

# AQT0663 - JAK1 (JH1) Assay Validation

PhosphoSens®-Kinetic Assay Format

# Outline for this Study



### PhosphoSens-Kinetic Assay Validation

### **Enzyme Source, Construct, and Lot Information:**

Invitrogen (Cat # PR8767C) Recombinant human, catalytic domain (amino acids 866-1154), GSTtagged, expressed in insect cells.

### **Reference Compound Information:**

Staurosporine MedChemExpress(Cat#/Lot#: HY-15141/125391)

### **Experiments to be run:**

**Enzyme Titration** 

Sensor Peptide K<sub>m</sub> Determination

ATP K<sub>m</sub> Determination

**DMSO Tolerance Test** 

Reference Compound  $IC_{50}$  Determination at ATP  $K_m$ 

# **Enzyme Titration**

# AssayQuant®

### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

1mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

20 μM AQT0663

0.02, 0.04, 0.08, 0.16, 0.31, 0.63, 1.25, 2.5, 5, 10, 20, 40 nM JAK1 (JH1)

### **Reaction Set Up:**

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 or 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

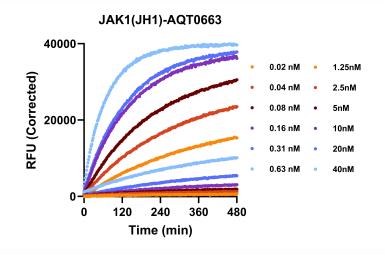
#### Notes:

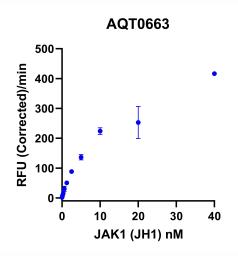
# **Enzyme Titration**

AssayQuant®

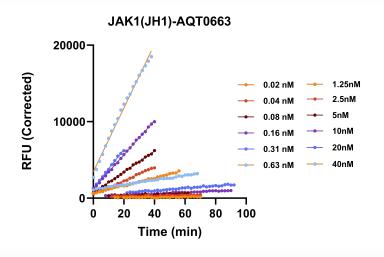
**Progress Curves** 

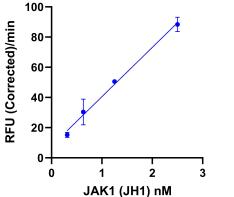
Complete Progress Curves





Linear Region of Progress Curves





**AQT0663-Linear-range** 

Linear Range

# **Enzyme Titration**



### Reaction Rate Table

Enzyme Conc. (nM)	Reaction Rate (RFU/min)	Standard Error (RFU/min)	Normalized Reaction Rate (RFU/pmole/min)	Normalized Rate Standard Error (RFU/pmole/min)
0.02	1.7	0.6	4368	1500
0.04	2.7	0.9	3353	1125
0.08	5.6	4.4	3506	2738
0.16	8.4	2.2	2634	680
0.31	15	2	2448	272
0.63	30	9	2413	677
1.25	51	1	2022	24
2.50	88	5	<i>17</i> 67	95
5.0	136	9	1364	90
10	224	11	1120	55
20	253	54	632	134
40	416	6	521	8

The reaction is linear from 0.31 - 2.5 nM

# Sensor Peptide K<sub>m</sub> Determination



### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

1mM ATP

1.2 mM DTT

0.012% Brij-35

1%glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

1.2,1.7,2.6,3.9,5.9,8.8,13,20,30,44,67,100 μM AQT0663

10 nM JAK1(JH1)

### **Reaction Set Up:**

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

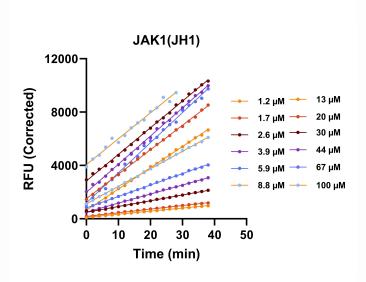
#### Notes:

# Sensor Peptide K<sub>m</sub> Determination

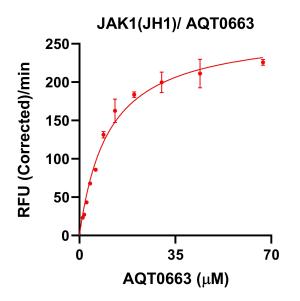


Titration Curves and K<sub>m</sub> Plot and Table

### Sensor Peptide Titration Curves



## Sensor Peptide K<sub>m</sub> Plot



Sensor Peptide K<sub>m</sub> is 11 µM

## Sensor Peptide K<sub>m</sub> Table

Michaelis-Menten	
Best-fit values	
Vmax	268.6
Km	10.58
Std. Error	
Vmax	10.49
Km	1.176
95% CI (asymptotic)	
Vmax	244.9 to 292.4
Km	7.923 to 13.24
Goodness of Fit	
Degrees of Freedom	9
R squared	0.9870

# ATP K<sub>m</sub> Determination



### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

0, 2.0, 3.9, 7.8, 16, 31, 63, 125, 250, 500, 1000, and 2000 μM ATP

1.2 mM DTT

0.012% Brij-35

1%glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

20 μM AQT0663

10 nM JAK1(JH1)

### **Reaction Set Up:**

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

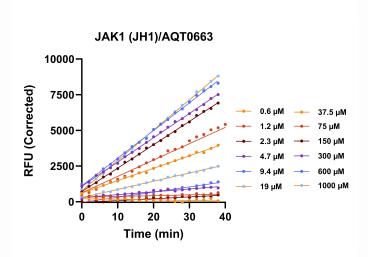
#### Notes:

# ATP K<sub>m</sub> Determination

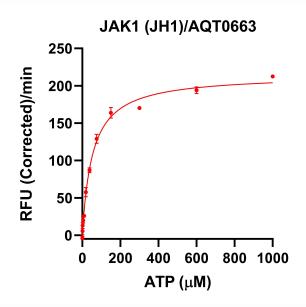


Titration Curves and K<sub>m</sub> Plot and Table

# ATP Titration Curves



### ATP K<sub>m</sub> Plot



ATP  $K_m$  is 54  $\mu$ M

## ATP K<sub>m</sub> Table

Michaelis-Menten	
Best-fit values	
Vmax	215.0
Km	53.70
Std. Error	
Vmax	4.811
Km	4.654
95% CI (asymptotic)	
Vmax	204.2 to 225.7
Km	43.33 to 64.07
Goodness of Fit	
Degrees of Freedom	10
R squared	0.9946

## **DMSO Tolerance Test**



### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

1 mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

0, .01, .02, .04, .08, .16, .31, .63, 1.3, 2.5, 5.0, and 10% DMSO

20 μM AQT0663

10 nM JAK1(JH1)

### **Reaction Set Up:**

2 or 2.5 μL 10x DMSO dilutions

14 or 17.5 μL Reaction Mix with Sensor Peptide, ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

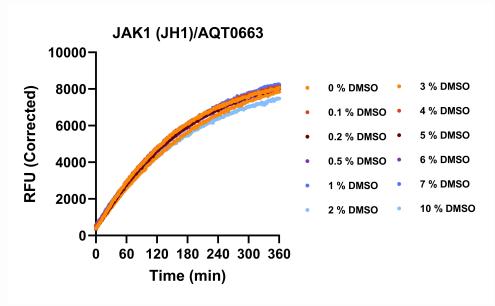
#### Notes:

# **DMSO Tolerance Test**

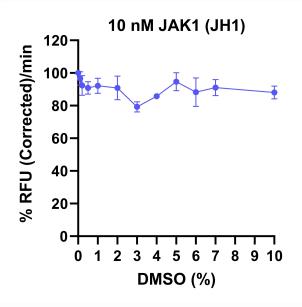


Titration Curves and Inhibition Plot

## Complete Progress Curves



# Reaction Rate vs [DMSO] Plot



No change in enzyme activity out to 2% DMSO

# IC<sub>50</sub> Determination

# AssayQuant®

### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

ATP at K<sub>m</sub>

1.2 mM DTT

0.012% Brij-35

1%glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

2% DMSO

20 µM AQT0663

10 nM JAK1(JH1)

0, 0.02, 0.05, 0.15, 0.46, 1.37, 4.12, 12.35, 37.04, 111.11, 333.33, and 1000 nM Staurosporine

### **Reaction Set Up:**

16 μL Reaction Mix with Sensor Peptide and Inhibitor

 $4 \mu L$  1x EDB or Kinase dilutions (5x in EDB)

20 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Inhibitors are added via direct (0.4  $\mu$ L of 50X stock in 100% DMSO) or intermediate dilutions (2.0  $\mu$ L of 10X stock in 10% DMSO).

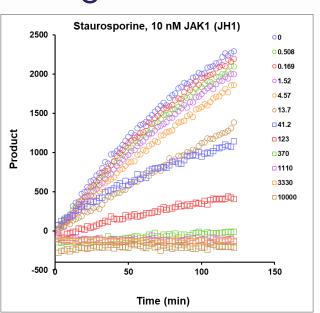
#### Notes:

# IC<sub>50</sub> Determination

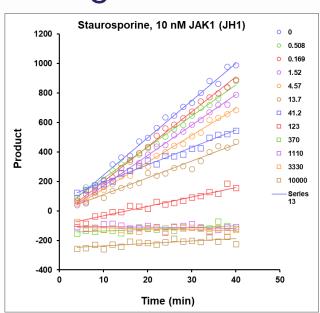


Progress Curves and IC<sub>50</sub> Curves and Table

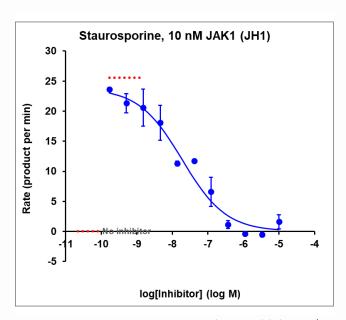
## Inhibitor Titration Progress Curves



# Linear Region of Progress Curves



IC<sub>50</sub> Curve



The Y-axis label is RFU/min.

IC<sub>50</sub> Table

Paramete	Value
Bottom	0.0
Тор	23.8
log IC50	-7.72
IC50 (nM)	19.06
Ki (nM)	9.53
Slope	-0.708
R squared	0.972
C50 approx SE (nM)	1.44
50% inhibition (nM)	19.06

Staurosporine IC<sub>50</sub> at ATP K<sub>m</sub> is 19 nM

# Summary

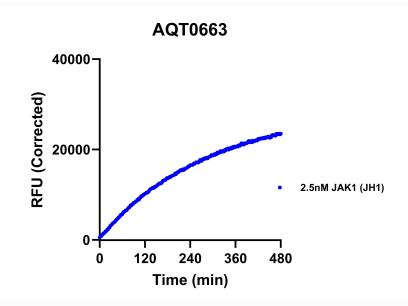


### Assay Validation Results and Progress Curve and Assay Strength at 1 mM ATP

Experiment	Result
Enzyme Titration Linear Range	0.31 - 2.5 nM
Sensor Peptide K <sub>m</sub> Value	11 μΜ
ATP K <sub>m</sub> Value	54 μΜ
DMSO Tolerance (highest % recommended)	2
Staurosporine IC <sub>50</sub> Determination at ATP K <sub>m</sub>	19 nM

Kinase Name	Conc. (nM)	Sox-based substrate name	Normalized Reaction Rate (RFU/pmol/min)	Normalized Reaction Rate Stnd Error (RFU/pmol/min)
JAK1 (JH1) (INVT)	2.5	AQT0663	1767	95

### **Progress Curve**



Assay Strength Key	
Very Strong	>1,000 (RFU/pmole/min)
Strong	300 to 999 (RFU/pmole/min)
Moderate	100 to 299 (RFU/pmole/min)
Weak	30 to 99 (RFU/pmole/min)

Under the conditions utilized for this experiment, the assay is Very Strong