

AQT0258 - CDK16/CycY Assay Validation

PhosphoSens®-Kinetic Assay Format

Outline for this Study



PhosphoSens-Kinetic Assay Validation

Enzyme Source, Construct, and Lot Information:

Carna CDK16/CycY (Cat/Lot #, 04-116/13CBS-0287G) amino acids 107-496(end), N-term GST tag & full length GST-Cyclin Y

Reference Compound Information:

Staurosporine MedChemExpress (Cat#/Lot#: HY-15141/125391) CAS No.: 62996-74-1

Experiments to be run:

Enzyme Titration

Sensor Peptide K_m Determination

ATP K_m Determination

DMSO Tolerance Test

Reference Compound IC₅₀ Determination at ATP K_m

Enzyme Titration

AssayQuant®

Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5

1mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl₂

15 μM AQT0258

0.02, 0.04, 0.08, 0.16, 0.3125, 0.625, 1.25, 2.5, 5, 10, 20, and 40 nM Carna CDK16/CycY

Reaction Set Up:

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

 $4 \text{ or } 5 \mu L$ 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 or 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

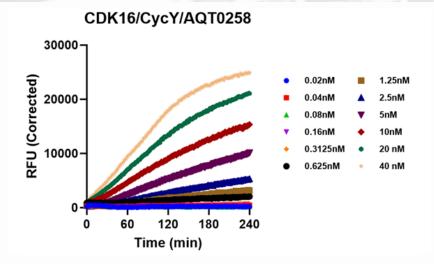
Notes:

Enzyme Titration

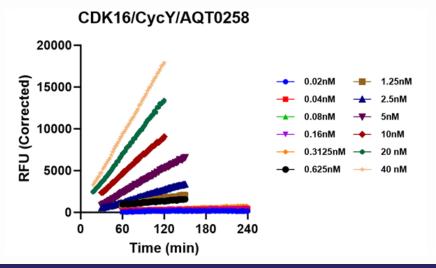


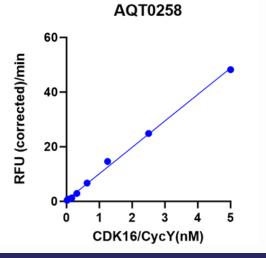
Progress Curves

Complete Progress Curves



Linear Region of Progress Curves





Linear Range

Enzyme Titration



Reaction Rate Table

Enzyme Conc. (nM)	Normalized Reaction Rate (RFU/pmole/min)	Normalized Rate Stnd Error (RFU/pmole/min)
0.02	1,118	203
0.04	940	80
0.08	1,071	104
0.16	800	41
0.3125	918	21
0.625	1,073	27
1.25	1,170	17
2.5	995	6
5	964	5
10	752	4
20	551	2
40	363	1

The reaction is linear from 0.04-5 nM

Sensor Peptide K_m Determination



Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5

1mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl₂

1, 2, 3, 5, 7, 10, 15, 20, 30, 50, 70, & 100 μM AQT0258

10 nM Carna CDK16/CycY

Reaction Set Up:

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

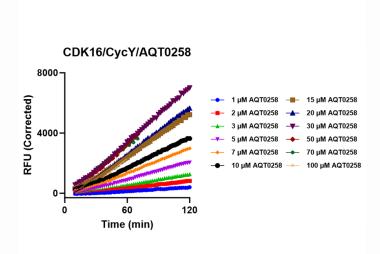
Notes:

Sensor Peptide K_m Determination



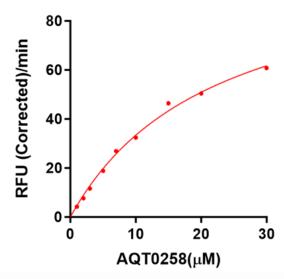
Titration Curves and K_m Plot and Table

Sensor Peptide Titration Curves



Sensor Peptide K_m Plot

CDK16/CycY/AQT0258



Sensor Peptide K_m is 22 μM

Sensor Peptide K_m Table

Michaelis-Menten	
Best-fit values	
Vmax	107.2
Km	22.16
Std. Error	
Vmax	7.155
Km	2.603
95% CI (profile likelihood)	
Vmax	92.97 to 126.9
Km	17.09 to 29.44
Goodness of Fit	
Degrees of Freedom	7
R squared	0.9948

ATP K_m Determination



Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5

0, 2.0, 3.9, 7.8, 16, 31, 63, 125, 250, 500, 1000, and 2000 μM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl₂

15 μM AQT0258

10 nM Carna CDK16/CycY

Reaction Set Up:

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

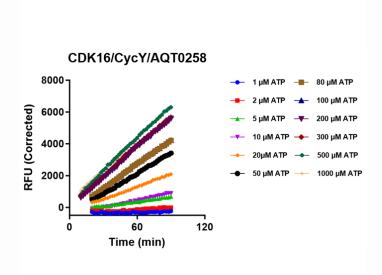
Notes:

ATP K_m Determination



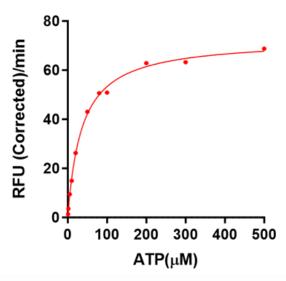
Titration Curves and K_m Plot and Table

ATP Titration Curves



ATP K_m Plot





ATP K_m is 36 μM

ATP K_m Table

Michaelis-Menten	
Best-fit values	
Vmax	72.55
Km	36.22
Std. Error	
Vmax	1.060
Km	2.019
95% CI (profile likelihood)	
Vmax	70.20 to 75.01
Km	31.90 to 41.09
Goodness of Fit	
Degrees of Freedom	9
R squared	0.9978

DMSO Tolerance Test



Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5

1 mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl₂

0, .01, .02, .04, .08, .16, .31, .63, 1.3, 2.5, 5.0, and 10% DMSO

15 μM AQT0258

10 nM Carna CDK16/CycY

Reaction Set Up:

2 or 2.5 μL 10x DMSO dilutions

14 or 17.5 µL Reaction Mix with Sensor Peptide, ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

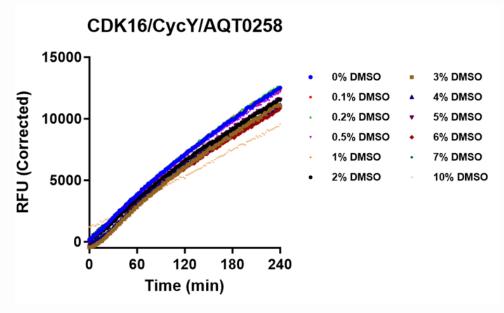
Notes:

DMSO Tolerance Test



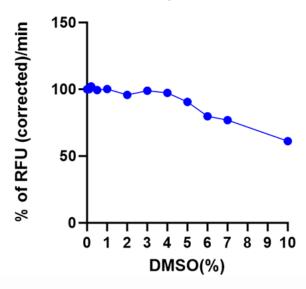
Titration Curves and Inhibition Plot

Complete Progress Curves



Reaction Rate vs [DMSO] Plot

CDK16/CycY/AQT0258



No change in enzyme activity out to 3% DMSO

IC₅₀ Determination

Assay Quant®

Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5

ATP at K_m

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl₂

1% DMSO

15 μM AQT0258

10 nM Carna CDK16/CycY

0.1 mM Staurosporine was serially diluted (3-fold, 11-point) in 100% DMSO. The series was then diluted 10-fold into BSA (with a final concentration of 0.2 mg/mL BSA in 10% DMSO) to prepare the 10x compound stocks.

Reaction Set Up:

16 μL Reaction Mix with Sensor Peptide and Inhibitor

 $4 \mu L$ 1x EDB or Kinase dilutions (5x in EDB)

20 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Inhibitors are added via direct (0.4 μ L of 50X stock in 100% DMSO) or intermediate dilutions (2.0 μ L of 10X stock in 10% DMSO).

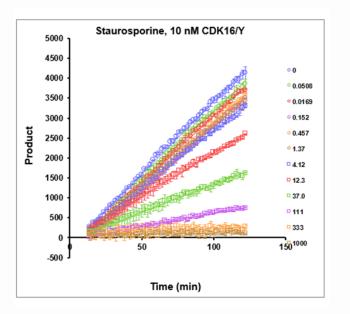
Notes:

IC₅₀ Determination

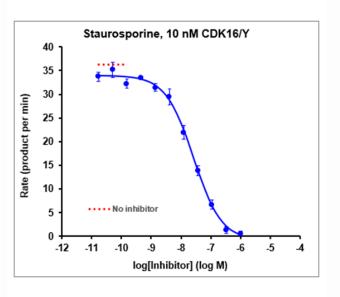


Progress Curves and IC₅₀ Curves and Table

Linear Region of Progress Curves



IC₅₀ Curve



IC₅₀ Table

Parameter	Value
Bottom	-1.0
Тор	33.9
log IC50	-7.58
IC50 (nM)	26.52
Ki (nM)	13.26
Slope	-0.931
R squared	0.997
IC50 approx SE (nM)	1.20
50% inhibition (nM)	24.92

The Y-axis label is RFU/min.

Staurosporine IC₅₀ Determination at ATP K_m is 27 nM

Summary



Assay Validation Results and Progress Curve and Assay Strength at 1 mM ATP

Experiment	Result
Enzyme Titration Linear Range	0.04 - 5 nM
Sensor Peptide K _m Value	22 μΜ
ATP K _m Value	36 μΜ
DMSO Tolerance (highest % recommended)	3
Staurosporine IC ₅₀ Determination at ATP K _m is	27 nM

Kinase Name	Conc. (nM)	Sox-based Substrate Name	Normalized Reaction Rate (RFU/pmole/min)	Normalized Rate StndError (RFU/pmole/min)
CDK16/CycY	10	AQT0258	752	4

Progress Curve CDK16/CycY/AQT0258 20000 15000 10000

Assa	Assay Strength Key	
Very Strong	Very Strong > 1,000 (RFU/pmole/min)	
Strong	300 to 999 (RFU/pmole/min)	
Moderate	100 to 299 (RFU/pmole/min)	
Weak	30 to 99 (RFU/pmole/min)	

Time (min)

Under the conditions utilized for this experiment, the assay is Strong