

# AQT0661 - EPHB2 Assay Validation

PhosphoSens®-Kinetic Assay Format

# Outline for this Study



### PhosphoSens-Kinetic Assay Validation

### **Enzyme Source, Construct, and Lot Information:**

Carna EPHB2 (Cat/Lot #, 08-129/10CBS-0314H) amino acids 581-987(end), N-term GST tag

### **Reference Compound Information:**

Staurosporine MedChemExpress (Cat#/Lot#: HY-15141/125391) CAS No.: 62996-74-1

### **Experiments to be run:**

**Enzyme Titration** 

Sensor Peptide K<sub>m</sub> Determination

ATP K<sub>m</sub> Determination

**DMSO Tolerance Test** 

Reference Compound  $IC_{50}$  Determination at ATP  $K_m$ 

# **Enzyme Titration**

# AssayQuant®

### Reaction Conditions and Set Up

### **Reaction Conditions:**

54 mM HEPES, pH 7.5

1mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

 $15 \, \mu M \, AQT0661$ 

0.005, 0.01, 0.02, 0.04, 0.08, 0.16, 0.3125, 0.625, 1.25, 2.5, 5 and 10 nM Carna EPHB2

### **Reaction Set Up:**

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 or 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

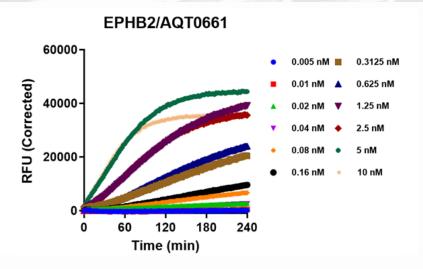
#### Notes:

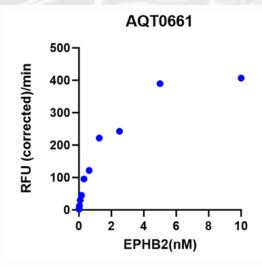
# **Enzyme Titration**

AssayQuant®

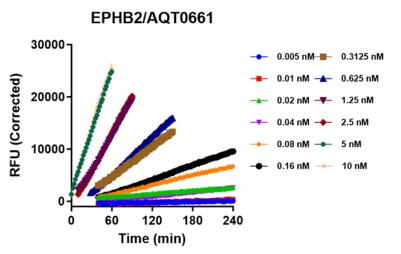
**Progress Curves** 

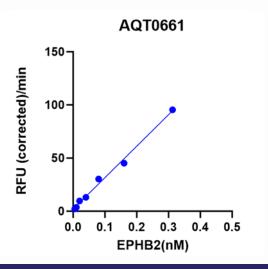
Complete Progress Curves





Linear Region of Progress Curves





Linear Range

# **Enzyme Titration**



### Reaction Rate Table

France Cons. (nDA)	Normalized	Normalized Rate
Enzyme Conc. (nM)	Reaction Rate (RFU/pmole/min)	Stnd Error (RFU/pmole/min)
0.005	21,950	927
0.01	18,190	536
0.02	48,610	450
0.04	32,800	338
0.08	37,938	158
0.16	28,263	89
0.3125	29,228	99
0.625	19,488	76
1.25	17,760	127
2.5	9,708	55
5	7,796	26
10	4,068	39

The reaction is linear from 0.04 - 0.31 nM

# Sensor Peptide K<sub>m</sub> Determination



### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

1mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

1, 2, 3, 5, 7, 10, 15, 20, 30, 50, 70, & 100 μM AQT0661

0.6 nM Carna EPHB2

### **Reaction Set Up:**

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

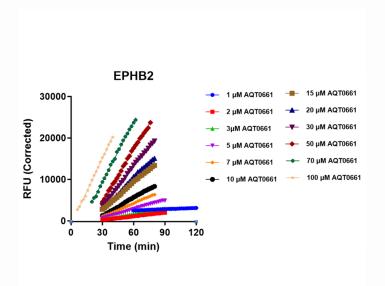
#### Notes:

# Sensor Peptide K<sub>m</sub> Determination

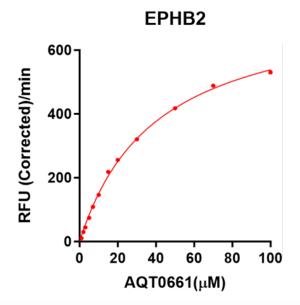


Titration Curves and K<sub>m</sub> Plot and Table

### Sensor Peptide Titration Curves



## Sensor Peptide K<sub>m</sub> Plot



Sensor Peptide K<sub>m</sub> is 40 µM

## Sensor Peptide K<sub>m</sub> Table

Michaelis-Menten	
Best-fit values	
Vmax	754.1
Km	40.16
Std. Error	
Vmax	17.67
Km	2.019
95% CI (profile likelihood)	
Vmax	717.1 to 795.1
Km	36.00 to 44.91
Goodness of Fit	
Degrees of Freedom	10
R squared	0.9983

# ATP K<sub>m</sub> Determination



### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

0, 2.0, 3.9, 7.8, 16, 31, 63, 125, 250, 500, 1000, and 2000 μM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

15 μM AQT0661

2 nM Carna EPHB2

### **Reaction Set Up:**

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

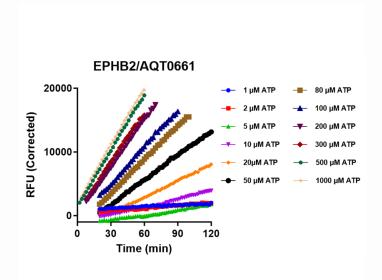
#### Notes:

# ATP K<sub>m</sub> Determination

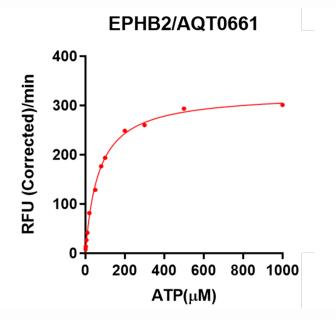


Titration Curves and K<sub>m</sub> Plot and Table

# ATP Titration Curves



### ATP K<sub>m</sub> Plot



ATP K<sub>m</sub> is 68 µM

## ATP K<sub>m</sub> Table

Michaelis-Menten	
Best-fit values	
Vmax	325.3
Km	67.86
Std. Error	
Vmax	4.808
Km	3.554
95% CI (profile likelihood)	
Vmax	314.7 to 336.2
Km	60.30 to 76.24
Goodness of Fit	
Degrees of Freedom	10
R squared	0.9977

## **DMSO Tolerance Test**



### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

1 mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

0, .01, .02, .04, .08, .16, .31, .63, 1.3, 2.5, 5.0, and 10% DMSO

15 μM AQT0661

2 nM Carna EPHB2

### **Reaction Set Up:**

2 or 2.5 μL 10x DMSO dilutions

14 or 17.5 μL Reaction Mix with Sensor Peptide, ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

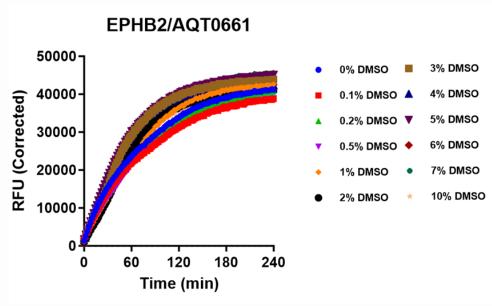
#### Notes:

# **DMSO Tolerance Test**

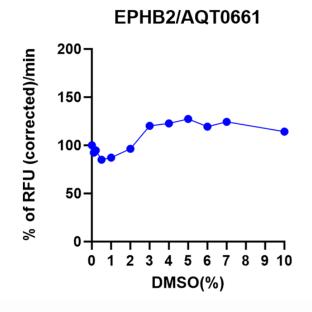


Titration Curves and Inhibition Plot

## Complete Progress Curves



# Reaction Rate vs [DMSO] Plot



No change in enzyme activity out to 2% DMSO

# IC<sub>50</sub> Determination

# Assay Quant®

### Reaction Conditions and Set Up

### **Reaction Conditions:**

54 mM HEPES, pH 7.5

ATP at K<sub>m</sub>

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

1% DMSO

15 μM AQT0661

2 nM Carna EPHB2

0.1 mM Staurosporine was serially diluted (3-fold, 11-point) in 100% DMSO. The series was then diluted 10-fold into BSA (with a final concentration of 0.2 mg/mL BSA in 10% DMSO) to prepare the 10x compound stocks.

### Reaction Set Up:

16 μL Reaction Mix with Sensor Peptide and Inhibitor

 $4 \mu L$  1x EDB or Kinase dilutions (5x in EDB)

20 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Inhibitors are added via direct (0.4  $\mu$ L of 50X stock in 100% DMSO) or intermediate dilutions (2.0  $\mu$ L of 10X stock in 10% DMSO).

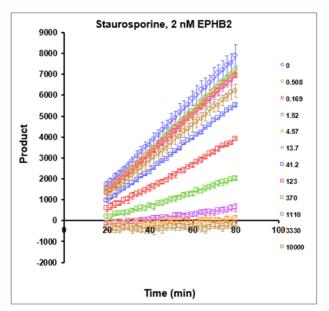
#### Notes:

# IC<sub>50</sub> Determination

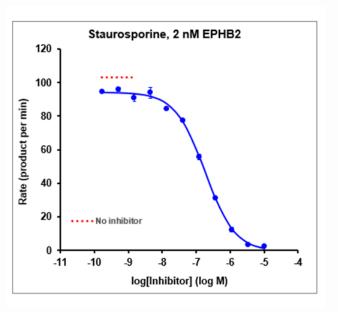


Progress Curves and IC<sub>50</sub> Curves and Table

# Linear Region of Progress Curves



### IC<sub>50</sub> Curve



IC<sub>50</sub> Table

_	Parameter	Value
	Bottom	-0.8
	Top	94.2
	log IC50	-6.74
	IC50 (nM)	180.11
	Ki (nM)	90.06
	Slope	-0.985
	R squared	0.998
C50 a <sub>l</sub>	0.00	
50% in	hibition (nM)	177.05

The Y-axis label is RFU/min.

Staurosporine IC<sub>50</sub> Determination at ATP K<sub>m</sub> is 180 nM

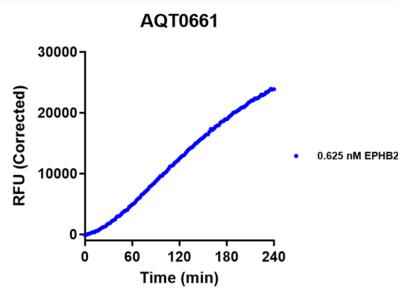
# Summary



### Assay Validation Results and Progress Curve and Assay Strength at 1 mM ATP

Experiment	Result
Enzyme Titration Linear Range	0.04 - 0.31 nM
Sensor Peptide K <sub>m</sub> Value	40 μΜ
ATP K <sub>m</sub> Value	68 μΜ
DMSO Tolerance (highest % recommended)	2
Staurosporine IC50 Determination at ATP Km is	180 nM

## Progress Curve



Kinase Name	Conc. (nM)	Sox-based Substrate Name	Normalized Reaction Rate (RFU/pmole/min)	Normalized Rate StndError (RFU/pmole/min)	Maximum Signal:Bkgd (S/B) Kinetic
EPHB2	0.625	AQT0661	19,488	76	2.7

Assa	Assay Strength Key	
Very Strong	>1,000 (RFU/pmole/min)	
Strong	300 to 999 (RFU/pmole/min)	
Moderate	100 to 299 (RFU/pmole/min)	
Weak	30 to 99 (RFU/pmole/min)	

Under the conditions utilized for this experiment, the assay is Very Strong