

# AQT0887 – NEK7 Assay Validation

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*PhosphoSens<sup>®</sup>*-Kinetic Assay Format

# Outline for this Study

## PhosphoSens<sup>®</sup>-Kinetic Assay Validation



### **Enzyme Source, Construct, and Lot Information:**

Carna NEK7 (05-131/13CBS-0250F) amino acid full length; N-terminal GST tag

### **Reference Compound Information:**

Staurosporine      MedChemExpress (Cat. HY-15141)

### **Experiments to be run:**

**Enzyme Titration**

**Sensor Peptide  $K_M$  Determination**

**ATP  $K_M$  Determination**

**DMSO Tolerance Test**

**Reference Compound  $IC_{50}$  Determination at ATP  $K_M$**

# Enzyme Titration

## Reaction Conditions and Set Up

### Reaction Conditions:

54 mM HEPES, pH 7.5

1 mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

15 uM AQT0887

0.1, 0.2, 0.4, 0.6, 0.8, 1, 2, 3, 5, 10, 15, and 20 nM NEK7

### Reaction Set Up:

2 or 2.5  $\mu$ L

10x Sensor Peptide

14 or 17.5  $\mu$ L

Reaction Mix with ATP & DTT

4 or 5  $\mu$ L

1x EDB or Kinase dilutions (5x in EDB)

20 or 25  $\mu$ L

Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 or 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

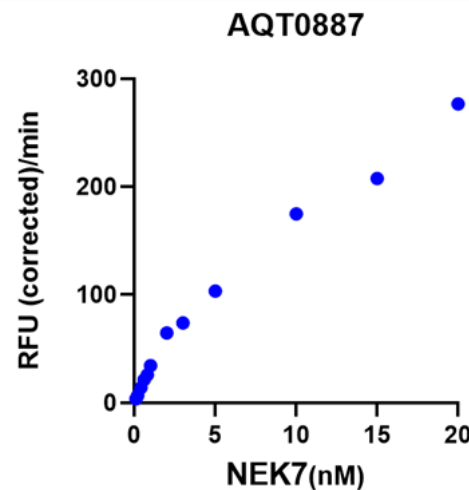
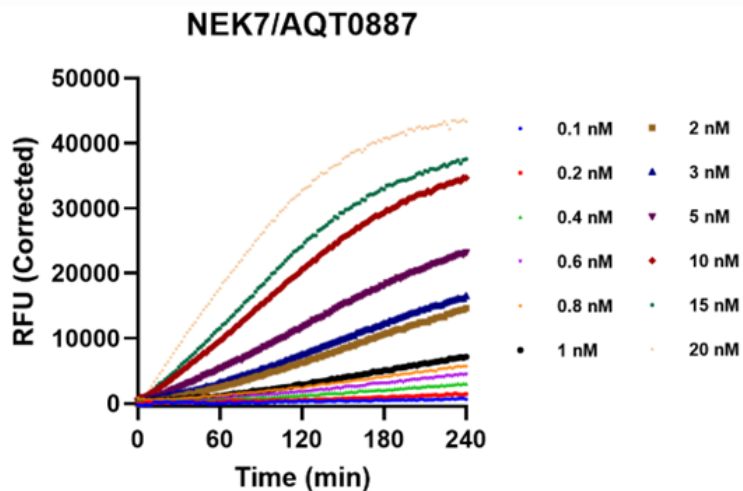
### Notes:

Enzyme Dilution Buffer (EDB): 20 mM HEPES, pH 7.5, 0.01% Brij-35, 5% Glycerol, 0.5 mM EGTA, 1 mM DTT, 1 mg/ml Bovine Serum Albumin.

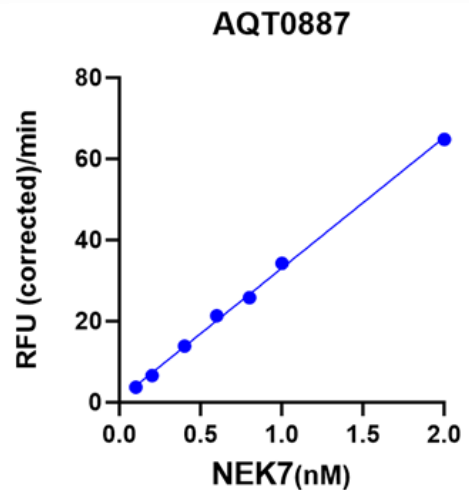
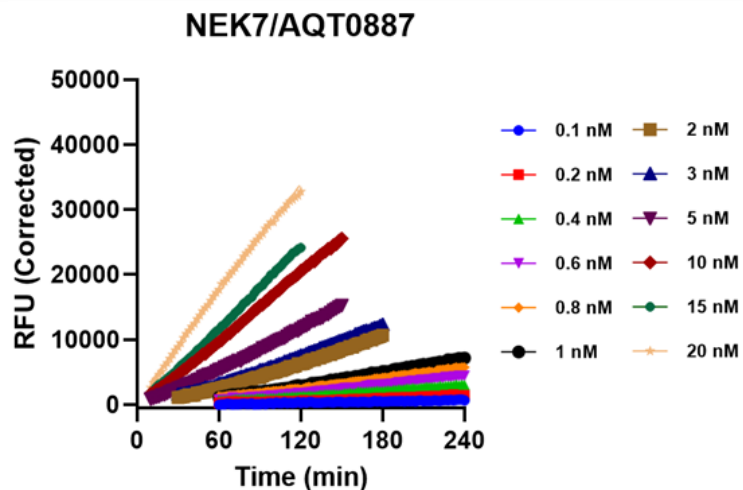
# Enzyme Titration

## Progress Curves

Complete  
Progress  
Curves



Linear  
Region of  
Progress  
Curves



Linear  
Range

# Enzyme Titration

## Reaction Rate Table

Enzyme Conc. (nM)	Normalized Reaction Rate (RFU/pmole/min)	Normalized Rate Stnd Error (RFU/pmole/min)
0.1	1,838	55
0.2	1,635	37
0.4	1,725	19
0.6	1,773	15
0.8	1,611	12
1	1,713	10
2	1,619	13
3	1,231	8
5	1,033	6
10	875	3
15	692	3
20	691	5

The reaction is linear from 0.1 – 2.0 nM

# Sensor Peptide $K_M$ Determination



## Reaction Conditions and Set Up

### Reaction Conditions:

54 mM HEPES, pH 7.5

1 mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM  $MgCl_2$

1, 2, 3, 5, 7, 10, 15, 20, 30, 50, 70, & 100  $\mu M$  AQT0887

10 nM NEK7

### Reaction Set Up:

2 or 2.5  $\mu L$

10x Sensor Peptide

14 or 17.5  $\mu L$

Reaction Mix with ATP & DTT

4 or 5  $\mu L$

1x EDB or Kinase dilutions (5x in EDB)

20 or 25  $\mu L$

Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu L$  final well volume or in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu L$  final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

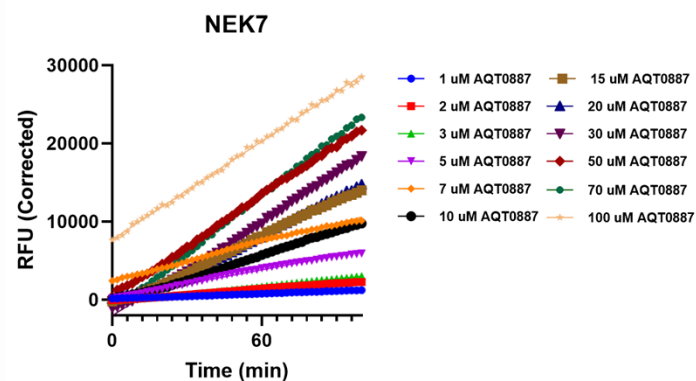
### Notes:

Enzyme Dilution Buffer (EDB): 20 mM HEPES, pH 7.5, 0.01% Brij-35, 5% Glycerol, 0.5 mM EGTA, 1 mM DTT, 1 mg/ml Bovine Serum Albumin.

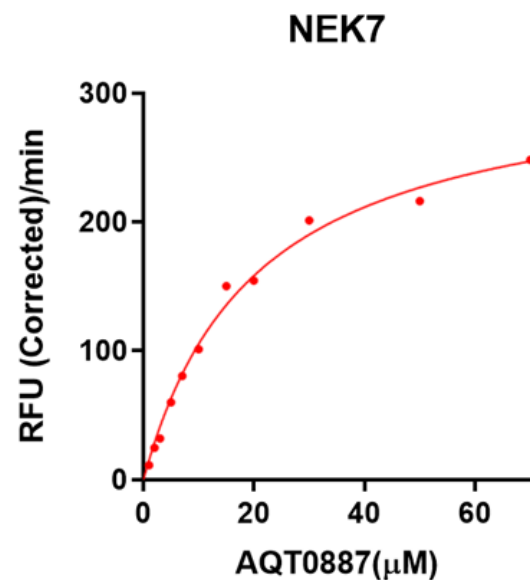
# Sensor Peptide $K_M$ Determination

Titration Curves and  $K_M$  Plot and Table

## Sensor Peptide Titration Curves



## Sensor Peptide $K_M$ Plot



## Sensor Peptide $K_M$ Table

Michaelis-Menten	
Best-fit values	
Vmax	318.5
Km	20.27
Std. Error	
Vmax	14.06
Km	2.107
95% CI (profile likelihood)	
Vmax	289.8 to 352.6
Km	16.16 to 25.56
Goodness of Fit	
Degrees of Freedom	9
R squared	0.9912

Sensor Peptide  $K_M$  is 20  $\mu$ M



# ATP $K_M$ Determination

## Reaction Conditions and Set Up

### Reaction Conditions:

54 mM HEPES, pH 7.5

0, 2.0, 3.9, 7.8, 16, 31, 63, 125, 250, 500, 1000, and 2000  $\mu$ M ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM  $MgCl_2$

15  $\mu$ M AQT0887

10 nM NEK7

### Reaction Set Up:

2 or 2.5  $\mu$ L

10x Sensor Peptide

14 or 17.5  $\mu$ L

Reaction Mix with ATP & DTT

4 or 5  $\mu$ L

1x EDB or Kinase dilutions (5x in EDB)

20 or 25  $\mu$ L

Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

### Notes:

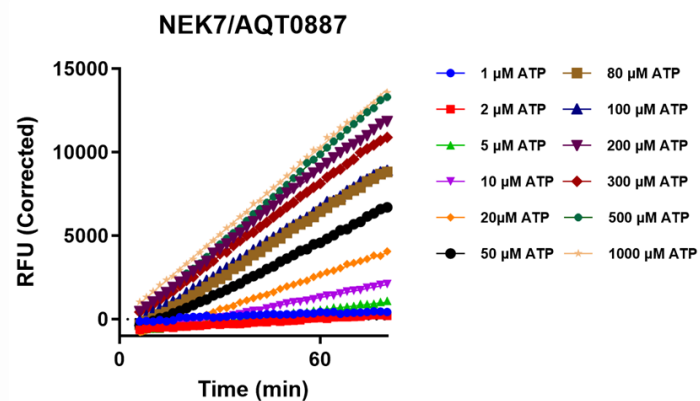
Enzyme Dilution Buffer (EDB): 20 mM HEPES, pH 7.5, 0.01% Brij-35, 5% Glycerol, 0.5 mM EGTA, 1 mM DTT, 1 mg/ml Bovine Serum Albumin.



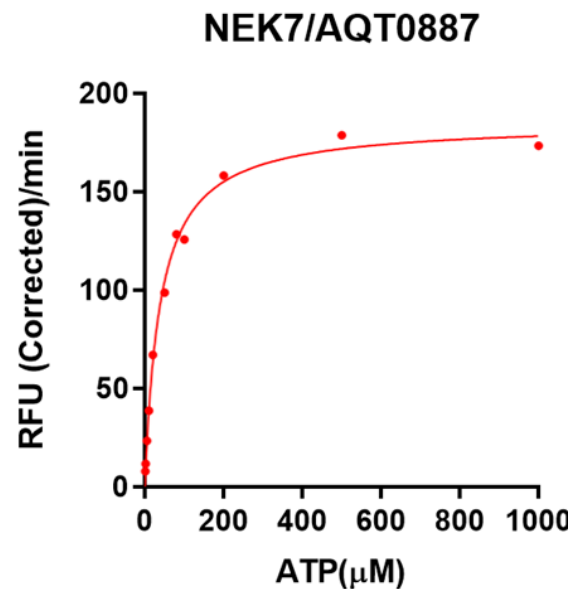
# ATP $K_M$ Determination

Titration Curves and  $K_M$  Plot and Table

## ATP Titration Curves



## ATP $K_M$ Plot



## ATP $K_M$ Table

Michaelis-Menten	
Best-fit values	
Vmax	185.0
Km	38.83
Std. Error	
Vmax	3.803
Km	3.090
95% CI (profile likelihood)	
Vmax	176.5 to 193.9
Km	32.24 to 46.53
Goodness of Fit	
Degrees of Freedom	9
R squared	0.9947

ATP  $K_M$  is 39  $\mu$ M

# DMSO Tolerance Test



## Reaction Conditions and Set Up

### Reaction Conditions:

54 mM HEPES, pH 7.5

1 mM ATP

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

0, .01, .02, .04, .08, .16, .31, .63, 1.3, 2.5, 5.0, and 10% DMSO

15 uM AQT0887

10 nM NEK7

### Reaction Set Up:

2 or 2.5  $\mu$ L

10x DMSO dilutions

14 or 17.5  $\mu$ L

Reaction Mix with Sensor Peptide, ATP & DTT

4 or 5  $\mu$ L

1x EDB or Kinase dilutions (5x in EDB)

20 or 25  $\mu$ L

Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

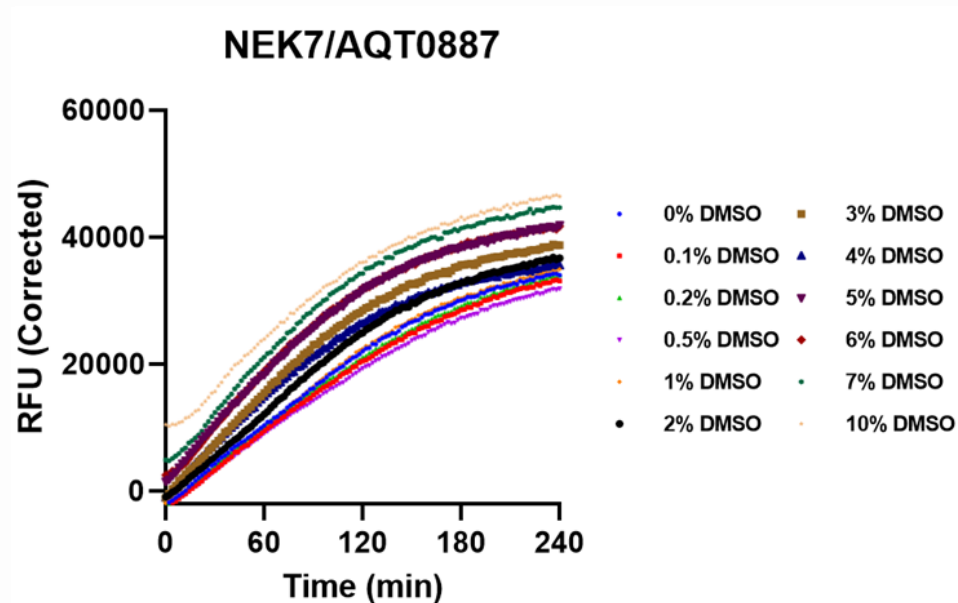
### Notes:

Enzyme Dilution Buffer (EDB): 20 mM HEPES, pH 7.5, 0.01% Brij-35, 5% Glycerol, 0.5 mM EGTA, 1 mM DTT, 1 mg/ml Bovine Serum Albumin.

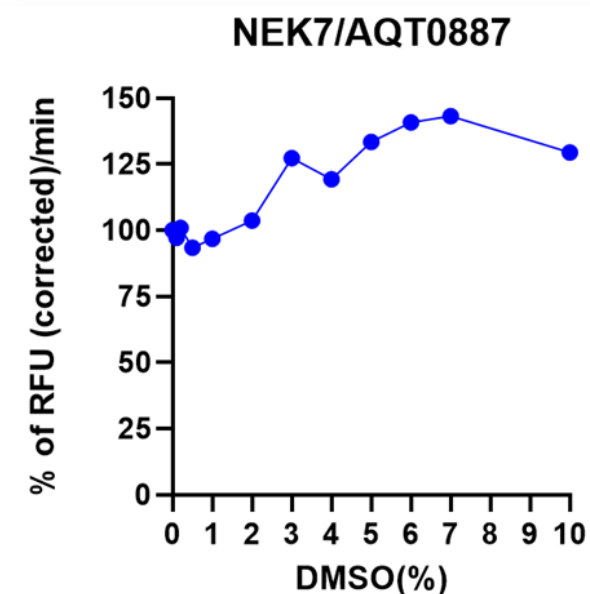
# DMSO Tolerance Test

## Titration Curves and Inhibition Plot

### Complete Progress Curves



### Reaction Rate vs [DMSO] Plot



There is no significant inhibitory effect up to 2% DMSO

# IC<sub>50</sub> Determination

## Reaction Conditions and Set Up

### Reaction Conditions:

54 mM HEPES, pH 7.5

ATP at K<sub>M</sub>

1.2 mM DTT

0.012% Brij-35

1% glycerol

0.2 mg/ml BSA

0.55 mM EGTA

10 mM MgCl<sub>2</sub>

1% DMSO

15 uM AQT0887

10 nM NEK7

5 mM Staurosporine with 3-fold titration in 100% DMSO then diluted 10-fold into BSA (with a final concentration of 0.2 mg/ml) for a DMSO concentration of 10% before diluted 10-fold into reaction mixture with a final DMSO concentration of 1%

### Reaction Set Up:

16 μL            Reaction Mix with Sensor Peptide and Inhibitor

4 μL            1x EDB or Kinase dilutions (5x in EDB)

20 μL            Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 μL final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 μL final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Inhibitors are added via direct (0.4 μL of 50X stock in 100% DMSO) or intermediate dilutions (2.0 μL of 10X stock in 10% DMSO).

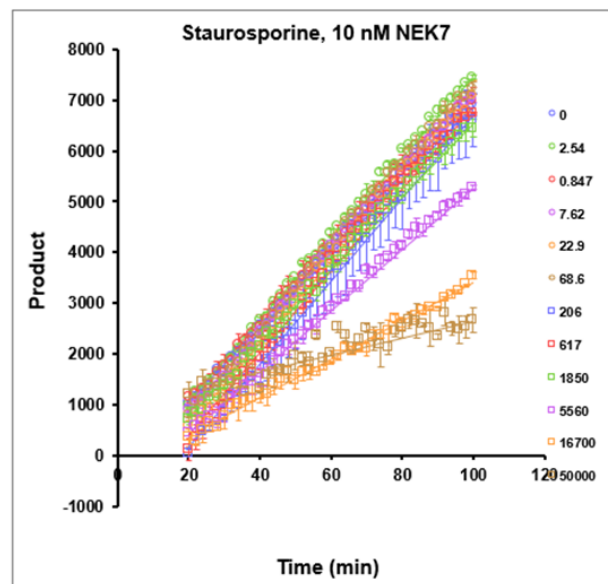
### Notes:

Enzyme Dilution Buffer (EDB): 20 mM HEPES, pH 7.5, 0.01% Brij-35, 5% Glycerol, 0.5 mM EGTA, 1 mM DTT, 1 mg/ml Bovine Serum Albumin.

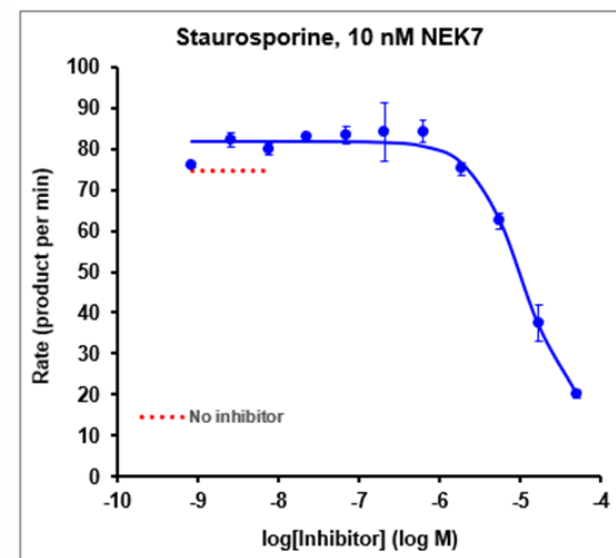
# IC<sub>50</sub> Determination

## Progress Curves and IC<sub>50</sub> Curves and Table

### Linear Region of Progress Curves



### IC<sub>50</sub> Curve



### IC<sub>50</sub> Table

Parameter	Value
Bottom	13.4
Top	81.9
log IC50	-4.97
IC50 (nM)	10709.05
Ki (nM)	5354.53
Slope	-1.425
R squared	0.986
IC50 approx SE (nM)	446.42
50% inhibition (nM)	14130.06

The Y-axis label is RFU/min.

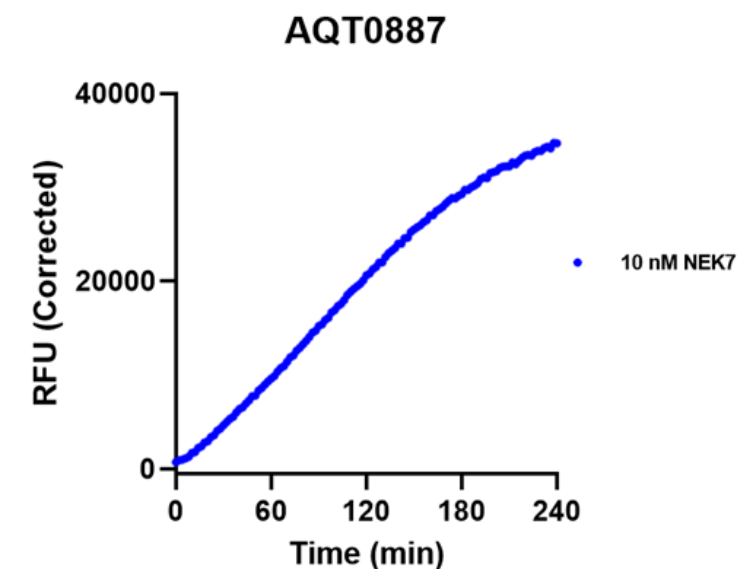
Staurosporine IC<sub>50</sub> at ATP K<sub>M</sub> is 10700 nM

# Summary

## Assay Validation Results and Progress Curve and Assay Strength at 1 mM ATP

Experiment	Result
Enzyme Titration Linear From	0.1 - 2.0 nM
Sensor Peptide $K_M$ Determination	20 $\mu$ M
ATP $K_M$ Determination	39 $\mu$ M
DMSO Tolerance Test	2%
Staurosporine IC50 Determination	10700 nM

### Progress Curve



Kinase Name	Conc. (nM)	Sox-based Substrate Name	Normalized Reaction Rate (RFU/pmole/mi)	Normalized Rate StdError (RFU/pmole/mi)
NEK7	10	AQT0887	875	3

Assay Strength Key	
Very Strong	> 1,000 (RFU/pmole/min)
Strong	300 to 999 (RFU/pmole/min)
Moderate	100 to 299 (RFU/pmole/min)
Weak	30 to 99 (RFU/pmole/min)

Under the conditions utilized for this experiment, the assay is Strong