

# AQT0232 - RPS6KA1 (RSK1) Assay Validation

PhosphoSens®-Kinetic Assay Format

# Outline for this Study



### PhosphoSens-Kinetic Assay Validation

### **Enzyme Source, Construct, and Lot Information:**

Carna RSK1 (Cat#/Lot#: 01-149/12CBS-0749F) amino acids full length; N-terminal GST tag

### **Reference Compound Information:**

Staurosporine MedChemExpress (Cat#/Lot#: HY-15141/125391) CAS No.: 62996-74-1

### **Experiments to be run:**

**Enzyme Titration** 

Sensor Peptide K<sub>m</sub> Determination

ATP K<sub>m</sub> Determination

**DMSO Tolerance Test** 

Reference Compound  $IC_{50}$  Determination at ATP  $K_m$ 

## **Enzyme Titration**

# AssayQuant®

### Reaction Conditions and Set Up

### **Reaction Conditions:**

54 mM HEPES, pH 7.5

1mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

15 μM AQT0232

0.01, 0.02, 0.04, 0.08, 0.16, 0.3125, 0.625, 1.25, 2.5, 5, 10, and 20 nM RSK1

### Reaction Set Up:

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 or 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

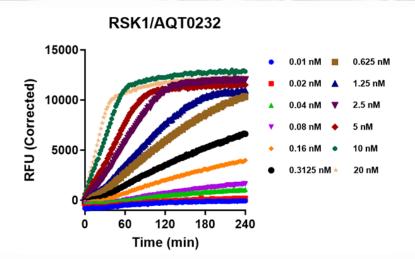
#### Notes:

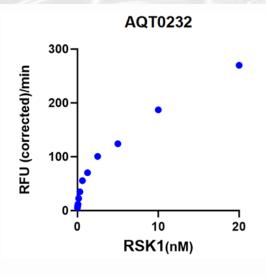
## **Enzyme Titration**



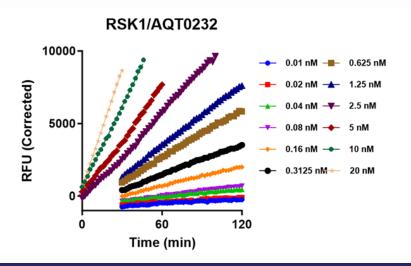
**Progress Curves** 

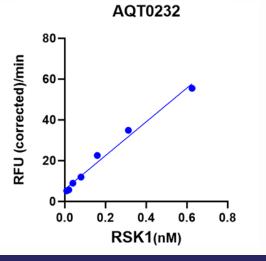
Complete Progress Curves





Linear Region of Progress Curves





Linear Range

# **Enzyme Titration**



### Reaction Rate Table

Enzyme Conc. (nM)	Normalized	Normalized Rate
Elizyille Colic. (Ilivi)	Reaction Rate (RFU/pmole/min)	Stnd Error (RFU/pmole/min)
0.01	26,315	970
0.02	14,320	433
0.04	11,171	186
0.08	7,488	95
0.16	7,047	56
0.3125	5,587	41
0.625	4,440	25
1.25	2,818	10
2.5	2,010	12
5	1,240	12
10	936	7
20	675	4

The reaction is linear from 0.08 - 0.625 nM

# Sensor Peptide K<sub>m</sub> Determination



### Reaction Conditions and Set Up

### **Reaction Conditions:**

54 mM HEPES, pH 7.5

1mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

1, 2, 3, 5, 7, 10, 15, 20, 30, 50, 70, and 100  $\mu$ M AQT0232

1 nM RSK1

### Reaction Set Up:

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

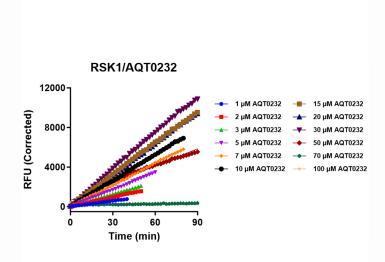
#### Notes:

# Sensor Peptide K<sub>m</sub> Determination

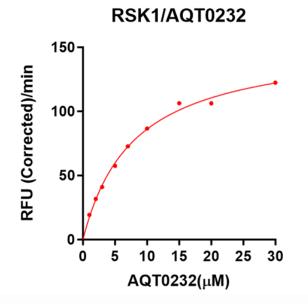


Titration Curves and K<sub>m</sub> Plot and Table

## Sensor Peptide Titration Curves



## Sensor Peptide K<sub>m</sub> Plot



Sensor Peptide K<sub>m</sub> is 7.9 µM

## Sensor Peptide K<sub>m</sub> Table

Michaelis-Menten	
Best-fit values	
Vmax	154.3
Km	7.921
Std. Error	
Vmax	4.822
Km	0.6236
95% CI (profile likelihood)	
Vmax	143.7 to 166.5
Km	6.585 to 9.547
Goodness of Fit	
Degrees of Freedom	7
R squared	0.9945

## ATP K<sub>m</sub> Determination



## Reaction Conditions and Set Up

### **Reaction Conditions:**

54 mM HEPES, pH 7.5

0, 2.0, 3.9, 7.8, 16, 31, 63, 125, 250, 500, 1000, and 2000 μM ATP

1.2 mM DTT

0.012% Brij-35

1%glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

15 μM AQT0232

1 nM RSK1

### **Reaction Set Up:**

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

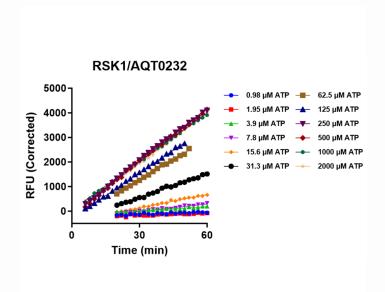
#### Notes:

# ATP K<sub>m</sub> Determination

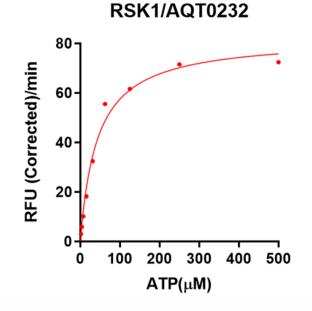


Titration Curves and K<sub>m</sub> Plot and Table

# ATP Titration Curves



## ATP K<sub>m</sub> Plot



ATP  $K_m$  is 43  $\mu M$ 

## ATP K<sub>m</sub> Table

Michaelis-Menten	
Best-fit values	
Vmax	82.32
Km	43.29
Std. Error	
Vmax	3.467
Km	6.206
95% CI (profile likelihood)	
Vmax	74.82 to 90.68
Km	31.44 to 59.54
Goodness of Fit	
Degrees of Freedom	7
R squared	0.9870

## **DMSO Tolerance Test**



### Reaction Conditions and Set Up

### **Reaction Conditions:**

54 mM HEPES, pH 7.5

1 mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

0, .01, .02, .04, .08, .16, .31, .63, 1.3, 2.5, 5.0, and 10% DMSO

15 μM AQT0232

1 nM RSK1

### **Reaction Set Up:**

2 or 2.5 μL 10x DMSO dilutions

14 or 17.5 μL Reaction Mix with Sensor Peptide, ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

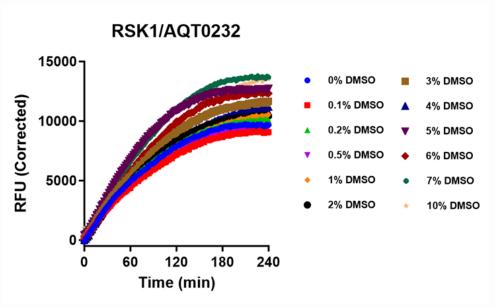
#### Notes:

## **DMSO Tolerance Test**

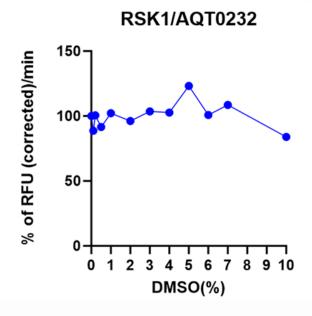


Titration Curves and Inhibition Plot

## Complete Progress Curves



# Reaction Rate vs [DMSO] Plot



No change in enzyme activity out to 4% DMSO

## IC<sub>50</sub> Determination

# Assay Quant®

### Reaction Conditions and Set Up

### **Reaction Conditions:**

54 mM HEPES, pH 7.5

ATP at K<sub>m</sub>

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

1% DMSO

15 μM AQT0232

1 nM RSK1

0.1 mM Staurosporine was serially diluted (3-fold, 11-point) in 100%DMSO. The series was then diluted 10-fold into BSA (with a final concentration of 0.2 mg/mL BSA in 10% DMSO) to prepare the 10x compound stocks.

### **Reaction Set Up:**

16 μL Reaction Mix with Sensor Peptide and Inhibitor

 $4 \mu L$  1x EDB or Kinase dilutions (5x in EDB)

20 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Inhibitors are added via direct (0.4  $\mu$ L of 50X stock in 100% DMSO) or intermediate dilutions (2.0  $\mu$ L of 10X stock in 10% DMSO).

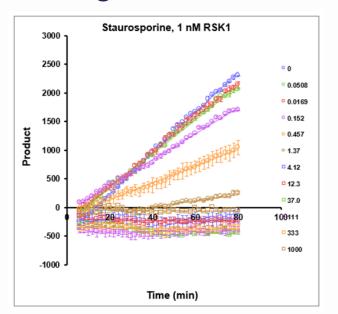
#### Notes:

# IC<sub>50</sub> Determination

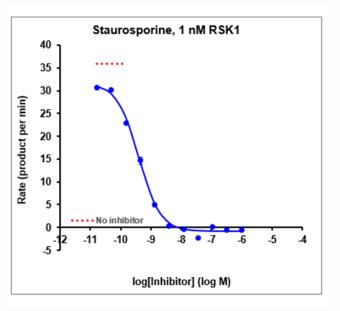


Progress Curves and IC<sub>50</sub> Curves and Table

# Linear Region of Progress Curves



## IC<sub>50</sub> Curve



IC<sub>50</sub> Table

Bottom -0	0.9
Top 3:	1.6
log IC50 -9	.40
IC50 (nM) 0.	39
Ki (nM) 0.	20
Slope -1.	225
R squared 0.9	996
IC50 approx SE (nM) 0.	.00
50% inhibition (nM) 0.	38

The Y-axis label is RFU/min.

Staurosporine IC<sub>50</sub> at ATP K<sub>m</sub> is 0.39 nM

## Summary

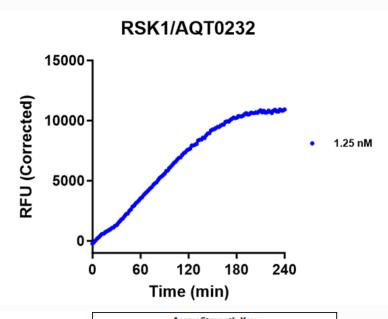


## Assay Validation Results and Progress Curve and Assay Strength at 1 mM ATP

Experiment	Result
Enzyme Titration Linear Range	0.08 - 0.625 nM
Sensor Peptide K <sub>m</sub> Value	7.9 μΜ
ATP K <sub>m</sub> Value	43 μΜ
DMSO Tolerance (highest % recommended)	4
Staurosporine IC <sub>50</sub> Determination at ATP K <sub>m</sub>	0.39 nM

Kinase Name	Conc. (nM)	Sox-based Substrate Name	Reaction Rate	Normalized Rate StndError (RFU/pmole/mi
			(III 0 / pillolo / IIII	(iii o pino io jini
RSK1	1.25	AQT0232	2,818	10

## **Progress Curve**



Assa	Assay Strength Key	
Very Strong	>1,000 (RFU/pmole/min)	
Strong	300 to 999 (RFU/pmole/min)	
Moderate	100 to 299 (RFU/pmole/min)	
Weak	30 to 99 (RFU/pmole/min)	

Under the conditions utilized for this experiment, the assay is Very Strong