

AQT0732 - MTOR Assay Validation

PhosphoSens®-Kinetic Assay Format

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Outline for this Study



PhosphoSens-Kinetic Assay Validation

Enzyme Source, Construct, and Lot Information:

Carna mTOR/MLST8 (Cat#/Lot#: 11-431/21CBS-0121B) amino acids 1362-2549(end); N-terminal DYKDDDDK tag

Reference Compound Information:

Wortmannin MedChemExpress (Cat#: HY-10197)

Experiments to be run:

Enzyme Titration

Sensor Peptide K_m Determination

ATP K_m Determination

DMSO Tolerance Test

Reference Compound IC₅₀ Determination at ATPK_m

Enzyme Titration

Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5 1mM ATP 1.2 mM DTT 0.012% Brij-35 1% glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl₂ 20 μM AQT0732

0, 0.040, 0.080, 0.16, 0.31, 0.63, 1.3, 2.5, 5.0, 10, 20, and 40 $\,$ nM mTOR/MLST8

Reaction Set Up:2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 or 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Final reaction volume

Notes:

20 or 25 µL





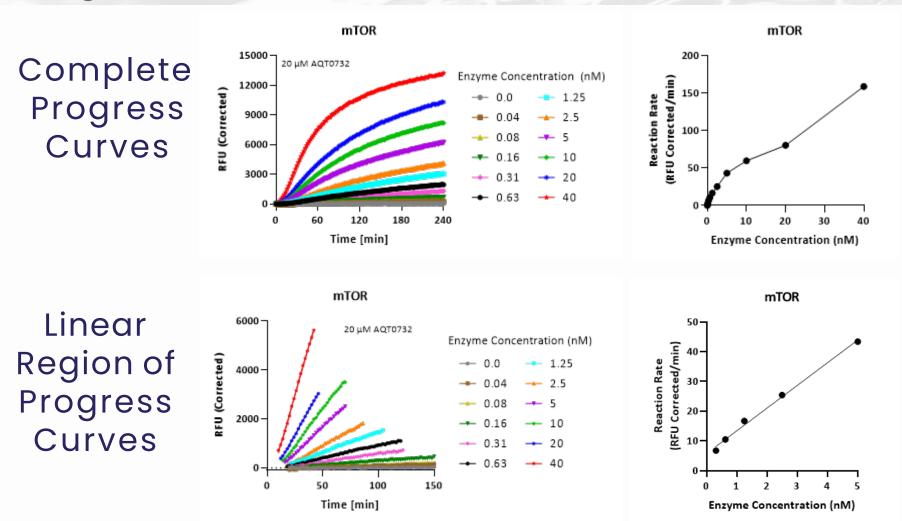
Enzyme Titration



Linear

Range

Progress Curves



Enzyme Titration

AssayQuant®

Reaction Rate Table

Enzyme Conc. (nM)	Reaction Rate (RFU/min)	Normalized Reaction Rate (RFU/pmole/min)		
0.04	1.4	1788		
0.08	1.9	1240		
0.16	3.4	1089		
0.31	6.8	1082		
0.63	11	842		
1.3	17	669		
2.5	25	508		
5.0	43	434		
10	60	299		
20	81	201		
40	159	199		

The reaction is linear from 0.63-5.0 nM

Sensor Peptide K_m Determination



Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5 1mM ATP 1.2 mM DTT 0.012% Brij-35 1%glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl₂ 0, 0.20, 0.39, 0.78, 1.6, 3.1, 6.3, 13, 25, 50, 100, and 200 μM AQT0732 8 nM mTOR/MLST8

Reaction Set Up:2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

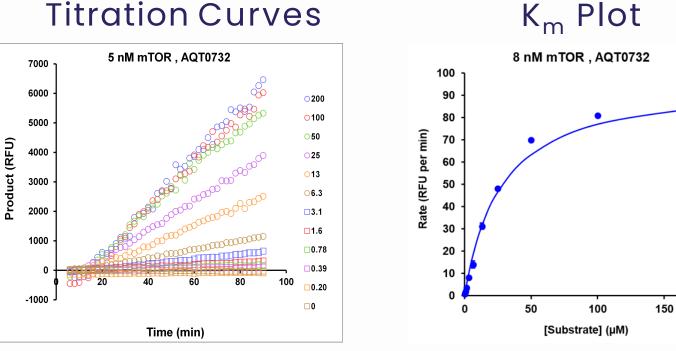
Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Notes:

Sensor Peptide K_m Determination

Titration Curves and K_m Plot and Table

Sensor Peptide Titration Curves



Sensor Peptide K_m Table

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Parameter	Value	Approx SE
Vmax (RFU per min)	98.5	3.3
Vmax (RFU per pmol per min)	616	21
Km (μM)	27.8	0.9
R squared	0.992	

Sensor Peptide K_m is 28 µM

200

Sensor Peptide



ATP K_m Determination



Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5	
0, 2.0, 3.9, 7.8, 16, 31, 63, 125, 250, 500, 1000, and 2000 μM	Л АТР
1.2 mM DTT	
0.012% Brij-35	
1%glycerol	
0.2 mg/ml BSA	
0.55 mM EGTA	
10 mM MgCl ₂	
20 μM AQT0732	
8 nM mTOR/MLST8	

Reaction Set Up:
2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Notes:

ATP K_m Determination

Titration Curves and K_m Plot and Table

2000

01000

0500

250

0130
063

31

□7.8 □3.9

2.0

□0

100

ATP Titration Curves

8 nM mTOR , AQT0732

12000

10000

8000

6000

4000

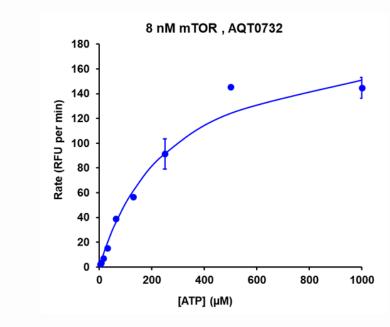
2000

-2000

20

1000-**40**0000

Product (RFU)



ATP K_m Plot

ATP K_m Table

Parameter	Value	Approx SE
Vmax (RFU per min)	192.8	3.7
Vmax (RFU per pmol per min)	1205	23
Km (μM)	276.6	23.4
Rsquared	0.982	

ATP K_m is 277 µM

Time (min)

60 80





DMSO Tolerance Test



Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5 1 mM ATP 1.2 mM DTT 0.012% Brij-35 1% glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl₂ 0, .01, .02, .04, .08, .16, .31, .63, 1.3, 2.5, 5.0, and 10% DMSO 40 μM AQT0732 5 nM mTOR/MLST8

Reaction Set Up:
2 or 2.5 μL10x DMSO dilutions14 or 17.5 μLReaction Mix with Sensor Peptide, ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

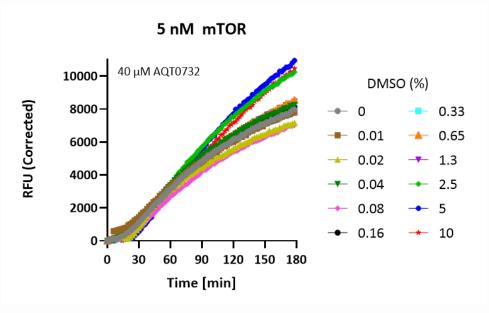
Notes:

DMSO Tolerance Test

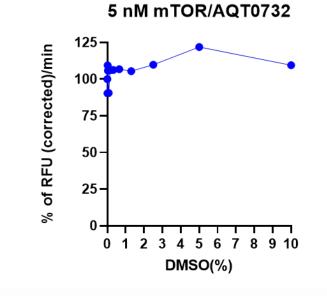


Titration Curves and Inhibition Plot

Complete Progress Curves



Reaction Rate vs [DMSO] Plot



No change in enzyme activity out to 3% DMSO

IC₅₀ Determination

Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5 ATP at K_m 1.2 mM DTT 0.012% Brij-35 1%glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl₂ 2% DMSO

40 µM AQT0732

5 nM mTOR/MLST8

0, 0.17, 0.51, 1.5, 4.6, 14, 41, 123, 370, 1,110, 3,330, and 10,000 nM Wortmannin

Reaction Set Up:

- 16 μLReaction Mix with Sensor Peptide and Inhibitor4 μL1 522 μ/μ
- $4 \mu L$ 1x EDB or Kinase dilutions (5x in EDB)
- 20 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Inhibitors are added via direct (0.4 μ L of 50X stock in 100% DMSO) or intermediate dilutions (2.0 μ L of 10X stock in 10% DMSO).

Notes:



IC₅₀ Determination



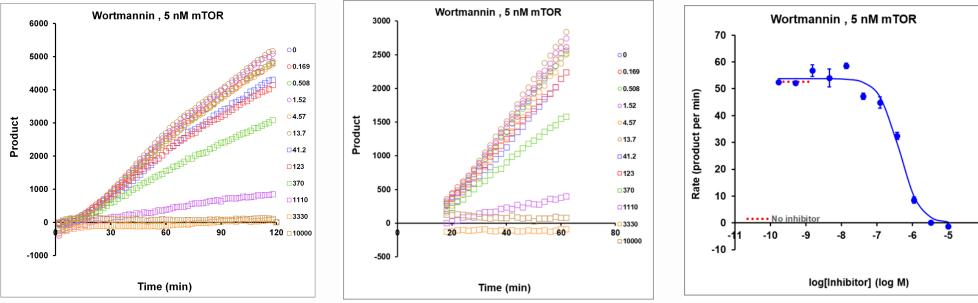
Progress Curves and IC₅₀ Curves and Table

Inhibitor Titration Progress Curves

Linear Region of Progress Curves

IC₅₀ Curve

IC₅₀ Table



Parameter	Value		
Bottom	0.0		
Тор	53.7		
log IC50	-6.37		
IC50 (nM)	425.93		
Ki (nM)	212.97		
Slope	-1.544		
R squared	0.985		
IC50 approx SE (nM)	17.80		
50% inhibition (nM)	425.93		

The Y-axis label is RFU/min.

Wortmannin IC₅₀ at ATP K_m is 426 nM

Summary



Assay Validation Results and Progress Curve and Assay Strength at 1 mM ATP

Experiment	Result	Progress Curve		
Enzyme Titration Linear Range	0.63-5.0 nM	mTOR/AQT0732		
Sensor Peptide K _m Value	28 μΜ			
ATP K _m Value	277 μΜ	-indig 1000 - → 5 nM		
DMSOTolerance (highest % recommended)	1	2000-		
Wortmannin $\rm IC_{50}$ Determination at ATP $\rm K_m$	426 nM			
		Time [min]		

		Sox-based		Normalized Rate	Assa	y Strength Key
Kinase Name	Conc. (nM)	Substrate	Normalized Reaction	Stnd Error	Very Strong	> 1,000 (RFU/pmole/min)
Kinase Name			Rate (RFU/pmole/min)		Strong	300 to 999 (RFU/pmole/min)
		Name		(RFU/pmole/min)	Moderate	100 to 299 (RFU/pmole/min)
mTOR	5.0	AQT0732	434	3	Weak	30 to 99 (RFU/pmole/min)

Under the conditions utilized for this experiment, the assay is Strong