

# AQT0232 - RPS6KA6 (RSK4) Assay Validation

PhosphoSens®-Kinetic Assay Format

# **Outline for this Study**



PhosphoSens-Kinetic Assay Validation

### **Enzyme Source, Construct, and Lot Information:**

Carna RSK4 (Cat#/Lot#: 01-152/07CBS-2401J) amino acids full length; N-terminal GST tag

#### **Reference Compound Information:**

Staurosporine MedChemExpress (Cat#/Lot#: HY-15141/125391) CAS No.: 62996-74-1

#### **Experiments to be run:**

**Enzyme Titration** 

Sensor Peptide K<sub>m</sub> Determination

ATP K<sub>m</sub> Determination

**DMSO Tolerance Test** 

Reference Compound IC<sub>50</sub> Determination at ATPK<sub>m</sub>

# **Enzyme Titration**

# AssayQuant TECHNOLOGIES INC.

## **Reaction Conditions and Set Up**

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5 1mM ATP 1.2 mM DTT 0.012% Brij-35 1% glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl<sub>2</sub>

 $15 \, \mu M \, AQT0232$ 

0.01, 0.02, 0.04, 0.08, 0.16, 0.3125, 0.625, 1.25, 2.5, 5, 10, and 20 nM RSK4

# Reaction Set Up:<br/>2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 or 25 µL final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 µL final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

#### Notes:

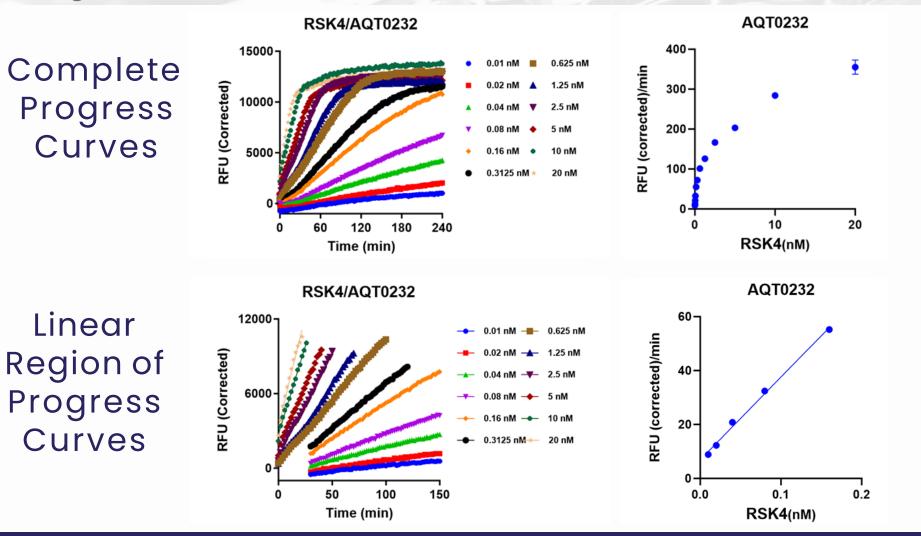
# **Enzyme Titration**

AssayQuant®

Linear

Range

### **Progress Curves**





# **Enzyme Titration**

# AssayQuant<sup>®</sup> TECHNOLOGIES INC.

## **Reaction Rate Table**

Enzyme Conc. (nM)	Normalized	Normalized Rate
Elizyille conc. (IIIVI)	Reaction Rate (RFU/pmole/min)	Stnd Error (RFU/pmole/min)
0.01	44,395	751
0.02	30,650	336
0.04	26,050	211
0.08	20,256	104
0.16	17,263	89
0.3125	11,554	51
0.625	8,104	36
1.25	5,032	53
2.5	3,328	35
5	2,032	28
10	1,421	32
20	888	44

## The reaction is linear from 0.02 - 0.16 nM

# Sensor Peptide K<sub>m</sub> Determination



**Reaction Conditions and Set Up** 

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5 1mM ATP 1.2 mM DTT 0.012% Brij-35 1% glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl<sub>2</sub> 1, 2, 3, 5, 7, 10, 15, 20, 30, 50, 70, and 100 μM AQT0232 0.3 nM RSK4

# Reaction Set Up:<br/>2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

#### Notes:

# **Sensor Peptide K<sub>m</sub> Determination**

Titration Curves and K<sub>m</sub> Plot and Table

Sensor Peptide Titration Curves

**RSK4/AQT0232** 

60

Time (min)

90

120

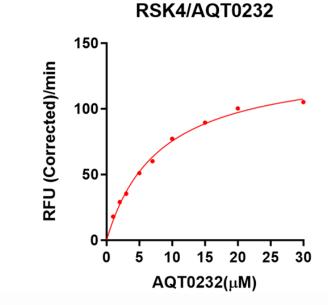
30

12000

8000

4000

RFU (Corrected)



Sensor Peptide

K<sub>m</sub> Plot

## Sensor Peptide K<sub>m</sub> Table

Michaelis-Menten	
Best-fit values	
Vmax	136.5
Km	8.090
Std. Error	
Vmax	4.349
Km	0.6446
95% CI (profile likelihood)	
Vmax	126.8 to 147.5
Km	6.704 to 9.780
Goodness of Fit	
Degrees of Freedom	7
R squared	0.9943

## Sensor Peptide K<sub>m</sub> is 8.1 µM



# **ATP K<sub>m</sub> Determination**



### **Reaction Conditions and Set Up**

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5		
0, 2.0, 3.9, 7.8, 16, 31, 63, 125, 250, 500, 1000, and 2000 $\mu M$ ATP		
1.2 mM DTT		
0.012% Brij-35		
1%glycerol		
0.2 mg/ml BSA		
0.55 mM EGTA		
10 mM MgCl <sub>2</sub>		
15 μM AQT0232		
0.3 nM RSK4		

# Reaction Set Up:<br/>2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

#### Notes:

# **ATP K<sub>m</sub> Determination**

Titration Curves and K<sub>m</sub> Plot and Table

2000 µM ATF

ATP Titration Curves

RSK4/AQT0232

30

0

60

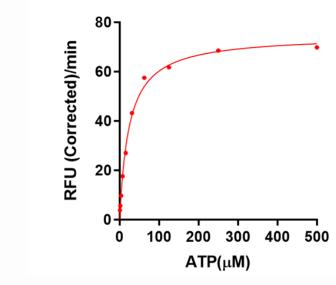
Time (min)

90

120

6000-

RFU (Corrected) 00 00



ATP K<sub>m</sub> Plot

**RSK4/AQT0232** 

## ATP K<sub>m</sub> Table

Michaelis-Menten	
Best-fit values	
Vmax	74.70
Km	24.07
Std. Error	
Vmax	1.354
Km	1.676
95% CI (profile likelihood)	
Vmax	71.67 to 77.84
Km	20.57 to 28.13
Goodness of Fit	
Degrees of Freedom	8
R squared	0.9963

## ATP $K_m$ is 24 $\mu M$

#### How Can We Help? For technical questions, please reach out at hello@assayquant.com



# **DMSO Tolerance Test**



### **Reaction Conditions and Set Up**

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5 1 mM ATP 1.2 mM DTT 0.012% Brij-35 1% glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl<sub>2</sub> 0, .01, .02, .04, .08, .16, .31, .63, 1.3, 2.5, 5.0, and 10% DMSO 15 μM AQT0232

0.3 nM RSK4

# Reaction Set Up:<br/>2 or 2.5 μL10x DMSO dilutions14 or 17.5 μLReaction Mix with Sensor Peptide, ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

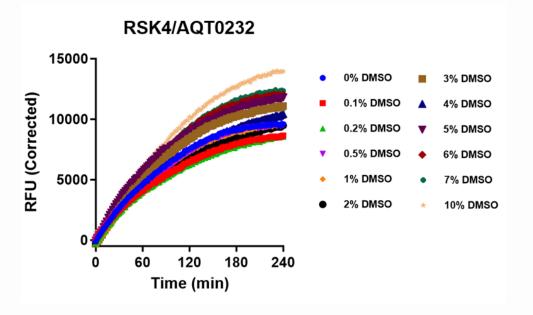
#### Notes:

# **DMSO Tolerance Test**

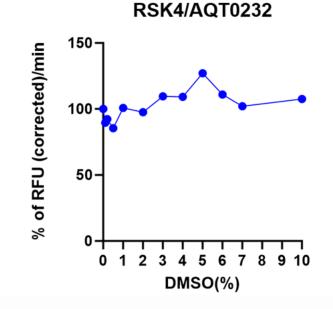


**Titration Curves and Inhibition Plot** 

## Complete Progress Curves



## Reaction Rate vs [DMSO] Plot



## No change in enzyme activity out to 2% DMSO

# IC<sub>50</sub> Determination

## **Reaction Conditions and Set Up**

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5 ATP at K<sub>m</sub> 1.2 mM DTT 0.012% Brij-35 1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

 $10 \,\mathrm{mM}\,\mathrm{MgCl}_2$ 

1% DMSO

15 µM AQT0232

0.3 nM RSK4

0.1 mM Staurosporine was serially diluted (3-fold, 11-point) in 100%DMSO. The series was then diluted 10-fold into BSA (with a final concentration of 0.2 mg/mL BSA in 10% DMSO) to prepare the 10x compound stocks.

#### **Reaction Set Up:**

- 16 μL Reaction Mix with Sensor Peptide and Inhibitor
- $4 \mu L$  1x EDB or Kinase dilutions (5x in EDB)
- 20 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Inhibitors are added via direct (0.4  $\mu$ L of 50X stock in 100% DMSO) or intermediate dilutions (2.0  $\mu$ L of 10X stock in 10% DMSO).

#### Notes:



# IC<sub>50</sub> Determination

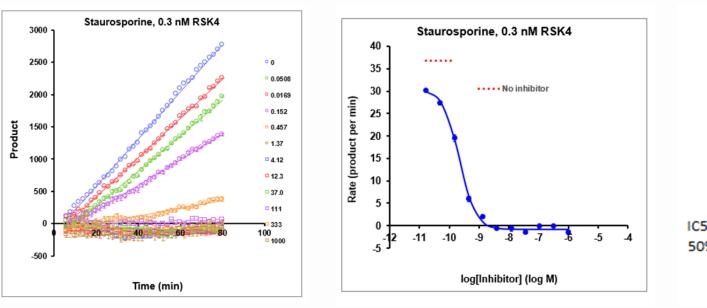


Progress Curves and IC<sub>50</sub> Curves and Table

## Linear Region of Progress Curves

IC<sub>50</sub> Curve





Parameter	Value
Bottom	-0.8
Тор	30.4
log IC50	-9.66
IC50 (nM)	0.22
Ki (nM)	0.11
Slope	-1.591
R squared	0.997
50 approx SE (nM)	0.00
% inhibition (nM)	0.21

The Y-axis label is RFU/min.

Staurosporine IC<sub>50</sub> at ATP K<sub>m</sub> is 0.22 nM

# Summary



## Assay Validation Results and Progress Curve and Assay Strength at 1 mM ATP

Experiment	Result	Progress Curve
Enzyme Titration Linear Range	0.02 - 0.16 nM	RSK4/AQT0232
Sensor Peptide K <sub>m</sub> Value	8.1 μM	ਿ ਉਹ 10000-
ATP K <sub>m</sub> Value	24 μM	• 0.3125 nM RSK4
DMSO Tolerance (highest % recommended)	2	000- 5000-
Staurosporine $\rm IC_{50}$ Determination at ATP $\rm K_m$	0.22 nM	0 60 120 180 240
		Time (min)
	Sox-based Normalized Normalized Rate	Assay Strength Key
Kinase Name Conc. (nM)	Strate Name (DELL/nmole/mi) (DELL/nmole/mi)	Very Strong > 1,000 (RFU/pmole/min)   Strong 300 to 999 (RFU/pmole/min)

(RFU/pmole/mi (RFU/pmole/mi

51

## Under the conditions utilized for this experiment, the assay is Very Strong

11,554

AQT0232

0.3125

RSK4

Moderate

Weak

100 to 299 (RFU/pmole/min)

30 to 99 (RFU/pmole/min)