

# AQT0661 - JAK3 (JH1) Assay Validation

PhosphoSens®-Kinetic Assay Format

# **Outline for this Study**



PhosphoSens-Kinetic Assay Validation

### **Enzyme Source, Construct, and Lot Information:**

Carna JAK3(JH1) (Cat/Lot #, 08-046/19CBS-0798B) amino acids 795-1124, N-term His tag

### **Reference Compound Information:**

Staurosporine MedChemExpress(Cat#/Lot#: HY-15141/125391)

### **Experiments to be run:**

**Enzyme Titration** 

Sensor Peptide K<sub>m</sub> Determination

ATP K<sub>m</sub> Determination

**DMSO Tolerance Test** 

Reference Compound IC<sub>50</sub> Determination at ATPK<sub>m</sub>

# **Enzyme Titration**

# Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5 1mM ATP 1.2 mM DTT 0.012% Brij-35 1% glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl<sub>2</sub>

15 µM AQT0661

 $0.005,\,0.01,\,0.02,\,0.04,\,0.08,\,0.16,\,0.3125,\,0.625,\,1.25,\,2.5,\,5,\,\text{and}\,10$  nM JAK3 (JH1)

# Reaction Set Up:<br/>2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 or 25 µL final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 µL final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

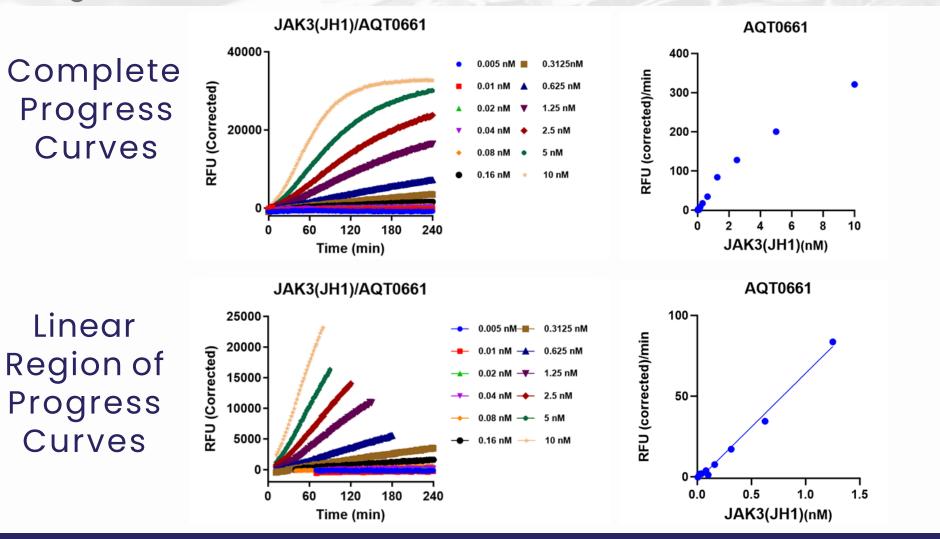
#### Notes:





# **Enzyme Titration**

**Progress Curves** 





Linear

Range

# **Enzyme Titration**

## AssayQuant<sup>®</sup> TECHNOLOGIES INC.

## **Reaction Rate Table**

Enzyme Conc. (nM)	Normalized	Normalized Rate			
Enzyme conc. (mvi)	Reaction Rate (RFU/pmole/min)	Stnd Error (RFU/pmole/min)			
0.005	-3,257	1,159			
0.01	5,550	496			
0.02	8,170	503			
0.04	4,793	296			
0.08	4,735	89			
0.16	4,713	50			
0.3125	5,331	33			
0.625	5,506	18			
1.25	6,693	32			
2.5	5,112	32			
5	4,004	23			
10	3,209	22			

## The reaction is linear from 0.16 - 1.25 nM

# Sensor Peptide K<sub>m</sub> Determination



**Reaction Conditions and Set Up** 

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5 1mM ATP 1.2 mM DTT 0.012% Brij-35 1%glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl<sub>2</sub> 1, 2, 3, 5, 7, 10, 15, 20, 30, 50, 70, & 100 μM AQT0661 2 nM JAK3(JH1)

# Reaction Set Up:<br/>2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

#### Notes:

# **Sensor Peptide K<sub>m</sub> Determination**

Titration Curves and K<sub>m</sub> Plot and Table

Sensor Peptide Titration Curves

1 JIM AOT066

JAK3(JH1)/AQT0661

25000

20000

15000

10000· 5000·

30

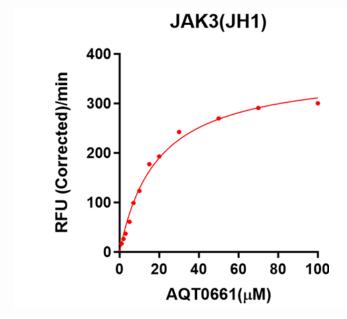
60

Time (min)

90

120

**RFU (Corrected)** 



Sensor Peptide

K<sub>m</sub> Plot

## Sensor Peptide K<sub>m</sub> Table

Michaelis-Menten	
Best-fit values	
Vmax	370.3
Km	19.01
Std. Error	
Vmax	13.00
Km	1.797
95% CI (profile likelihood)	
Vmax	343.9 to 400.0
Km	15.56 to 23.27
Goodness of Fit	
Degrees of Freedom	10
R squared	0.9911

## Sensor Peptide K<sub>m</sub> is 19 µM



# **ATP K<sub>m</sub> Determination**



## **Reaction Conditions and Set Up**

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5
0, 2.0, 3.9, 7.8, 16, 31, 63, 125, 250, 500, 1000, and 2000 $\mu M$ ATP
1.2 mM DTT
0.012% Brij-35
1%glycerol
0.2 mg/ml BSA
0.55 mM EGTA
10 mM MgCl <sub>2</sub>
15 μM AQT0661
3 nM JAK3(JH1)

# Reaction Set Up:<br/>2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

#### Notes:

# **ATP K<sub>m</sub> Determination**

Titration Curves and K<sub>m</sub> Plot and Table

**ATP Titration** Curves

JAK3(JH1)/AQT0661

20

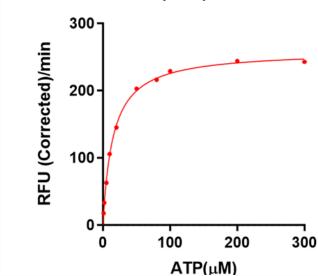
15000.

10000.

5000

0

RFU (Corrected)



ATP K<sub>m</sub> Plot

**JAK3(JH1)/AQT0661** 

## ATP K<sub>m</sub> Table

Michaelis-Menten	
Best-fit values	
Vmax	259.1
Km	14.97
Std. Error	
Vmax	2.198
Km	0.5491
95% CI (profile likelihood)	
Vmax	254.1 to 264.3
Km	13.76 to 16.29
Goodness of Fit	
Degrees of Freedom	8
R squared	0.9989

## ATP K<sub>m</sub> is 15 µM

#### 40 60 Time (min)



# **DMSO Tolerance Test**



### **Reaction Conditions and Set Up**

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5 1 mM ATP 1.2 mM DTT 0.012% Brij-35 1% glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl<sub>2</sub> 0, .01, .02, .04, .08, .16, .31, .63, 1.3, 2.5, 5.0, and 10% DMSO 15 μM AQT0661 2 nM JAK3(JH1)

# Reaction Set Up:<br/>2 or 2.5 μL10x DMSO dilutions14 or 17.5 μLReaction Mix with Sensor Peptide, ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

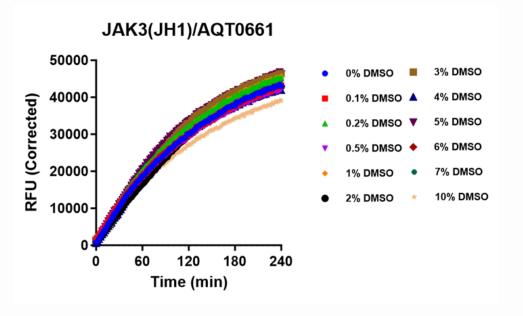
#### Notes:

## **DMSO Tolerance Test**

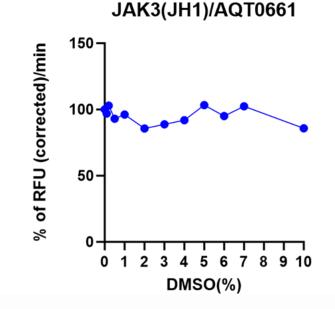


**Titration Curves and Inhibition Plot** 

## Complete Progress Curves







No change in enzyme activity out to 1% DMSO

# IC<sub>50</sub> Determination

## **Reaction Conditions and Set Up**

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5 ATP at K<sub>m</sub> 1.2 mM DTT 0.012% Brij-35 1%glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl<sub>2</sub>

1% DMSO

15 µM AQT0661

2 nM JAK3(JH1)

0.1 mM Staurosporine with 3-fold titration in 100% DMSO then diluted 10-fold into BSA (with a final concentration of 0.2 mg/ml) for a DMSO concentration of 10% before diluted 10-fold into reaction mixture with a final DMSO concentration of 1%

#### **Reaction Set Up:**

- 16 μL Reaction Mix with Sensor Peptide and Inhibitor
- $4 \mu L$  1x EDB or Kinase dilutions (5x in EDB)
- 20 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Inhibitors are added via direct (0.4  $\mu$ L of 50X stock in 100% DMSO) or intermediate dilutions (2.0  $\mu$ L of 10X stock in 10% DMSO).

#### Notes:



# IC<sub>50</sub> Determination

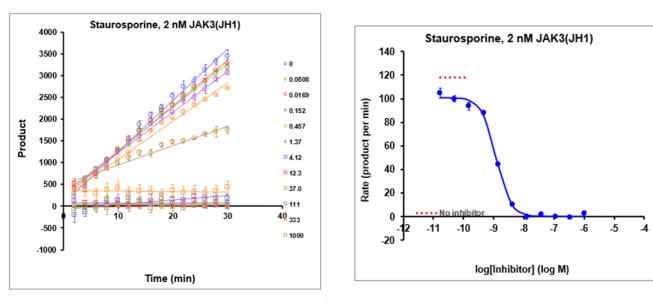


Progress Curves and IC<sub>50</sub> Curves and Table

## Linear Region of Progress Curves

IC<sub>50</sub> Curve





Parameter	Value				
Bottom	0.4				
Тор	100.8				
log IC50	-8.92				
IC50 (nM)	1.21				
Ki (nM)	0.61				
Slope	-1.826				
R squared	0.997				
IC50 approx SE (nM)	0.00				
50% inhibition (nM)	1.22				

The Y-axis label is RFU/min.

Staurosporine IC<sub>50</sub> at ATP K<sub>m</sub> is 1.2 nM

# Summary



## Assay Validation Results and Progress Curve and Assay Strength at 1 mM ATP

Experiment				Res	sult		Progress Curve				ve		
Enzyme Titration Linear Range				0.16 - 1.25 nM				AQT0661					
Sensor Peptide K <sub>m</sub> Value				19	μM		चि 20000-						
ATP K <sub>m</sub> Value				15 μΜ			or	000- 000-		•	2.5 nM JAK3(JH1)		
DMSOTolerance (highest % recommended)			)	1			Ð	000-					
Staurosporine $\rm IC_{50}$ Determination at ATP $\rm K_m$				1.2 nM			0- <b> </b> 0	0-  0	60 120	180 240	240		
									Time (m	in)			
	Kinase Name	e Name   Conc. (nM)	Sox-based	Normalized Reaction Rate	Normalized Rate StndError				Very Strong	> 1,000 (RFU/pmole			
		Substrate Name		(RFU/pmole/min)				Strong Moderate	300 to 999 (RFU/pmo 100 to 299 (RFU/pmo				
	JAK3 (JH1)	2.5	AQT0661	5,112	32				Weak	30 to 99 (RFU/pmole	/min)		

## Under the conditions utilized for this experiment, the assay is Very Strong