

# AQT0661 - JAK2 (JH1) Assay Validation

PhosphoSens®-Kinetic Assay Format

## **Outline for this Study**



PhosphoSens-Kinetic Assay Validation

### **Enzyme Source, Construct, and Lot Information:**

Carna JAK2(JH1) (Cat/Lot #, 08-045/14CBS-0374D) amino acids 826-1132(end), N-term GST tag

### **Reference Compound Information:**

Staurosporine MedChemExpress(Cat#/Lot#: HY-15141/125391)

### **Experiments to be run:**

**Enzyme Titration** 

Sensor Peptide K<sub>m</sub> Determination

ATP K<sub>m</sub> Determination

**DMSO Tolerance Test** 

Reference Compound IC<sub>50</sub> Determination at ATPK<sub>m</sub>

# **Enzyme Titration**

## Reaction Conditions and Set Up

### **Reaction Conditions:**

54 mM HEPES, pH 7.5 1mM ATP 1.2 mM DTT 0.012% Brij-35 1% glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl<sub>2</sub>

15 µM AQT0661

 $0.005,\,0.01,\,0.02,\,0.04,\,0.08,\,0.16,\,0.3125,\,0.625,\,1.25,\,2.5,\,5$  and 10 nM JAK2 (JH1)

# Reaction Set Up:<br/>2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 or 25 µL final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 µL final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

### Notes:





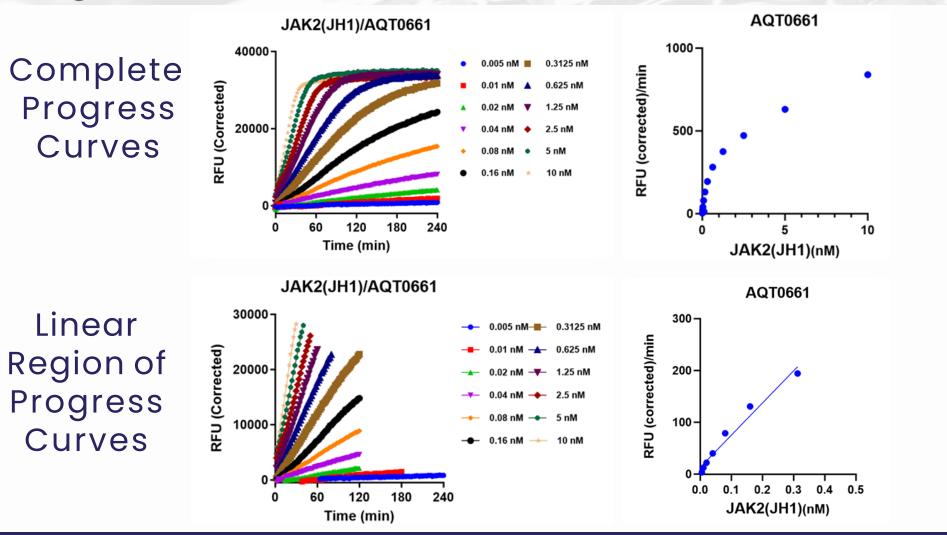
## **Enzyme Titration**

AssayQuant®

Linear

Range

### **Progress Curves**



#### How Can We Help? For technical questions, please reach out at hello@assayguant.com

# **Enzyme Titration**

## AssayQuant<sup>®</sup> TECHNOLOGIES INC.

### **Reaction Rate Table**

	Normalized	Normalized Rate		
Enzyme Conc. (nM)	Reaction Rate (RFU/pmole/min)	Stnd Error (RFU/pmole/min)		
0.005	34,170	886		
0.01	60,300	1,128		
0.02	109,100	1,233		
0.04	99,975	938		
0.08	98,550	347		
0.16	81,563	388		
0.3125	60,688	304		
0.625	44,928	153		
1.25	30,040	212		
2.5	18,876	285		
5	12,612	119		
10	8,392	120		

## The reaction is linear from 0.01 - 0.3125 nM

# Sensor Peptide K<sub>m</sub> Determination



**Reaction Conditions and Set Up** 

### **Reaction Conditions:**

54 mM HEPES, pH 7.5 1mM ATP 1.2 mM DTT 0.012% Brij-35 1%glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl<sub>2</sub> 1, 2, 3, 5, 7, 10, 15, 20, 30, 50, 70, & 100 μM AQT0661 0.2 nM JAK2(JH1)

# Reaction Set Up:<br/>2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

#### Notes:

# **Sensor Peptide K<sub>m</sub> Determination**

Titration Curves and K<sub>m</sub> Plot and Table

Sensor Peptide Titration Curves

JAK2(JH1)/AQT0661

30

60

Time (min)

90

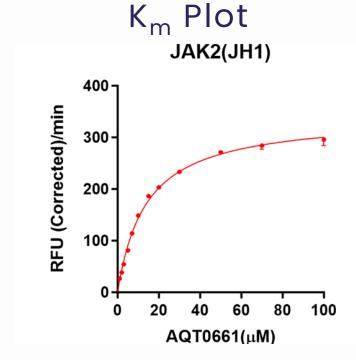
120

20000-

15000

10000

**RFU (Corrected)** 



Sensor Peptide

## Sensor Peptide K<sub>m</sub> Table

Michaelis-Menten	
Best-fit values	
Vmax	341.6
Km	13.84
Std. Error	
Vmax	5.656
Km	0.6763
95% CI (profile likelihood)	
Vmax	329.4 to 354.3
Km	12.43 to 15.40
Goodness of Fit	
Degrees of Freedom	10
R squared	0.9971

### Sensor Peptide K<sub>m</sub> is 13.8 µM



# **ATP K<sub>m</sub> Determination**



### **Reaction Conditions and Set Up**

### **Reaction Conditions:**

54 mM HEPES, pH 7.5
0, 2.0, 3.9, 7.8, 16, 31, 63, 125, 250, 500, 1000, and 2000 $\mu M$ ATP
1.2 mM DTT
0.012% Brij-35
1%glycerol
0.2 mg/ml BSA
0.55 mM EGTA
10 mM MgCl <sub>2</sub>
15 μM AQT0661
3 nM JAK2(JH1)

# Reaction Set Up:<br/>2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

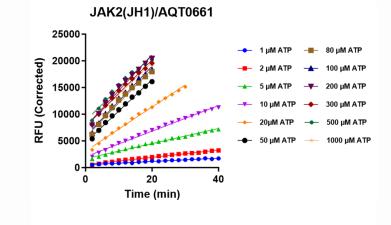
Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

#### Notes:

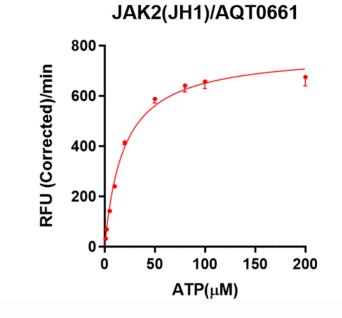
# **ATP K<sub>m</sub> Determination**

Titration Curves and K<sub>m</sub> Plot and Table

ATP Titration Curves



## ATP $K_m$ Plot



## ATP K<sub>m</sub> Table

Michaelis-Menten	
Best-fit values	
Vmax	774.5
Km	19.07
Std. Error	
Vmax	21.68
Km	1.939
95% CI (profile likelihood)	
Vmax	727.1 to 826.7
Km	15.17 to 23.99
Goodness of Fit	
Degrees of Freedom	7
R squared	0.9939

## ATP K<sub>m</sub> is 19.1 µM



## **DMSO Tolerance Test**



### **Reaction Conditions and Set Up**

### **Reaction Conditions:**

54 mM HEPES, pH 7.5 1 mM ATP 1.2 mM DTT 0.012% Brij-35 1% glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl<sub>2</sub> 0, .01, .02, .04, .08, .16, .31, .63, 1.3, 2.5, 5.0, and 10% DMSO 15 μM AQT0661 0.2 nM JAK2(JH1)

# Reaction Set Up:<br/>2 or 2.5 μL10x DMSO dilutions14 or 17.5 μLReaction Mix with Sensor Peptide, ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

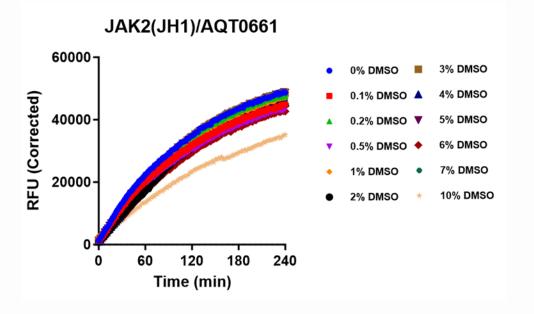
#### Notes:

## **DMSO Tolerance Test**

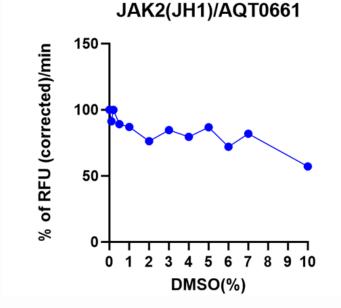


**Titration Curves and Inhibition Plot** 

## Complete Progress Curves







No change in enzyme activity out to 1% DMSO

# IC<sub>50</sub> Determination

### **Reaction Conditions and Set Up**

### **Reaction Conditions:**

54 mM HEPES, pH 7.5 ATP at K<sub>m</sub> 1.2 mM DTT 0.012% Brij-35 1%glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl<sub>2</sub>

1% DMSO

15 µM AQT0661

2 nM JAK2(JH1)

0.1 mM Staurosporine with 3-fold titration in 100% DMSO then diluted 10-fold into BSA (with a final concentration of 0.2 mg/ml) for a DMSO concentration of 10% before diluted 10-fold into reaction mixture with a final DMSO concentration of 1%

### **Reaction Set Up:**

- 16 μL Reaction Mix with Sensor Peptide and Inhibitor
- $4 \mu L$  1x EDB or Kinase dilutions (5x in EDB)
- 20 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Inhibitors are added via direct (0.4  $\mu$ L of 50X stock in 100% DMSO) or intermediate dilutions (2.0  $\mu$ L of 10X stock in 10% DMSO).

### Notes:



# IC<sub>50</sub> Determination



Progress Curves and IC<sub>50</sub> Curves and Table

16000

14000

12000

10000

8000

6000

4000

2000

-2000

Product

## Linear Region of Progress Curves

Staurosporine, 2nM JAK2(JH1)

Time (min)

IC<sub>50</sub> Curve

Staurosporine, 2nM JAK2(JH1)

.....

••••No inhibitor

-10

-9

-8

log[Inhibitor] (log M)

-11

500

450

400

350

300

250 200

150

100

50

-50

Rate (product per min)



Parameter	Value
Bottom	-2.6
Тор	368.1
log IC50	-8.67
IC50 (nM)	2.13
Ki (nM)	1.06
Slope	-1.280
R squared	0.996
IC50 approx SE (nM)	0.00
50% inhibition (nM)	2.10

The Y-axis label is RFU/min.

-5

-6

Staurosporine IC<sub>50</sub> at ATP K<sub>m</sub> is 2.13 nM

0.0508

0.0169

0.152

0.75

0 11 1

30 1000

# Summary



### Assay Validation Results and Progress Curve and Assay Strength at 1 mM ATP

Experiment			Result Progress			essCur	ve		
Enzyme Titration Linear Range			0.01 - 0.3125 nM			AQT0661			
Sensor Peptide K <sub>m</sub> Value			13.8 μM						
ATP K <sub>m</sub> Value		19.1 μM		- 20000 - 2000 - 20000 - 2000 - 200 - 2000 - 2000		•	0.16 nM JAK2(JH1)		
DMSO Tolerance (highest	d)	1							
Staurosporine IC <sub>50</sub> Determin		2.13 nM		0- <b>-</b>	60 120	180 240			
						Time (m	in)		
		Cov boood	Normalized	Normalized Rate StndError		Assay Strength Key			
Kinase Name	Kinase Name Conc. (nM)	Sox-based	<b>Reaction Rate</b>			Very Strong			
		Substrate Name	Substrate Name (RFU/pmole/min			Strong Moderate	300 to 999 (RFU/pmole 100 to 299 (RFU/pmole	· · ·	

## Under the conditions utilized for this experiment, the assay is Very Strong

81,563

388

AQT0661

0.16

JAK2(JH1)

Weak

30 to 99 (RFU/pmole/min)