

# AQT0235 - DDR2 Assay Validation

PhosphoSens®-Kinetic Assay Format

## Outline for this Study



#### PhosphoSens-Kinetic Assay Validation

#### **Enzyme Source, Construct, and Lot Information:**

Carna DDR2 (Cat/Lot #, 08-114/17CBS-0819H) amino acids 422-855(end), N-term GST tag

#### **Reference Compound Information:**

Staurosporine

#### **Experiments to be run:**

**Enzyme Titration** 

Sensor Peptide K<sub>m</sub> Determination

ATP K<sub>m</sub> Determination

**DMSO Tolerance Test** 

Reference Compound  $IC_{50}$  Determination at ATP  $K_m$ 

## **Enzyme Titration**

# AssayQuant®

#### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

1mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

15 μM AQT0235

0.01, 0.02, 0.04, 0.08, 0.16, 0.3125, 0.625, 1.25, 2.5, 5, 10 and 20 nM DDR2

#### Reaction Set Up:

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 or 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

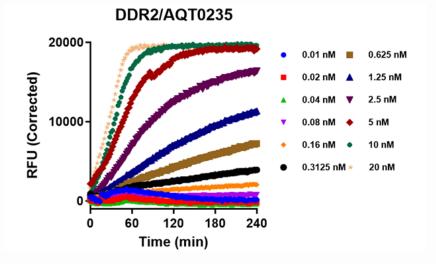
#### Notes:

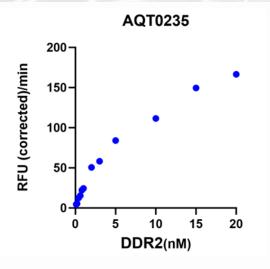
## **Enzyme Titration**



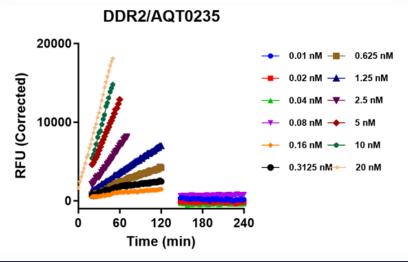
**Progress Curves** 

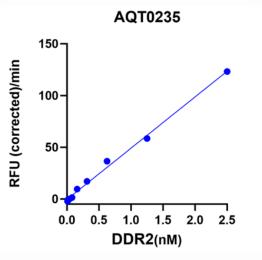
Complete Progress Curves





Linear Region of Progress Curves





Linear Range

## **Enzyme Titration**

# AssayQuant®

#### Reaction Rate Table

Enzyme Conc. (nM)	Normalized Reaction Rate (RFU/pmole/min)	Normalized Rate Stnd Error (RFU/pmole/min)
0.01	0	0
0.02	0	0
0.04	76	693
80.0	1,700	275
0.16	5,956	238
0.3125	5,309	198
0.625	5,848	106
1.25	4,674	22
2.5	4,928	57
5	4,172	33
10	3,227	44
20	1,683	16

The reaction is linear from 0.31 - 5.0 nM

## Sensor Peptide K<sub>m</sub> Determination



#### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

1mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

1, 2, 3, 5, 7, 10, 15, 20, 30, 50, 70, & 100 μM AQT0235

2 nM DDR2

#### Reaction Set Up:

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

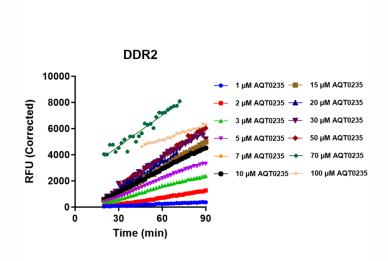
#### Notes:

## Sensor Peptide K<sub>m</sub> Determination

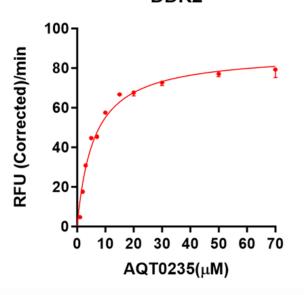


Titration Curves and K<sub>m</sub> Plot and Table

#### Sensor Peptide Titration Curves



## Sensor Peptide K<sub>m</sub> Plot



Sensor Peptide K<sub>m</sub> is 5.9 µM

#### Sensor Peptide K<sub>m</sub> Table

Michaelis-Menten	
Best-fit values	
Vmax	87.67
Km	5.925
Std. Error	
Vmax	3.164
Km	0.7362
95% CI (profile likelihood)	
Vmax	81.04 to 94.97
Km	4.515 to 7.727
Goodness of Fit	
Degrees of Freedom	9
R squared	0.9784

## ATP K<sub>m</sub> Determination



#### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

0, 2.0, 3.9, 7.8, 16, 31, 63, 125, 250, 500, 1000, and 2000 μM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

15 μM AQT0235

10 nM DDR2

#### **Reaction Set Up:**

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

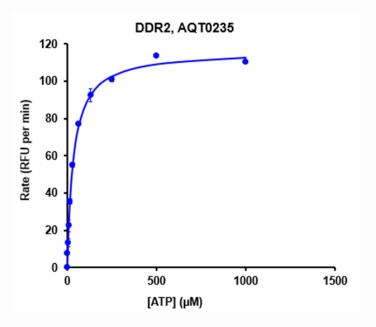
#### Notes:

## ATP K<sub>m</sub> Determination



Titration Curves and K<sub>m</sub> Plot and Table

#### ATP K<sub>m</sub> Plot



ATP  $K_m$  is 34  $\mu M$ 

### **DMSO Tolerance Test**



#### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

1 mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

0, .01, .02, .04, .08, .16, .31, .63, 1.3, 2.5, 5.0, and 10% DMSO

15 μM AQT0235

2 nM DDR2

#### Reaction Set Up:

2 or 2.5 μL 10x DMSO dilutions

14 or 17.5 μL Reaction Mix with Sensor Peptide, ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

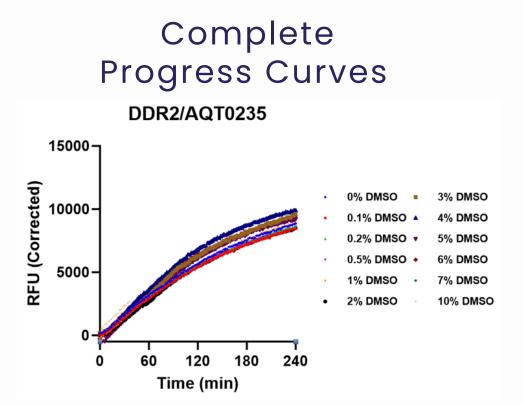
Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

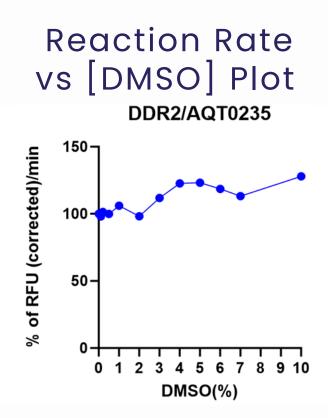
#### Notes:

## **DMSO Tolerance Test**



Titration Curves and Inhibition Plot





No change in enzyme activity out to 2% DMSO

## IC<sub>50</sub> Determination

# Assay Quant®

#### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

ATP at K<sub>m</sub>

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

1% DMSO

15 μM AQT0235

10 nM DDR2

0.1 mM Staurosporine with 3-fold titration in 100% DMSO then diluted 10-fold into BSA (with a final concentration of 0.2 mg/ml) for a DMSO concentration of 10% before diluted 10-fold into reaction mixture with a final DMSO concentration of 1%

#### **Reaction Set Up:**

16 μL Reaction Mix with Sensor Peptide and Inhibitor

 $4 \mu L$  1x EDB or Kinase dilutions (5x in EDB)

20 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Inhibitors are added via direct (0.4  $\mu$ L of 50X stock in 100% DMSO) or intermediate dilutions (2.0  $\mu$ L of 10X stock in 10% DMSO).

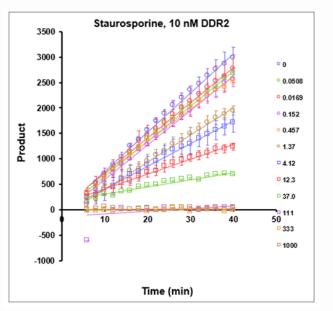
#### Notes:

## IC<sub>50</sub> Determination

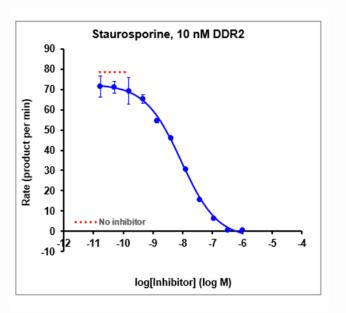


Progress Curves and IC<sub>50</sub> Curves and Table

## Linear Region of Progress Curves



IC<sub>50</sub> Curve



IC<sub>50</sub> Table

Parameter	Value
Bottom	-3.7
Top	72.7
log IC50	-8.06
IC50 (nM)	8.78
Ki (nM)	4.39
Slope	-0.720
R squared	0.998
IC50 approx SE (nM)	0.37
50% inhibition (nM)	7.66

The Y-axis label is RFU/min.

Staurosporine IC<sub>50</sub> Determination at ATP K<sub>m</sub> is 8.8 nM

## Summary



#### Assay Validation Results and Progress Curve and Assay Strength at 1 mM ATP

Experiment	Result	
Enzyme Titration Linear Range	0.31 - 5.0 nM	
Sensor Peptide K <sub>m</sub> Value	5.9 μΜ	
ATP K <sub>m</sub> Value	34 μΜ	
DMSO Tolerance (highest % recommended)	2	
Staurosporine IC50 Determination at ATP Km	8.8 nM	

Kinase Name	Conc. (nM)	Sox-based Substrate Name	Normalized Reaction Rate (RFU/pmole/min)	Normalized Rate StndError (RFU/pmole/min)
DDR2	2.5	AQT0235	4,928	57

# AQT0235 AQT0235 15000 10000 5000 10000 Time (min)

Assa	Assay Strength Key  Very Strong >1,000 (RFU/pmole/min)	
Very Strong		
Strong	300 to 999 (RFU/pmole/min)	
Moderate	100 to 299 (RFU/pmole/min)	
Weak	30 to 99 (RFU/pmole/min)	

Under the conditions utilized for this experiment, the assay is Very Strong