

AQT0663 - JAKI (JH1 JH2) Assay Validation

PhosphoSens®-Kinetic Assay Format

Outline for this Study



PhosphoSens-Kinetic Assay Validation

Enzyme Source, Construct, and Lot Information:

SignalChem JAK1 (Cat/Lot # J01-11G/E4140-3) (438 - end) was expressed as an N-terminal GST tag in SF9 insect cells.

Reference Compound Information:

Staurosporine MedChemExpress(Cat#/Lot#: HY-15141/125391)

Experiments to be run:

Enzyme Titration

Sensor Peptide K_m Determination

ATP K_m Determination

DMSO Tolerance Test

Reference Compound IC₅₀ Determination at ATPK_m

Enzyme Titration

Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5 1mM ATP 1.2 mM DTT 0.012% Brij-35 1% glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl₂

 $20 \ \mu\text{M} \ \text{AQT0663}$

0.04 ,0.08 ,0.16 ,0.31 ,0.63 ,1.25,2.5,5,10,20,40,80 nM JAK1 (JH1JH2)

Reaction Set Up:
2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 or 25 µL final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 µL final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Notes:





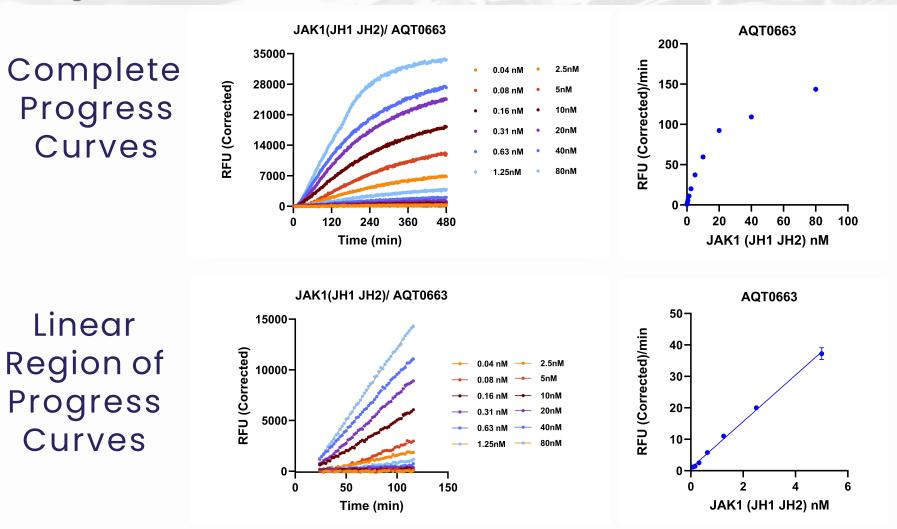
Enzyme Titration



Linear

Range

Progress Curves



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Enzyme Titration

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Reaction Rate Table

Enzyme Conc. (nM)	Reaction Rate Standard Erro (RFU/min) (RFU/min)		Normalized Reaction Rate (RFU/pmole/min)	Normalized Rate Standard Error (RFU/pmole/min)	
0.04	0.7	0.3	851	363	
0.08	1.2	0.4	753	228	
0.16	1.4	0.3	449	89 41	
0.31	2.5	0.3	404		
0.63	6	0	457	34	
1.25	11	0	441	16	
2.50	20	0	400	8	
5.00	37	2	372	19	
10.0	60	0	298	2	
20	92	0	231	1	
40	109	0	137	1	
80	144	1	90	0	

The reaction is linear from 0.16 - 5 nM

Sensor Peptide K_m Determination



Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5 1mM ATP 1.2 mM DTT 0.012% Brij-35 1%glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl₂ 1.2,1.7,2.6,3.9,5.9,8.8,13,20,30,44,67,100 μM AQT0663 15 nM JAK1(JH1 JH2)

Reaction Set Up:
2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Notes:

Sensor Peptide K_m Determination

Titration Curves and K_m Plot and Table

Sensor Peptide Titration Curves

100

JAK1(JH1 JH2)/ AQT0663

75

Time (min)

12000-

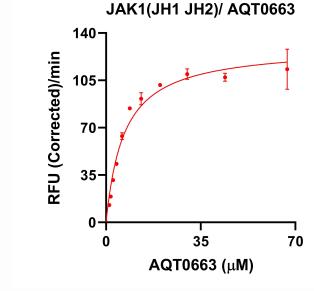
8000

4000

0

50

RFU (Corrected)



Sensor Peptide

K_m Plot

Sensor Peptide K_m Table

Michaelis-Menten			
Best-fit values			
Vmax	130.2		
Km	6.655		
Std. Error			
Vmax	5.702		
Km	0.9453		
95% CI (asymptotic)			
Vmax	117.3 to 143.1		
Km	4.516 to 8.793		
Goodness of Fit			
Degrees of Freedom	9		
R squared	0.9739		

Sensor Peptide K_m is 6.7 µM



ATP K_m Determination



Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5
0, 2.0, 3.9, 7.8, 16, 31, 63, 125, 250, 500, 1000, and 2000 μM ATP
1.2 mM DTT
0.012% Brij-35
1%glycerol
0.2 mg/ml BSA
0.55 mM EGTA
10 mM MgCl ₂
20 μM AQT0663
15 nM JAK1(JH1 JH2)

Reaction Set Up:
2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Notes:

ATP K_m Determination

Titration Curves and K_m Plot and Table

75 uM

300 uN

600 µМ 1000 µМ

ATP Titration Curves

JAK1 (JH1 JH2)/ AQT0663

50

Time (min)

100

150

4000-

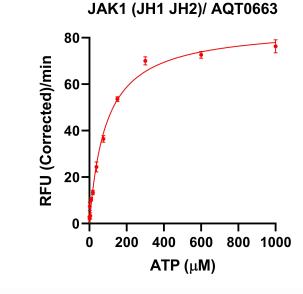
3000

2000-

1000

-1000

RFU (Corrected)



ATP K_m Plot

ATP K_m Table

84.89		
89.25		
2.583		
9.472		
79.13 to 90.64		
68.15 to 110.4		
10		
0.9923		

ATP K_m is 89 μM

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DMSO Tolerance Test



Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5 1 mM ATP 1.2 mM DTT 0.012% Brij-35 1% glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl₂ 0, .01, .02, .04, .08, .16, .31, .63, 1.3, 2.5, 5.0, and 10% DMSO 20 μM AQT0663

15 nM JAK1(JH1 JH2)

Reaction Set Up:
2 or 2.5 μL10x DMSO dilutions14 or 17.5 μLReaction Mix with Sensor Peptide, ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

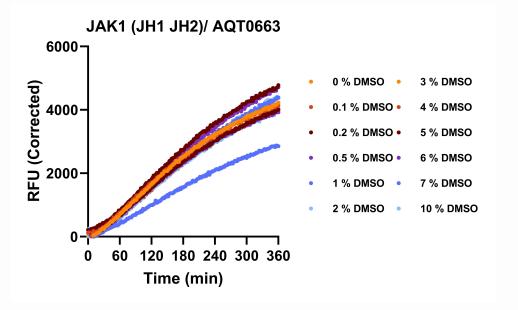
Notes:

DMSO Tolerance Test

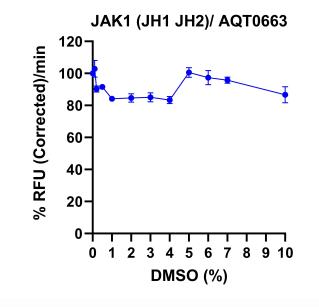


Titration Curves and Inhibition Plot

Complete Progress Curves



Reaction Rate vs [DMSO] Plot



No change in enzyme activity out to 2% DMSO

IC₅₀ Determination

Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5 ATP at K_m 1.2 mM DTT 0.012% Brij-35 1%glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl₂ 2% DMSO

20 µM AQT0663

15 nM JAK1(JH1 JH2)

0, 0.02, 0.05, 0.15, 0.46, 1.37, 4.12, 12.35, 37.04, 111.11, 333.33, and 1000 nM Staurosporine

Reaction Set Up:

- 16 μL Reaction Mix with Sensor Peptide and Inhibitor
- $4 \mu L$ 1x EDB or Kinase dilutions (5x in EDB)
- 20 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Inhibitors are added via direct (0.4 μL of 50X stock in 100% DMSO) or intermediate dilutions (2.0 μL of 10X stock in 10% DMSO).

Notes:



IC₅₀ Determination



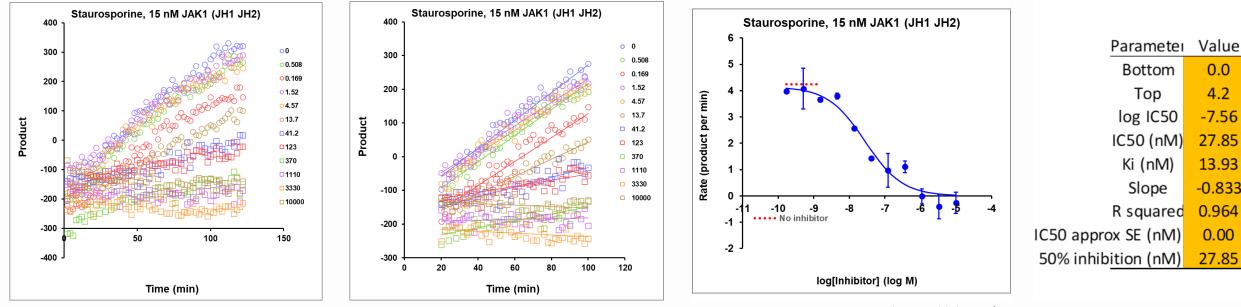
Progress Curves and IC₅₀ Curves and Table

Inhibitor Titration Progress Curves

Linear Region of Progress Curves

IC₅₀ Curve

IC₅₀ Table



The Y-axis label is RFU/min.

Staurosporine IC₅₀ at ATP K_m is 28 nM

Summary



Assay Validation Results and Progress Curve and Assay Strength at 1 mM ATP

Experiment	Result	Progress Curve
Enzyme Titration Linear Range	0.16 - 5 nM	AQT0663
Sensor Peptide K _m Value	6.7 μΜ	cted)
ATP K _m Value	89 µM	• 5nM JAK1(JH1 JH2)
DMSOTolerance (highest % recommended)	2	RE
Staurosporine IC_{50} Determination at ATP K_m	28 nM	0- 0 120 240 360 480 Time (min)

Kinase Name	Sox-based substrate name	Normalized	Normalized	Assay Strength Key		
		Sox-based substrate name	Reaction Rate	Reaction Rate	Very Strong	>1,000 (RFU/pmole/min)
				Stnd Error	Strong	300 to 999 (RFU/pmole/min)
				(RFU/pmol/min)	Moderate	100 to 299 (RFU/pmole/min)
AK1 (JH1 JH2) (SC)	5	AQT0663	372	19	Weak	30 to 99 (RFU/pmole/min)

Under the conditions utilized for this experiment, the assay is Strong

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