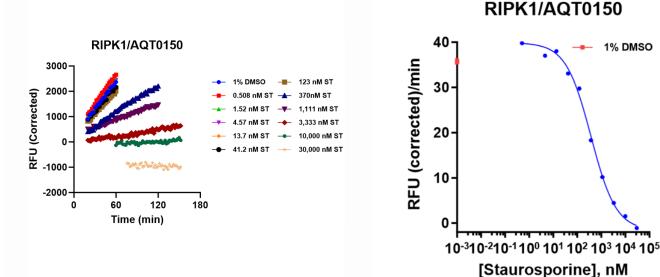


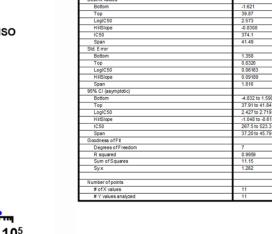
Progress Curves and IC<sub>50</sub> Curves and Table

## Linear Region of Progress Curves

## IC<sub>50</sub> Curve

IC<sub>50</sub> Table





The Y-axis label is RFU/min.

Staurosporine IC<sub>50</sub> Determination at ATP K<sub>m</sub> is 370 nM

# Summary



## Assay Validation Results and Progress Curve and Assay Strength at 1 mM ATP

Experiment				Resu	lt	Progress Curve	
Enzyme Titration Linear Range				3.75 - 30	) nM	RIPK1/AQT0150	
Sensor Peptide K <sub>m</sub> Value				67 μľ	M	() 10000-	
ATP K <sub>m</sub> Value				256 μΜ		DUBLE STORE	
DMSOTolerance (highest % recommended)			(1	4			
Staurosporine IC50 Determination at ATP Km				370 nM		0 60 120 180 240	
						Time (min)	
	Kinase Name	Conc. (nM)	Sox-based Substrate Name	Normalized Reaction Rate (RFU/pmole/mi	Normalized Rate StndError (RFU/pmole/mi	Assay Strength Key       Very Strong     > 1,000 (RFU/pmole/min)       Strong     300 to 999 (RFU/pmole/min)       Moderate     100 to 299 (RFU/pmole/min)	
	RIPK1	15	AQT0150	250	3	Weak 30 to 99 (RFU/pmole/min)	

## Under the conditions utilized for this experiment, the assay is Moderate