

# AQT1030 - VRK2 Assay Validation

PhosphoSens®-Kinetic Assay Format

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# **Outline for this Study**



PhosphoSens-Kinetic Assay Validation

### **Enzyme Source, Construct, and Lot Information:**

Invitrogen VRK2 (Cat#/Lot#: A30986/248990) amino acids 1-375, N-terminal GST tag

### **Reference Compound Information:**

Staurosporine MedChemExpress (Cat#/Lot#: HY-15141/125391)

### **Experiments to be run:**

**Enzyme Titration** 

Sensor Peptide K<sub>m</sub> Determination

ATP K<sub>m</sub> Determination

**DMSO Tolerance Test** 

Reference Compound IC<sub>50</sub> Determination at ATPK<sub>m</sub>

# **Enzyme Titration**

## **Reaction Conditions and Set Up**

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5 1mM ATP 1.2 mM DTT 0.012% Brij-35 1% glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl<sub>2</sub> 15 μM AQT1030

0, 0.08, 0.16, 0.31, 0.63, 1.3, 2.5, 5, 10, 20, 40, and 80 nM VRK2

# Reaction Set Up:2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5)

1x EDB or Kinase dilutions (5x in EDB) Final reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 or 25 µL final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 µL final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

#### Notes:

20 or 25 µL





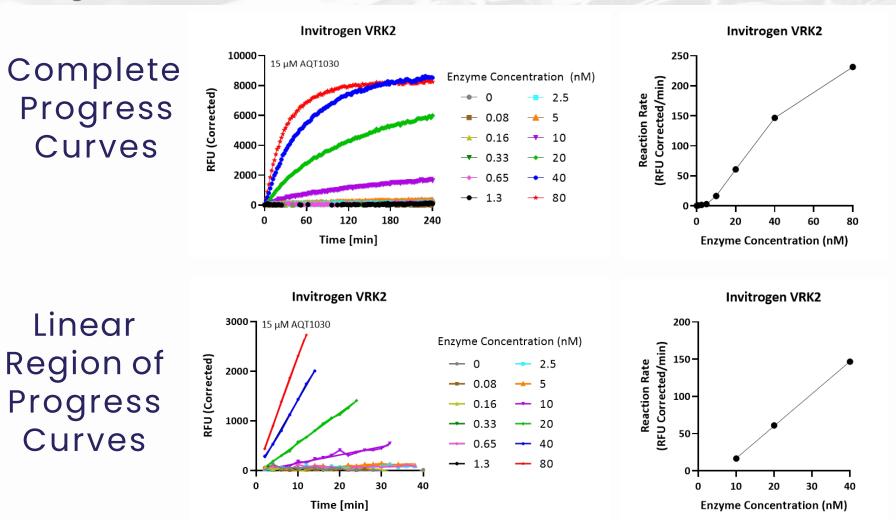
# **Enzyme Titration**



Linear

Range

## **Progress Curves**



# **Enzyme Titration**



## **Reaction Rate Table**

Enzyme Conc. (nM)	Reaction Rate (RFU/min)	Normalized Reaction Rate (RFU/pmole/min)		
0.08	0.39	241		
0.16	-0.71	-222		
0.33	0.48	72		
0.65	0.39	30		
1.3	-2.6	-99		
2.5	1.6	32		
5	4.1	41		
10	21	103		
20	61	153		
40	147	184		
80	232	145		

## The reaction is linear from 10-40 nM

# Sensor Peptide K<sub>m</sub> Determination



**Reaction Conditions and Set Up** 

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5 1mM ATP 1.2 mM DTT 0.012% Brij-35 1%glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl<sub>2</sub> 0, 0.6, 1.2, 2.3, 4.7, 9.4, 18.8, 37.5, 75, 150, 300, and 600 μM AQT1030 20 nM VRK2

# Reaction Set Up:<br/>2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

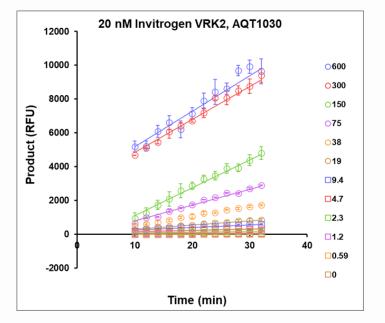
Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

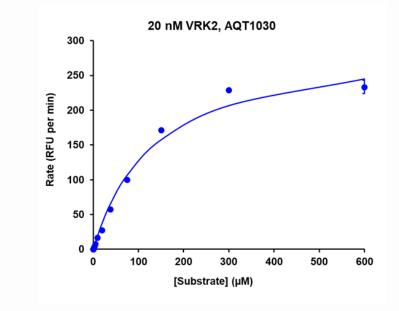
#### Notes:

# **Sensor Peptide K**<sub>m</sub> Determination

Titration Curves and K<sub>m</sub> Plot and Table

Sensor Peptide Titration Curves





Sensor Peptide

K<sub>m</sub> Plot

Sensor Peptide K<sub>m</sub> Table

Parameter	Value	Approx SE
Vmax (RFU per min)	301.0	7.6
Vmax (RFU per pmol per min)	752	19
Km (μM)	136.6	7.4
R squared	0.983	

## Sensor Peptide K<sub>m</sub> is 137 µM



# ATP K<sub>m</sub> Determination

## **Reaction Conditions and Set Up**

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5
0, 2.0, 3.9, 7.8, 16, 31, 63, 125, 250, 500, 1000, and 2000 $\mu M$ ATP
1.2 mM DTT
0.012% Brij-35
1%glycerol
0.2 mg/ml BSA
0.55 mM EGTA
10 mM MgCl <sub>2</sub>
15 μM AQT1030
20 nM VRK2



# Reaction Set Up:<br/>2 or 2.5 μL10x Sensor Peptide14 or 17.5 μLReaction Mix with ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

#### Notes:

# **ATP K<sub>m</sub> Determination**

Titration Curves and K<sub>m</sub> Plot and Table

**2000** 

01000

**○500** 

250

0130

063

□7.8 □3.9

2.0

ATP Titration Curves

20 nM VRK2, 15 µM AQT1030

2000

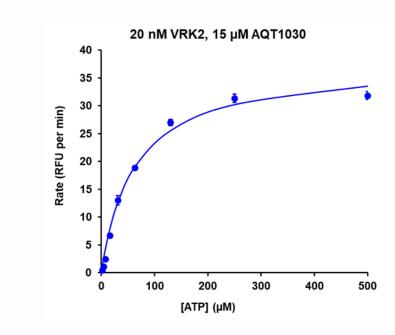
1500

1000

500

-500

Product (RFU)



ATP K<sub>m</sub> Plot

ATP K<sub>m</sub> Table

Parameter	Value	Approx SE
Vmax (RFU per min)	37.7	1.5
max (RFU per pmol per min)	94	4
Km (μM)	62.1	9.7
R squared	0.992	

## ATP $K_m$ is 62 $\mu M$

Time (min)





# **DMSO Tolerance Test**



## **Reaction Conditions and Set Up**

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5 1 mM ATP 1.2 mM DTT 0.012% Brij-35 1% glycerol 0.2 mg/ml BSA 0.55 mM EGTA 10 mM MgCl<sub>2</sub> 0, .01, .02, .04, .08, .16, .31, .63, 1.3, 2.5, 5.0, and 10% DMSO 15 μM AQT1030 20 nM VRK2

# Reaction Set Up:2 or 2.5 μL10x DMSO dilutions14 or 17.5 μLReaction Mix with Sensor Peptide, ATP & DTT4 or 5 μL1x EDB or Kinase dilutions (5x in EDB)20 or 25 μLFinal reaction volume

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 25  $\mu$ L final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20  $\mu$ L final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

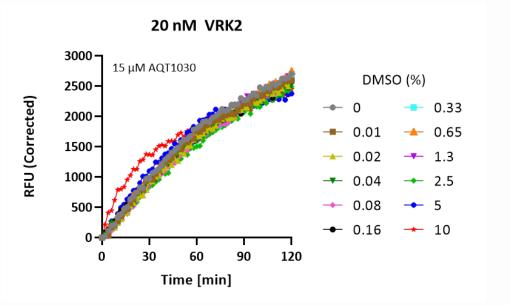
#### Notes:

# **DMSO Tolerance Test**

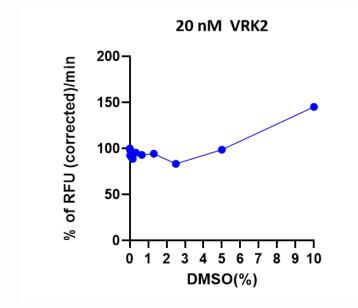


**Titration Curves and Inhibition Plot** 

## Complete Progress Curves



## Reaction Rate vs [DMSO] Plot



No change in enzyme activity out to 2% DMSO, but significant activation (150%) at 5% and 10% DMSO. This has been seen with other kinases.

# IC<sub>50</sub> Determination

## **Reaction Conditions and Set Up**

#### **Reaction Conditions:**

54 mM HEPES, pH 7.5 ATP at K<sub>m</sub> 1.2 mM DTT 0.012% Brij-35 1%glycerol 0.2 mg/ml BSA 0.55 mM EGTA  $10 \,\mathrm{mM} \,\mathrm{MgCl}_2$ 2% DMSO

15 µM AQT1030

20 nM VRK2

0, 0.169, 0.508, 1.52, 4.57, 13.7, 41.2, 123, 370, 1110, 3330, 10000 nM Staurosporine

## **Reaction Set Up:**

- 16 µL Reaction Mix with Sensor Peptide and Inhibitor 4 μL
- 1x EDB or Kinase dilutions (5x in EDB)
- Final reaction volume 20 µL

Reactions were run at 30°C for 240 minutes in either Corning, low volume 384-well, white flat round bottom polystyrene NBS microplates (Cat. #3824) at 20 µL final well volume or in in PerkinElmer, ProxiPlate-384 Plus, white shallow well microplates (Cat. #6008280) at 20 µL final well volume after sealing using optically-clear adhesive film (TopSealA-Plus plate seal, PerkinElmer [Cat. #6050185]) in a Biotek Synergy Neo 2 microplate reader with excitation (360 nm) and emission (485 nm) wavelengths.

Inhibitors are added via direct (0.4 µL of 50X stock in 100% DMSO) or intermediate dilutions (2.0 µL of 10X stock in 10% DMSO).

#### Notes:



# IC<sub>50</sub> Determination



Progress Curves and IC<sub>50</sub> Curves and Table

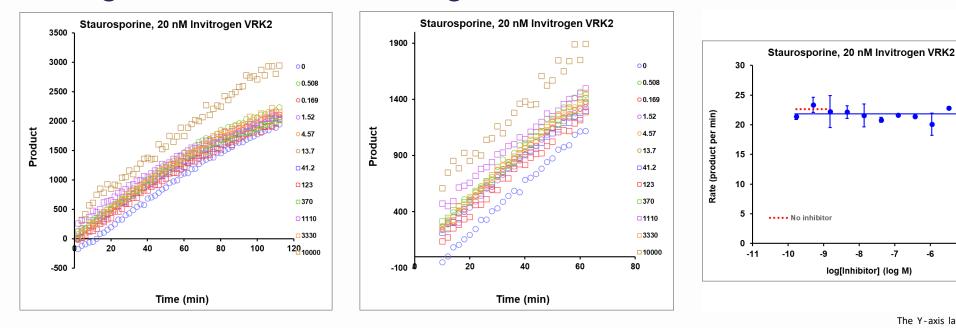
Inhibitor Titration **Progress Curves** 

## Linear Region of **Progress Curves**

IC<sub>50</sub> Curve

log[Inhibitor] (log M

IC<sub>50</sub> Table



Parameter	Value
Bottom	-0.1
Тор	21.9
log IC50	0.35
IC50 (nM)	> 10 µM
Ki (nM)	> 10 µM
Slope	-1.810
R squared	0.643
IC50 approx SE (nM)	ND
50% inhibition (nM)	ND

The Y-axis label is RFU/min.

Staurosporine IC<sub>50</sub> at ATP K<sub>m</sub> is > 10000 nM

# Summary



## Assay Validation Results and Progress Curve and Assay Strength at 1 mM ATP

Experiment	Result	Progress Curve		
Enzyme Titration Linear Range	10-40 nM	AQT1030		
Sensor Peptide K <sub>m</sub> Value	137 μM	10000 - 1000000 - 100000 - 100000 - 100000 - 100000 - 100000 - 100000 - 100000 - 100000 - 100000 - 100000 - 100000 - 100000 - 100000 - 100000 - 100000 - 100000 - 1000000 - 10000000 - 1000000 - 100000000		
ATP K <sub>m</sub> Value	62 μM	6000 - → 20 nM VRK2(Invitrogen)		
DMSOTolerance (highest % recommended)	2%			
Staurosporine $IC_{50}$ Determination at ATP $K_m$	> 10000 nM	20060 120 180 240 Time [min]		

Kinase Name Conc. (nM)	Conc. (nM) Su	Sox-based Substrate Name	Normalized	Normalized Rate	Assa	y Strength Key
			Reaction Rate	Stnd Error	Very Strong	> 1,000 (RFU/pmole/mir
					Strong	300 to 999 (RFU/pmole/m
		(RFU/pmole/min)	(RFU/pmole/min)	Moderate	100 to 299 (RFU/pmole/m	
VRK2	20	AQT1030	153	2.3	Weak	30 to 99 (RFU/pmole/mi

## Under the conditions utilized for this experiment, the assay is Moderate