

# AQT0682 - BRAF Assay Validation

PhosphoSens®-Kinetic Assay Format

## Outline for this Study



### PhosphoSens-Kinetic Assay Validation

### **Enzyme Source, Construct, and Lot Information:**

Invitrogen BRAF (Cat#/Lot#): PV3848/2243669P) amino acids 1-766 (end); GST-tagged

### **Reference Compound Information:**

GW5074 MedChemExpress (HY-10542)

### **Experiments to be run:**

**Enzyme Titration** 

Sensor Peptide K<sub>m</sub> Determination

ATP K<sub>m</sub> Determination

**DMSO Tolerance Test** 

Reference Compound IC<sub>50</sub> Determination at ATP K<sub>m</sub>

## **Enzyme Titration**

### Reaction Conditions and Set Up



### **Reaction Conditions:**

54 mM HEPES, pH 7.5

1mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

15 μM AQT0682

0, 0.08, 0.16, 0.33, 0.65, 1.3, 2.5, 5, 10, 20, 40, and 80 nM BRAF

#### **Reaction Set Up:**

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 or 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

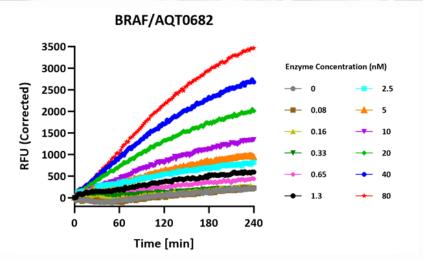
#### Notes:

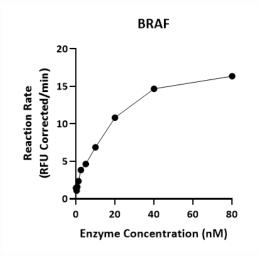
## **Enzyme Titration**

AssayQuant®

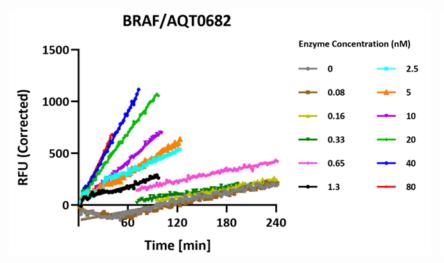
**Progress Curves** 

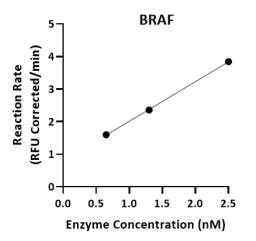
Complete Progress Curves





Linear Region of Progress Curves





Linear Range

## **Enzyme Titration**



### Reaction Rate Table

[Enzyme], nM	Normalized Reaction Rate (RFU/pmole/min)	Normalized Rate StndError (RFU/pmole/min)
0.08	896.25	24.95
0.16	463.44	12.79
0.31	175.00	5.10
0.63	126.90	2.37
1.3	90.65	3.87
2.5	76.86	2.02
5	46.35	0.73
10	34.42	0.48
20	27.08	0.27
40	18.34	0.20
80	10.22	0.30

The reaction is linear from 0.65 - 2.5 nM

## Sensor Peptide K<sub>m</sub> Determination



### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

1mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

 $0, 0.2, 0.4, 0.8, 1.6, 3.1, 6.3, 12.5, 25, 50, 100, or 200 \mu M AQT0682$ 

40 nM BRAF

### **Reaction Set Up:**

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

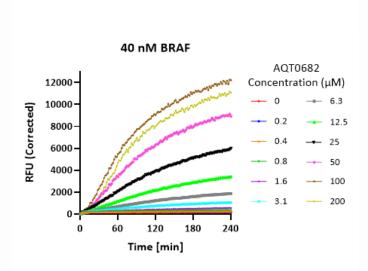
#### Notes:

## Sensor Peptide K<sub>m</sub> Determination

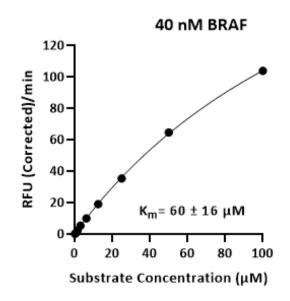


Titration Curves and K<sub>m</sub> Plot and Table

### Sensor Peptide Titration Curves



## Sensor Peptide K<sub>m</sub> Plot



Sensor Peptide  $K_m$  is 60  $\mu M$ 

## ATP K<sub>m</sub> Determination



### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

0, 2.0, 3.9, 7.8, 16, 31, 63, 125, 250, 500, 1000, and 2000 μM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

15 μM AQT0682

40 nM BRAF

### **Reaction Set Up:**

2 or 2.5 μL 10x Sensor Peptide

14 or 17.5 μL Reaction Mix with ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

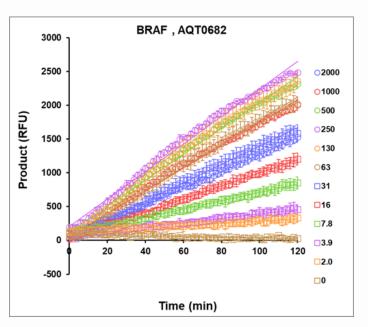
#### Notes:

## ATP K<sub>m</sub> Determination

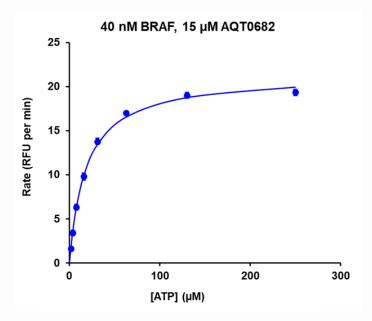


Titration Curves and K<sub>m</sub> Plot and Table

# ATP Titration Curves



### ATP K<sub>m</sub> Plot



ATP  $K_m$  is 19  $\mu M$ 

ATP K<sub>m</sub> Table

Parameter	Value	Approx SE
Vmax (RFU per min)	21.4	1.6
Vmax (RFU per pmol per min)	27	2
Km (μM)	18.5	3.6
R squared	0.998	

## **DMSO Tolerance Test**



### Reaction Conditions and Set Up

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

1 mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

0, .01, .02, .04, .08, .16, .31, .63, 1.3, 2.5, 5.0, and 10% DMSO

15 μM AQT0682

40 nM BRAF

### **Reaction Set Up:**

2 or 2.5 μL 10x DMSO dilutions

14 or 17.5 μL Reaction Mix with Sensor Peptide, ATP & DTT

4 or 5 μL 1x EDB or Kinase dilutions (5x in EDB)

20 or 25 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

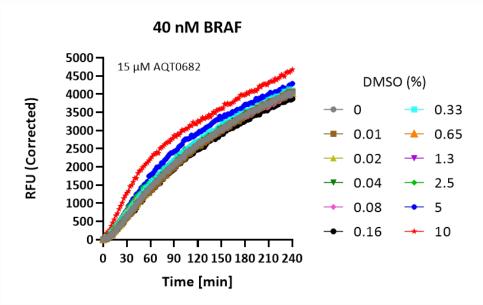
#### Notes:

## **DMSO Tolerance Test**

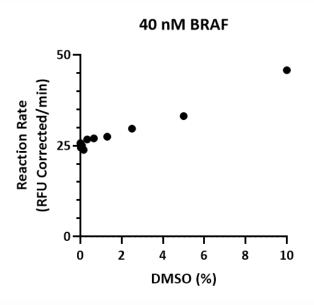


### Titration Curves and Inhibition Plot

## Complete Progress Curves



# Reaction Rate vs [DMSO] Plot



No change in enzyme activity out to 1% DMSO

## IC<sub>50</sub> Determination

# AssayQuant®

### Reaction Conditions and Set Up

### **Reaction Conditions:**

54 mM HEPES, pH 7.5

ATP at K<sub>m</sub>

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

1% DMSO

15 μM AQT0682

40 nM BRAF

0.1 mM GW5074 with 3-fold titration in 100% DMSO then diluted 10-fold into BSA (with a final concentration of 0.2 mg/ml) for a DMSO concentration of 10% before diluted 10-fold into reaction mixture with a final DMSO concentration of 1%

### **Reaction Set Up:**

16 μL Reaction Mix with Sensor Peptide and Inhibitor

 $4 \mu L$  1x EDB or Kinase dilutions (5x in EDB)

20 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Inhibitors are added via direct (0.4  $\mu$ L of 50X stock in 100% DMSO) or intermediate dilutions (2.0  $\mu$ L of 10X stock in 10% DMSO).

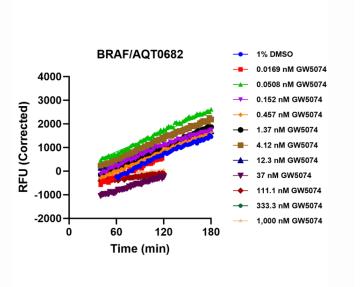
#### Notes:

## IC<sub>50</sub> Determination

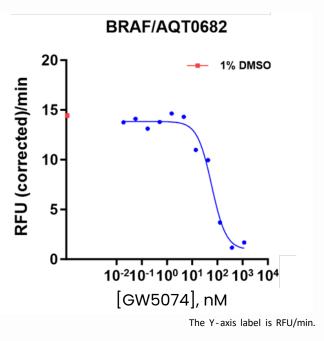


Progress Curves and IC<sub>50</sub> Curves and Table

# Linear Region of Progress Curves



### IC<sub>50</sub> Curve



IC<sub>50</sub> Table

log(inhibitor) vs. response Variable slope (four parameters)	
Best-fit values	
Bottom	1.167
Тор	14.02
LogIC50	1.717
HillSlope	-1.520
IC50	52.15
Span	12.85
Std. Error	
Bottom	0.8910
Тор	0.3904
LogIC50	0.08907
HillSlope	0.4038
Span	1.023
95% CI (asymptotic)	
Bottom	-0.9397 to 3.274
Тор	13.09 to 14.94
LogIC50	1.507 to 1.928
HillSlope	-2.475 to -0.5649
1C50	32.11 to 84.70
Span	10.43 to 15.27
Goodness of Fit	
Degrees of Freedom	7
R squared	0.9791

GW5074 IC<sub>50</sub> Determination at ATP K<sub>m</sub> is 52 nM

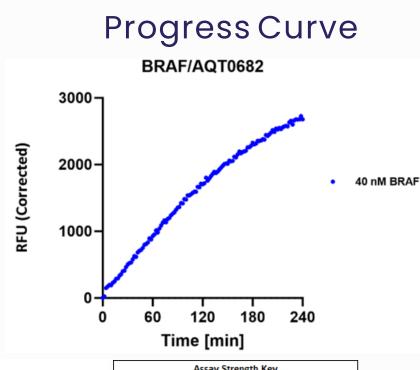
## Summary



### Assay Validation Results and Progress Curve and Assay Strength at 1 mM ATP

Experiment	Result
Enzyme Titration Linear Range	0.65 - 2.5 nM
Sensor Peptide K <sub>m</sub> Value	60 μΜ
ATP K <sub>m</sub> Value	19 μΜ
DMSO Tolerance (highest % recommended)	1 %
GW5074 IC <sub>50</sub> Determination at ATP K <sub>m</sub> is	52 nM

Kinase Name	Conc. (nM)	Sox-based Substrate Name	Normalized Reaction Rate (RFU/pmole/min)	Normalized Rate StndError (RFU/pmole/min)
BRAF	40	AQT0682	18.3	0.2



Assay Strength Key	
Very Strong	>1,000 (RFU/pmole/min)
Strong	300 to 999 (RFU/pmole/min)
Moderate	100 to 299 (RFU/pmole/min)
Weak	30 to 99 (RFU/pmole/min)

Under the conditions utilized for this experiment, the assay is Weak