

AQT0794 - FGFR1 Assay Validation

PhosphoSens®-Kinetic Assay Format

Outline for this Study



PhosphoSens-Kinetic Assay Validation

Enzyme Source, Construct, and Lot Information:

Carna FGFR1 (Cat#/Lot#: 08-133/17CBS-0499B) amino acids 398-822(end); N-terminal GST tag

Reference Compound Information:

Staurosporine MedChemExpress (Cat#/Lot#: HY-15141/125391) CAS No.: 62996-74-1

Experiments to be run:

Enzyme Titration

Sensor Peptide K_m Determination

ATP K_m Determination

DMSO Tolerance Test

Reference Compound IC₅₀ Determination at ATPK_m

Enzyme Titration

AssayQuant® TECHNOLOGIES INC.

Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5 1mM ATP 1.2 mM DTT 0.012% Brij-35 1% glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl₂ 15 μM AQT0794

 $0.01,\,0.02,\,0.04,\,0.08,\,0.16,\,0.3125,\,0.625,\,1.25,\,2.5,\,5,\,10,\,and\,20\;nM$ Carna FGFR1

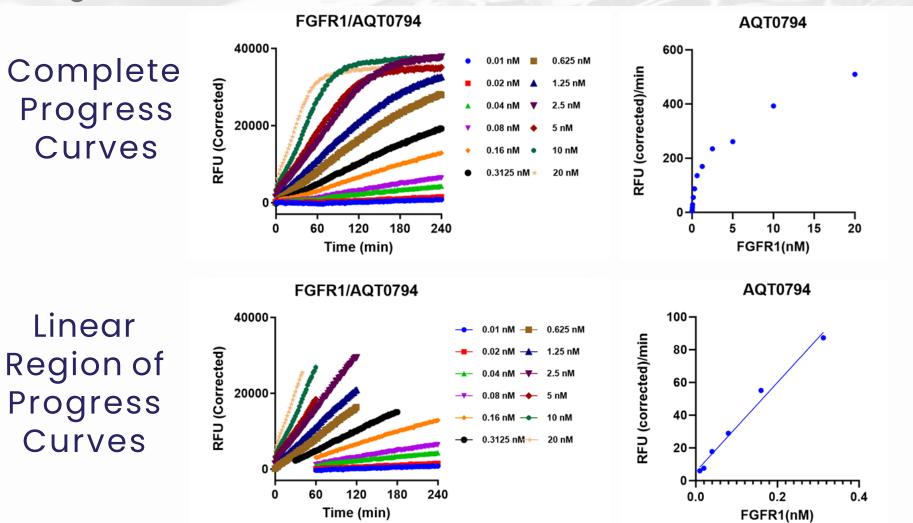
Reaction Set Up:
2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 or 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Notes:

Enzyme Titration

Progress Curves





Linear

Range

Enzyme Titration

AssayQuant®

Reaction Rate Table

Enzyme Conc. (nM)	Normalized Reaction Rate (RFU/pmole/min)	Normalized Rate Stnd Error (RFU/pmole/min)
0.01	30,235	646
0.02	19,158	311
0.04	44,525	454
0.08	36,150	244
0.16	34,450	164
0.3125	27,278	82
0.625	21,712	84
1.25	13,576	48
2.5	9,388	27
5	5,222	48
10	3,929	47
20	2,548	21

The reaction is linear from 0.04 - 0.31 nM

Sensor Peptide K_m Determination



Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5 1mM ATP 1.2 mM DTT 0.012% Brij-35 1% glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl₂ 1, 2, 3, 5, 7, 10, 15, 20, 30, 50, 70, and 100 μM AQT0794 2 nM Carna FGFR1

Reaction Set Up:
2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Notes:

Sensor Peptide K_m Determination

Titration Curves and K_m Plot and Table

Sensor Peptide Titration Curves

FGFR1/AQT0794

15000

10000

5000

10

0

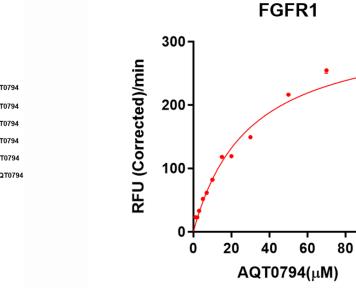
20 30

Time (min)

40

50

RFU (Corrected)



Sensor Peptide K_m Table

Michaelis-Menten	
Best-fit values	
Vmax	326.2
Km	29.43
Std. Error	
Vmax	22.56
Km	4.811
95% CI (profile likelihood)	
Vmax	281.8 to 384.6
Km	20.58 to 42.77
Goodness of Fit	
Degrees of Freedom	10
R squared	0.9771

Sensor Peptide K_m is 29 µM

Sensor Peptide

K_m Plot

100



ATP K_m Determination



Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5
0, 2.0, 3.9, 7.8, 16, 31, 63, 125, 250, 500, 1000, and 2000 μM ATP
1.2 mM DTT
0.012% Brij-35
1% glycerol
0.2 mg/ml BSA
0.55 mM EGTA
10 mM MgCl ₂
15 μM AQT0794
1.5 nM Carna FGFR1

Reaction Set Up:
2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Notes:

ATP K_m Determination

Titration Curves and K_m Plot and Table

ATP Titration Curves

FGFR1/AQT0794

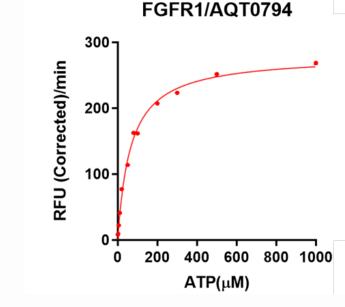
30

Time (min)

60

20000-

RFU (Corrected) - 0000 - 0000 - 0000 - 0000



ATP K_m Plot

ATP K_m Table

Michaelis-Menten	
Best-fit values	
Vmax	280.1
Km	65.30
Std. Error	
Vmax	5.775
Km	4.824
95% CI (profile likelihood)	
Vmax	267.3 to 293.6
Km	54.97 to 77.30
Goodness of Fit	
Degrees of Freedom	10
R squared	0.9955

ATP K_m is 65 µM

How Can We Help? For technical questions, please reach out at hello@assayquant.com



DMSO Tolerance Test

AssayQuant[®]

Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5
1 mM ATP
1.2 mM DTT
0.012% Brij-35
1% glycerol
0.2 mg/ml BSA
0.55 mM EGTA
10 mM MgCl₂
0, .01, .02, .04, .08, .16, .31, .63, 1.3, 2.5, 5.0, and 10% DMSO
15 μM AQT0794
0.5 nM Carna EGER1

Reaction Set Up:
2 or 2.5 μL10x DMSO dilutions14 or 17.5 μLReaction Mix with Sensor Peptide, ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

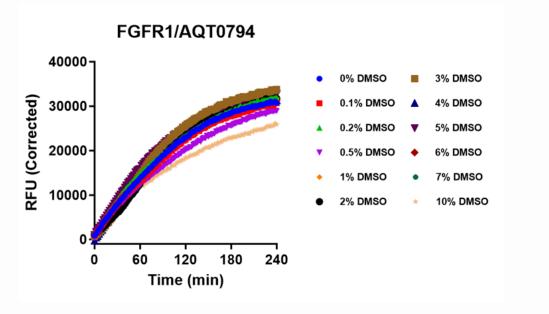
Notes:

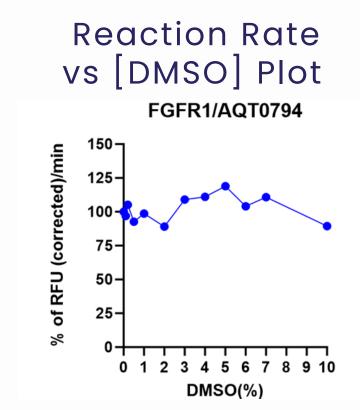
DMSO Tolerance Test



Titration Curves and Inhibition Plot

Complete Progress Curves





No change in enzyme activity out to 1% DMSO

IC₅₀ Determination

Reaction Conditions and Set Up

Reaction Conditions:

54 mM HEPES, pH 7.5

ATP at K_m

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl₂

1% DMSO

 $15 \, \mu M \, AQT0794$

2 nM Carna FGFR1

0.1 mM Staurosporine was serially diluted (3-fold, 11-point) in 100% DMSO. The series was then diluted 10-fold into BSA (with a final concentration of 0.2 mg/mL BSA in 10% DMSO) to prepare the 10x compound stocks.

Reaction Set Up:

16 µL	Reaction Mix with Sensor Peptide and Inhibitor
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- $4 \mu L$ 1x EDB or Kinase dilutions (5x in EDB)
- 20 μL Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 μ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μ L final well volumeafter sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Inhibitors are added via direct (0.4 μL of 50X stock in 100% DMSO) or intermediate dilutions (2.0 μL of 10X stock in 10% DMSO).

Notes:



IC₅₀ Determination

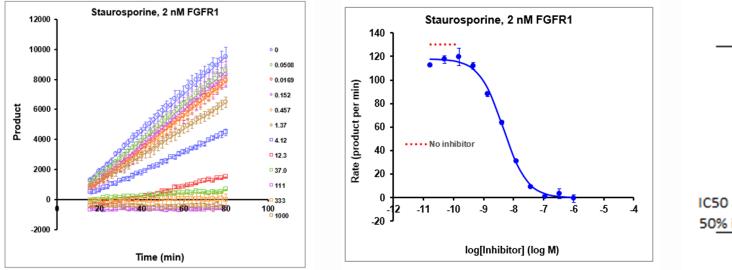


Progress Curves and IC₅₀ Curves and Table

Linear Region of Progress Curves

IC₅₀ Curve





Parameter	Value			
Bottom	0.0			
Тор	117.8			
log IC50	-8.34			
IC50 (nM)	4.54			
Ki (nM)	2.27			
Slope	-1.099			
R squared	0.997			
C50 approx SE (nM)	0.00			
0% inhibition (nM)	4.54			

The Y-axis label is RFU/min.

Staurosporine IC₅₀ Determination at ATP K_m is 4.5 nM

Summary



Assay Validation Results and Progress Curve and Assay Strength at 1 mM ATP

Experiment			Result				Progress Curve					
Enzyme Titration Linear Range			0.04 - 0.31 nM			AQT0794						
Sensor Peptide K _m Value			29 µM			ted)	20000-					
ATP K _m Value			65 μM			RFU (Corrected)	20000		/ · ·		0.625 nM FGFR1	
DMSOTolerance (highest % recommended)			1			RFU (10000-					
Staurosporine IC50 Determination at ATP Km is			4.5 nM				o	60 120	180	240		
								Time (n	nin)			
Kinase Name	Conc. (nM) Sox-based Substrate Name		Normalized Reaction Rate	Normalized Rate StndError	Maximum Signal:Bkgd			Very Strong		RFU/pmole/min)		
		Substrate Mame	(RFU/pmole/min)	(RFU/pmole/min)	(S/B) Kineti	ic		Strong		(RFU/pmole/min		

84

Under the conditions utilized for this experiment, the assay is Very Strong

21,712

0.625

AQT0794

FGFR1

100 to 299 (RFU/pmole/min)

30 to 99 (RFU/pmole/min)

Moderate

Weak

2.7