

AQT0663 - TYK2 (JH1JH2) Assay Validation

PhosphoSens®-Kinetic Assay Format

Outline for this Study



PhosphoSens-Kinetic Assay Validation

Enzyme Source, Construct, and Lot Information:

SignalChem TYK2(JH1&JH2 domains) (CAT#/LOT#: T21-11G/J4005-8) amino acids 442-end with N-terminal GST-tag

Reference Compound Information:

Staurosporine MedChemExpress(Cat#/Lot#: HY-15141/125391)

Experiments to be run:

Enzyme Titration

Sensor Peptide K_m Determination

ATP K_m Determination

DMSO Tolerance Test

Reference Compound IC_{50} Determination at ATP K_m

Enzyme Titration

AssayQuant®

Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5

1mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl₂

20 μM AQT0663

0.03 ,0.06 ,0.12 ,0.23 ,0.47 ,0.94 ,1.88 ,3.75,7.5,15,30,60 nM TYK2 (JH1JH2)

Reaction Set Up:

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 or 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

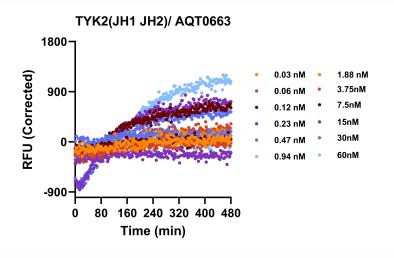
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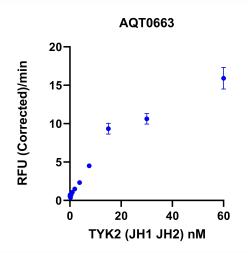
Enzyme Titration

AssayQuant®

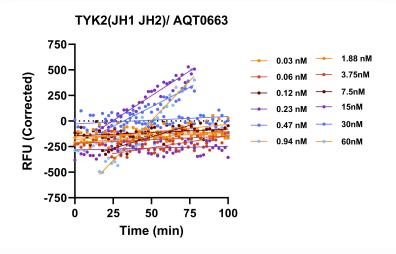
Progress Curves

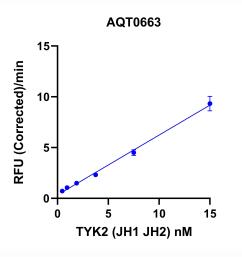
Complete Progress Curves





Linear Region of Progress Curves





Linear Range

Enzyme Titration



Reaction Rate Table

Enzyme Conc. (nM)	Reaction Rate (RFU/min)	Standard Error (RFU/min)	Normalized Reaction Rate (RFU/pmole/min)	Normalized Rate Standard Error (RFU/pmole/min)
0.03	0.5	0.09	901	153
0.06	0.7	0.11	599	96
0.12	0.7	0.10	274	40
0.23	0.4	0.08	79	18
0.47	0.7	0.08	77	9
0.94	1.1	0.08	56	4
1.88	1.5	0.08	40	2
3.75	2.3	0.17	31	2
7.5	4.5	0.28	30	2
15	9.3	0.71	31	2
30	10.6	0.68	18	1
60	15.9	1.40	13	1

The reaction is linear from 0.47 - 15 nM

Sensor Peptide K_m Determination



Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5

1mM ATP

1.2 mM DTT

0.012% Brij-35

1%glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl₂

1.2,1.7,2.6,3.9,5.9,8.8,13,20,30,44,67,100 μM AQT0663

40 nM TYK2(JH1 JH2)

Reaction Set Up:

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

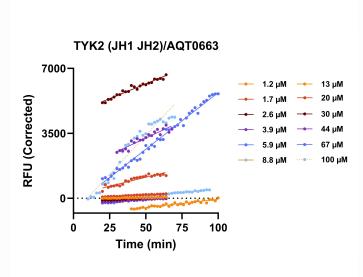
Notes:

Sensor Peptide K_m Determination

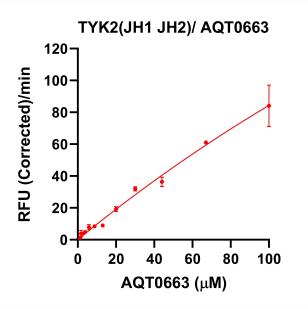


Titration Curves and K_m Plot and Table

Sensor Peptide Titration Curves



Sensor Peptide K_m Plot



Sensor Peptide K_m is 583 µM

Sensor Peptide K_m Table

Michaelis-Menten	
Best-fit values	
Vmax	575.9
Km	583.0
Std. Error	
Vmax	280.9
Km	321.0
95% CI (asymptotic)	
Vmax	-50.03 to 1202
Km	0.000 to 1298
Goodness of Fit	
Degrees of Freedom	10
R squared	0.9922

ATP K_m Determination



Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5

0, 2.0, 3.9, 7.8, 16, 31, 63, 125, 250, 500, 1000, and 2000 μM ATP

1.2 mM DTT

0.012% Brij-35

1%glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl₂

20 μM AQT0663

40 nM TYK2(JH1 JH2)

Reaction Set Up:

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

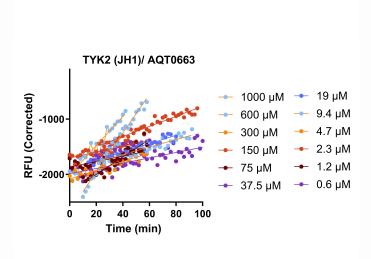
Notes:

ATP K_m Determination

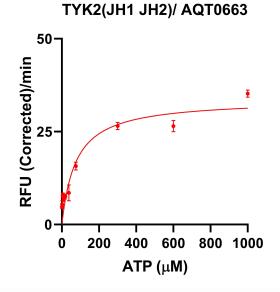


Titration Curves and K_m Plot and Table

ATP Titration Curves



ATP K_m Plot



ATP K_m is 76 μM

ATP K_m Table

Michaelis-Menten	
Best-fit values	
Vmax	33.68
Km	75.82
Std. Error	
Vmax	3.482
Km	29.48
95% CI (asymptotic)	
Vmax	25.80 to 41.56
Km	9.130 to 142.5
Goodness of Fit	
Degrees of Freedom	9
R squared	0.8809

DMSO Tolerance Test



Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5

1 mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl₂

0, .01, .02, .04, .08, .16, .31, .63, 1.3, 2.5, 5.0, and 10% DMSO

20 μM AQT0663

40 nM TYK2(JH1 JH2)

Reaction Set Up:

2 or 2.5 μL 10x DMSO dilutions

14 or 17.5 µL Reaction Mix with Sensor Peptide, ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

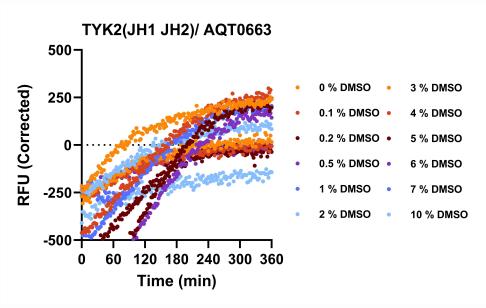
Notes:

DMSO Tolerance Test

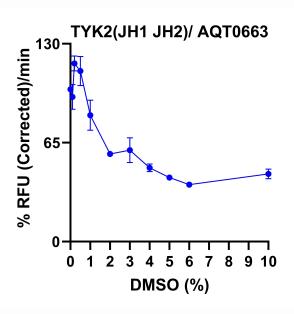


Titration Curves and Inhibition Plot

Complete Progress Curves



Reaction Rate vs [DMSO] Plot



No change in enzyme activity out to 2% DMSO

IC₅₀ Determination

AssayQuant®

Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5

ATP at K_m

1.2 mM DTT

0.012% Brij-35

1%glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl₂

2% DMSO

20 μM AQT0663

40 nM TYK2(JAK1)

0, 0.02, 0.05, 0.15, 0.46, 1.37, 4.12, 12.35, 37.04, 111.11, 333.33, and 1000 nM Staurosporine

Reaction Set Up:

16 μL Reaction Mix with Sensor Peptide and Inhibitor

 $4 \mu L$ 1x EDB or Kinase dilutions (5x in EDB)

20 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Inhibitors are added via direct (0.4 μ L of 50X stock in 100% DMSO) or intermediate dilutions (2.0 μ L of 10X stock in 10% DMSO).

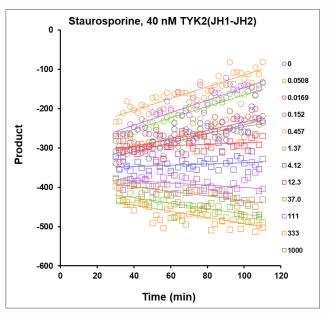
Notes:

IC₅₀ Determination

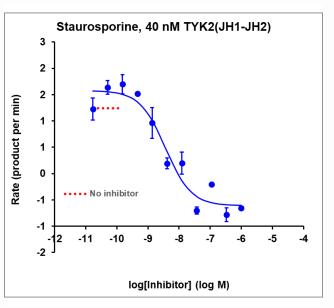


Progress Curves and IC₅₀ Curves and Table

Linear Region of Progress Curves



IC₅₀ Curve



The Y-axis label is RFU/min.

IC₅₀ Table

Paramete	Value
Bottom	-0.6
Тор	1.6
log IC50	-8.43
IC50 (nM)	3.67
Ki (nM)	1.84
Slope	-1.061
R squarec	0.946
IC50 approx SE (nM)	0.00
50% inhibition (nM)	2.14

Staurosporine IC₅₀ at ATP K_m is 3.7 nM

Summary

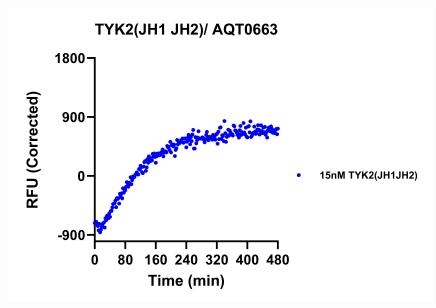


Assay Validation Results and Progress Curve and Assay Strength at 1 mM ATP

Experiment	Result
Enzyme Titration Linear Range	0.47 - 15 nM
Sensor Peptide K _m Value	583 μΜ
ATP K _m Value	76 μΜ
DMSO Tolerance (highest % recommended)	2
Staurosporine IC ₅₀ Determination at ATP K _m	3.7 nM

Kinase Name	Conc. (nM)	Sox-based substrate name	Normalized Reaction Rate (RFU/pmol/min)	Normalized Reaction Rate Stnd Error (RFU/pmol/min)
TYK2(JH1 JH2)	15	AQT0663	31	2

Progress Curve



Assay Strength Key	
Very Strong	>1,000 (RFU/pmole/min)
Strong	300 to 999 (RFU/pmole/min)
Moderate	100 to 299 (RFU/pmole/min)
Weak	30 to 99 (RFU/pmole/min)

Under the conditions utilized for this experiment, the assay is Weak