

# AQT0794 - FGFR1 Assay Validation

PhosphoSens®-Kinetic Assay Format

# **Outline for this Study**



PhosphoSens-Kinetic Assay Validation

#### **Enzyme Source, Construct, and Lot Information:**

Carna FGFR1 (Cat#/Lot#: 08-133/17CBS-0499B) amino acids 398-822(end); N-terminal GST tag

#### **Reference Compound Information:**

Staurosporine MedChemExpress (Cat#/Lot#: HY-15141/125391) CAS No.: 62996-74-1

#### **Experiments to be run:**

**Enzyme Titration** 

Sensor Peptide K<sub>m</sub> Determination

ATP K<sub>m</sub> Determination

**DMSO** Tolerance Test

Reference Compound IC<sub>50</sub> Determination at ATPK<sub>m</sub>

# **Enzyme Titration**

## AssayQuant® TECHNOLOGIES INC.

### **Reaction Conditions and Set Up**

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5 1mM ATP 1.2 mM DTT 0.012% Brij-35 1% glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl<sub>2</sub> 15 μM AQT0794

 $0.01,\,0.02,\,0.04,\,0.08,\,0.16,\,0.3125,\,0.625,\,1.25,\,2.5,\,5,\,10,\,and\,20\;nM$  Carna FGFR1

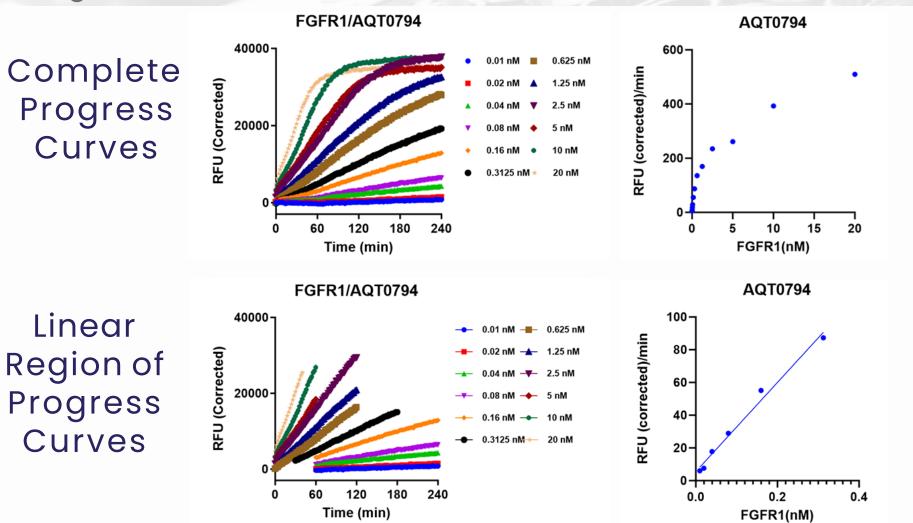
# Reaction Set Up:<br/>2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 or 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

#### Notes:

# **Enzyme Titration**

**Progress Curves** 





Linear

Range

# **Enzyme Titration**

# AssayQuant®

### **Reaction Rate Table**

Enzyme Conc. (nM)	Normalized Reaction Rate (RFU/pmole/min)	Normalized Rate Stnd Error (RFU/pmole/min)
0.01	30,235	646
0.02	19,158	311
0.04	44,525	454
0.08	36,150	244
0.16	34,450	164
0.3125	27,278	82
0.625	21,712	84
1.25	13,576	48
2.5	9,388	27
5	5,222	48
10	3,929	47
20	2,548	21

## The reaction is linear from 0.04 - 0.31 nM

# Sensor Peptide K<sub>m</sub> Determination



**Reaction Conditions and Set Up** 

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5 1mM ATP 1.2 mM DTT 0.012% Brij-35 1% glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl<sub>2</sub> 1, 2, 3, 5, 7, 10, 15, 20, 30, 50, 70, and 100 μM AQT0794 2 nM Carna FGFR1

# Reaction Set Up:<br/>2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

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#### Notes:

# **Sensor Peptide K**<sub>m</sub> Determination

Titration Curves and K<sub>m</sub> Plot and Table

Sensor Peptide Titration Curves

FGFR1/AQT0794

15000

10000

5000

10

0

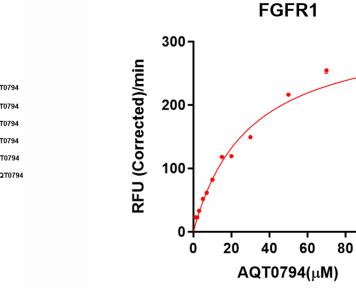
20 30

Time (min)

40

50

RFU (Corrected)



## Sensor Peptide K<sub>m</sub> Table

Michaelis-Menten	
Best-fit values	
Vmax	326.2
Km	29.43
Std. Error	
Vmax	22.56
Km	4.811
95% CI (profile likelihood)	
Vmax	281.8 to 384.6
Km	20.58 to 42.77
Goodness of Fit	
Degrees of Freedom	10
R squared	0.9771

## Sensor Peptide K<sub>m</sub> is 29 µM

Sensor Peptide

K<sub>m</sub> Plot

100



# **ATP K<sub>m</sub> Determination**



## **Reaction Conditions and Set Up**

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5
0, 2.0, 3.9, 7.8, 16, 31, 63, 125, 250, 500, 1000, and 2000 $\mu M$ ATP
1.2 mM DTT
0.012% Brij-35
1% glycerol
0.2 mg/ml BSA
0.55 mM EGTA
10 mM MgCl <sub>2</sub>
15 μM AQT0794
1.5 nM Carna FGFR1

# Reaction Set Up:<br/>2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

#### Notes:

# **ATP K<sub>m</sub> Determination**

Titration Curves and K<sub>m</sub> Plot and Table

ATP Titration Curves

FGFR1/AQT0794

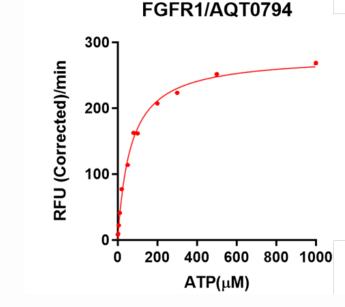
30

Time (min)

60

20000-

RFU (Corrected) - 0000 - 0000 - 0000 - 0000



ATP K<sub>m</sub> Plot

## ATP K<sub>m</sub> Table

Michaelis-Menten	
Best-fit values	
Vmax	280.1
Km	65.30
Std. Error	
Vmax	5.775
Km	4.824
95% CI (profile likelihood)	
Vmax	267.3 to 293.6
Km	54.97 to 77.30
Goodness of Fit	
Degrees of Freedom	10
R squared	0.9955

## ATP K<sub>m</sub> is 65 µM

#### How Can We Help? For technical questions, please reach out at hello@assayquant.com



# **DMSO Tolerance Test**

# AssayQuant<sup>®</sup>

### **Reaction Conditions and Set Up**

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5
1 mM ATP
1.2 mM DTT
0.012% Brij-35
1% glycerol
0.2 mg/ml BSA
0.55 mM EGTA
10 mM MgCl<sub>2</sub>
0, .01, .02, .04, .08, .16, .31, .63, 1.3, 2.5, 5.0, and 10% DMSO
15 μM AQT0794
0.5 nM Carna EGER1

# Reaction Set Up:<br/>2 or 2.5 μL10x DMSO dilutions14 or 17.5 μLReaction Mix with Sensor Peptide, ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

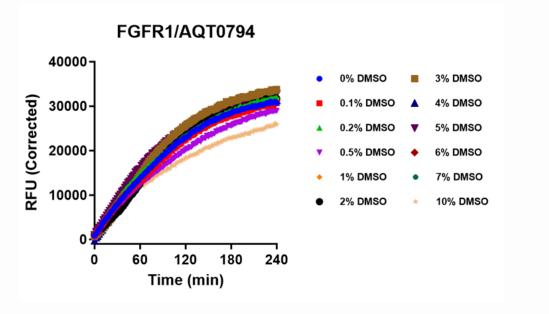
#### Notes:

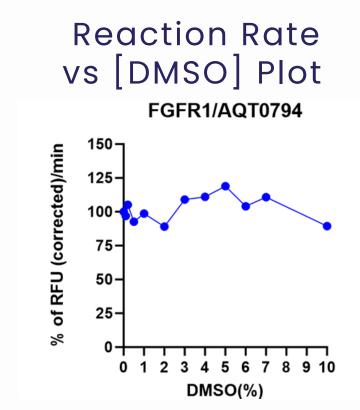
## **DMSO Tolerance Test**



**Titration Curves and Inhibition Plot** 

## Complete Progress Curves





No change in enzyme activity out to 1% DMSO

# IC<sub>50</sub> Determination

### **Reaction Conditions and Set Up**

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5

ATP at  $K_m$ 

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

1% DMSO

 $15 \, \mu M \, AQT0794$ 

2 nM Carna FGFR1

0.1 mM Staurosporine was serially diluted (3-fold, 11-point) in 100% DMSO. The series was then diluted 10-fold into BSA (with a final concentration of 0.2 mg/mL BSA in 10% DMSO) to prepare the 10x compound stocks.

#### **Reaction Set Up:**

16 µL	Reaction Mix with Sensor Peptide and Inhibitor
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- $4 \mu L$  1x EDB or Kinase dilutions (5x in EDB)
- 20 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Inhibitors are added via direct (0.4  $\mu L$  of 50X stock in 100% DMSO) or intermediate dilutions (2.0  $\mu L$  of 10X stock in 10% DMSO).

#### Notes:



# IC<sub>50</sub> Determination

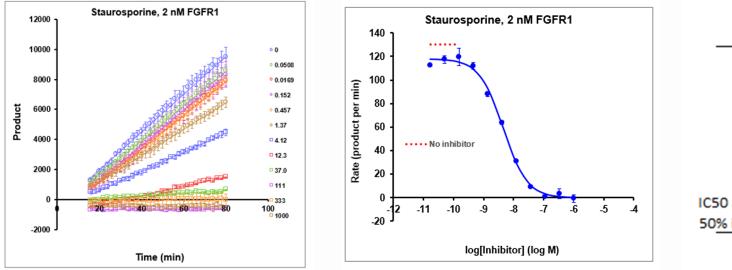


Progress Curves and IC<sub>50</sub> Curves and Table

## Linear Region of Progress Curves

IC<sub>50</sub> Curve





Parameter	Value			
Bottom	0.0			
Тор	117.8			
log IC50	-8.34			
IC50 (nM)	4.54			
Ki (nM)	2.27			
Slope	-1.099			
R squared	0.997			
C50 approx SE (nM)	0.00			
0% inhibition (nM)	4.54			

The Y-axis label is RFU/min.

Staurosporine IC<sub>50</sub> Determination at ATP K<sub>m</sub> is 4.5 nM

# Summary



## Assay Validation Results and Progress Curve and Assay Strength at 1 mM ATP

Experiment			Result				<b>Progress</b> Curve					
Enzyme Titration Linear Range			0.04 - 0.31 nM			AQT0794						
Sensor Peptide K <sub>m</sub> Value			29 µM			ted)	20000-					
ATP K <sub>m</sub> Value			65 μM			RFU (Corrected)	20000		/ · ·		0.625 nM FGFR1	
DMSOTolerance (highest % recommended)			1			RFU (	10000-					
Staurosporine IC50 Determination at ATP Km is			4.5 nM				o	60 120	180	240		
								Time (n	nin)			
Kinase Name	Conc. (nM) Sox-based Substrate Name		Normalized Reaction Rate	Normalized Rate StndError	Maximum Signal:Bkgd			Very Strong		RFU/pmole/min)		
		Substrate Mame	(RFU/pmole/min)	(RFU/pmole/min)	(S/B) Kineti	ic		Strong		(RFU/pmole/min		

84

## Under the conditions utilized for this experiment, the assay is Very Strong

21,712

0.625

AQT0794

FGFR1

100 to 299 (RFU/pmole/min)

30 to 99 (RFU/pmole/min)

Moderate

Weak

2.7