

# AQT0356 - MAP3K7 Assay Validation

PhosphoSens®-Kinetic Assay Format

## Outline for this Study



### PhosphoSens-Kinetic Assay Validation

### **Enzyme Source, Construct, and Lot Information:**

Carna BTN-TAK1-TAB1 (MAP3K7) (Cat#/Lot#: 09-419-21C/15CBS-0137B) amino acids 1-303 fused with TAB1 (437-504); C-terminal DYKDDDDK tag, biotinylated

### **Reference Compound Information:**

Staurosporine

### **Experiments to be run:**

**Enzyme Titration** 

Sensor Peptide K<sub>m</sub> Determination

ATP K<sub>m</sub> Determination

**DMSO Tolerance Test** 

Reference Compound  $IC_{50}$  Determination at ATP  $K_m$ 

## **Enzyme Titration**

# AssayQuant®

### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

1mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

15 μM AQT0356

0.02, 0.04, 0.08, 0.16, 0.3125, 0.625, 1.25, 2.5, 5, 10, 20, and 40 nM MAP3K7

### Reaction Set Up:

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 or 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

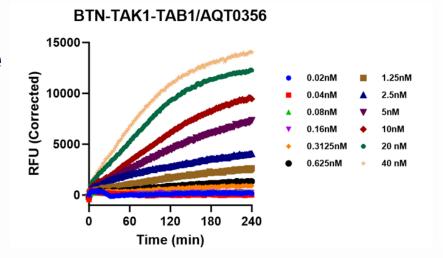
#### Notes:

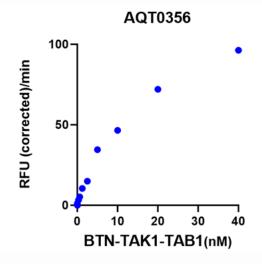
## **Enzyme Titration**



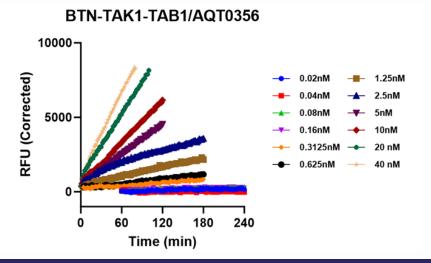
**Progress Curves** 

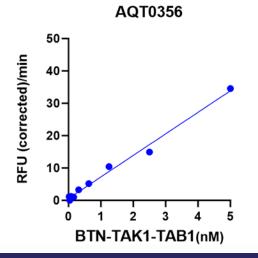
Complete Progress Curves





Linear Region of Progress Curves





Linear Range

## **Enzyme Titration**

# AssayQuant®

### Reaction Rate Table

Enzyme Conc. (nM)	Normalized Reaction Rate (RFU/pmole/min)	Normalized Rate Stnd Error (RFU/pmole/min)
0.02	2,863	198
0.04	6	85
0.08	756	36
0.16	320	21
0.3125	524	14
0.625	414	8
1.25	417	4
2.5	299	6
5	346	2
10	233	1
20	180	1
40	120	2

The reaction is linear from 0.16-5 nM

## Sensor Peptide K<sub>m</sub> Determination



### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

1mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

1, 2, 3, 5, 7, 10, 15, 20, 30, 50, 70, and 100 μM AQT0356

15 nM MAP3K7

### **Reaction Set Up:**

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

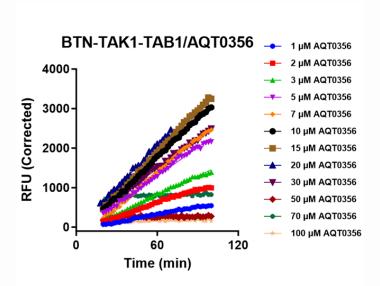
#### Notes:

## Sensor Peptide K<sub>m</sub> Determination

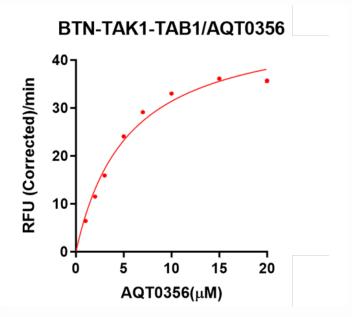


Titration Curves and K<sub>m</sub> Plot and Table

### Sensor Peptide Titration Curves



## Sensor Peptide K<sub>m</sub> Plot



Sensor Peptide K<sub>m</sub> is 5.4 µM

## Sensor Peptide K<sub>m</sub> Table

Michaelis-Menten	
Best-fit values	
Vmax	48.45
Km	5.440
Std. Error	
Vmax	2.938
Km	0.8544
95% CI (profile likelihood)	
Vmax	42.36 to 56.35
Km	3.787 to 7.860
Goodness of Fit	
Degrees of Freedom	6
R squared	0.9806

## ATP K<sub>m</sub> Determination



### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

0, 2.0, 3.9, 7.8, 16, 31, 63, 125, 250, 500, 1000, and 2000 μM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

15 μM AQT0356

10 nM MAP3K7

#### **Reaction Set Up:**

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

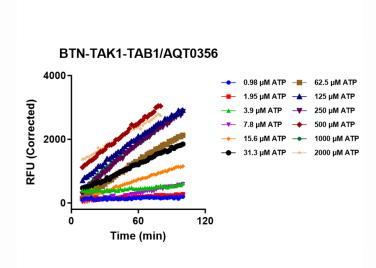
### Notes:

## ATP K<sub>m</sub> Determination



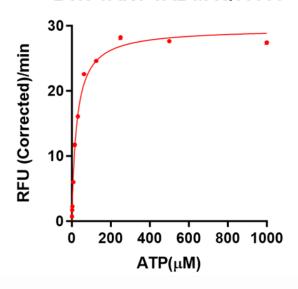
Titration Curves and K<sub>m</sub> Plot and Table

## ATP Titration Curves



### ATP K<sub>m</sub> Plot





ATP  $K_m$  is 25  $\mu M$ 

## ATP K<sub>m</sub> Table

Michaelis-Menten	
Best-fit values	
Vmax	29.55
Km	25.32
Std. Error	
Vmax	0.7154
Km	2.630
95% CI (profile likelihood)	
Vmax	28.00 to 31.16
Km	20.22 to 31.62
Goodness of Fit	
Degrees of Freedom	9
R squared	0.9914

## **DMSO Tolerance Test**



### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

1 mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

0, .01, .02, .04, .08, .16, .31, .63, 1.3, 2.5, 5.0, and 10% DMSO

15 μM AQT0356

15 nM MAP3K7

#### **Reaction Set Up:**

2 or 2.5 μL 10x DMSO dilutions

14 or 17.5 μL Reaction Mix with Sensor Peptide, ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

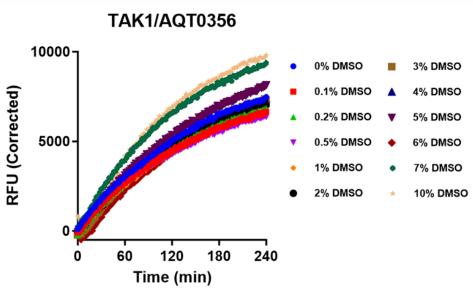
#### Notes:

## **DMSO Tolerance Test**

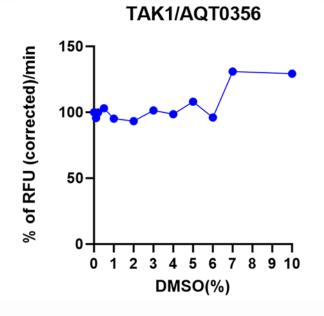


Titration Curves and Inhibition Plot





# Reaction Rate vs [DMSO] Plot



No change in enzyme activity out to 6% DMSO

## IC<sub>50</sub> Determination

# AssayQuant®

### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

ATP at K<sub>m</sub>

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

1% DMSO

15 μM AQT0356

15 nM MAP3K7

0.1 mM Staurosporine was serially diluted (3-fold, 11-point) in 100% DMSO. The series was then diluted 10-fold into BSA (with a final concentration of 0.2 mg/mL BSA in 10% DMSO) to prepare the 10x compound stocks.

#### **Reaction Set Up:**

16 μL Reaction Mix with Sensor Peptide and Inhibitor

 $4 \mu L$  1x EDB or Kinase dilutions (5x in EDB)

20 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Inhibitors are added via direct (0.4  $\mu$ L of 50X stock in 100% DMSO) or intermediate dilutions (2.0  $\mu$ L of 10X stock in 10% DMSO).

#### Notes:

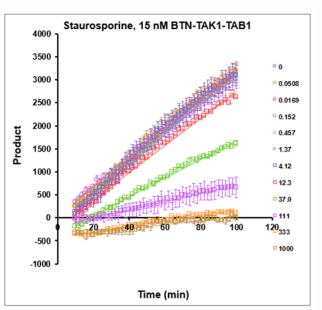
## IC<sub>50</sub> Determination

AssayQuant®

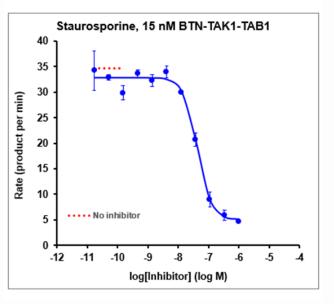
Progress Curves and IC<sub>50</sub> Curves and Table

## Inhibitor Titration Progress Curves

# Linear Region of Progress Curves



### IC<sub>50</sub> Curve



### IC<sub>50</sub> Table

Parameter	Value
Bottom	4.9
Top	32.8
log IC50	-7.37
IC50 (nM)	42.73
Ki (nM)	21.37
Slope	-1.883
R squared	0.990
IC50 approx SE (nM)	1.43
50% inhibition (nM)	51.71

The Y-axis label is RFU/min.

Staurosporine IC<sub>50</sub> Determination at ATP K<sub>m</sub> is 43 nM

## Summary

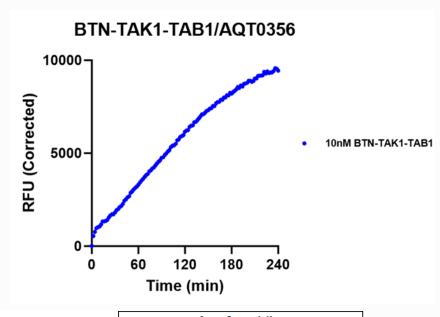


### Assay Validation Results and Progress Curve and Assay Strength at 1 mM ATP

Experiment	Result
Enzyme Titration Linear Range	0.16-5 nM
Sensor Peptide K <sub>m</sub> Value	5.4 μΜ
ATP K <sub>m</sub> Value	25 μΜ
DMSO Tolerance (highest % recommended)	6
Staurosporine IC <sub>50</sub> Determination at ATP K <sub>m</sub>	43 nM

Kinase Name	Conc. (nM)	Sox-based Substrate Name	Normalized Reaction Rate (RFU/pmole/min)	Normalized Rate StndError (RFU/pmole/min)
TAK1	10	AQT0356	233	1

### **Progress Curve**



Assa	Assay Strength Key	
Very Strong	>1,000 (RFU/pmole/min)	
Strong	300 to 999 (RFU/pmole/min)	
Moderate	100 to 299 (RFU/pmole/min)	
Weak	30 to 99 (RFU/pmole/min)	

Under the conditions utilized for this experiment, the assay is Moderate